Maternal-Fetal Calcium and Bone Metabolism During Pregnancy, Puerperium, and Lactation*

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- I. Introduction
- II. Maternal Physiology and Pathophysiology During Pregnancy
 - A. Maternal adaptive goals during pregnancy
 - B. Mineral ions and calcitropic hormones
 - C. Intestinal absorption of calcium
 - D. Renal handling of calcium
 - E. Skeletal calcium metabolism
 - F. Primary hyperparathyroidism
 - G. Hypoparathyroidism and pseudohypoparathyroidism
 - H. Summary
- III. Fetal-Placental Physiology and Pathophysiology
 - A. Fetal adaptive goals
 - B. Mineral ions and calcitropic hormones
 - C. Fetal-placental calcium transport
 - D. Renal handling of calcium and the amniotic fluid
 - E. Skeletal calcium metabolism
 - F. Fetal response to maternal hyper- or hypoparathyroidism
 - G. Integrated fetal calcium homeostasis
 - H. Summary
- IV. Maternal Physiology and Pathophysiology During Lactation
 - A. Maternal adaptive goals during lactation
 - B. Mineral ions and calcitropic hormones
 - C. Intestinal absorption of calcium
 - D. Renal handling of calcium
 - E. Skeletal calcium metabolism
 - F. Hypoparathyroidism and pseudohypoparathyroidism
 - G. Summary
- V. Neonatal Physiology and Pathophysiology
 - A. Neonatal adaptive goals
 - B. Mineral ions and calcitropic hormones
 - C. Intestinal absorption of calcium
 - D. Renal handling of calcium
 - E. Skeletal calcium metabolism

- F. Neonatal response to maternal hyper- or hypoparathyroidism
- G. Neonatal hypocalcemia
- H. Summary
- VI. Discussion and Conclusions

I. Introduction

IN THEIR classic text published in 1948, Albright and Reifenstein (1) noted the presentation of two young women with idiopathic osteoporosis worsened by pregnancy, and they recognized that significant maternal losses of skeletal calcium could occur during both pregnancy and lactation. They speculated that secondary hyperparathyroidism normally develops during pregnancy and lactation to resorb calcium from bone, and they concluded that, in certain cases, these skeletal calcium losses would cause a form of osteoporosis. Since that time, both pregnancy and lactation have been described in various endocrinology texts as states of "physiological, maternal hyperparathyroidism" (2, 3). However, this concept has not been supported by measurements of PTH with newer, more reliable assays.

Although Albright and Reifenstein's theory proved to be incorrect, it is now evident that mineralization of the fetal skeleton and continued skeletal growth in the infant both mandate a series of hormone-mediated adjustments in maternal calcium metabolism during pregnancy and lactation, respectively. These hormone-mediated adjustments normally satisfy the daily calcium needs of the fetus and infant without long-term consequences to the maternal skeleton. In addition, both fetal and neonatal calcium and bone metabolism are uniquely adapted to meet the specific needs of these developmental periods. The fetus must actively transport sufficient calcium across the placenta to meet the large demands of the rapidly mineralizing skeleton, whereas the neonate must quickly adjust to loss of placental calcium transport, while continuing to undergo rapid skeletal growth.

Here we review our present understanding of normal human calcium and bone metabolism during pregnancy, lactation, fetal development, and the neonatal period. We shall also discuss the relevant pathophysiology and management of clinical disorders of calcium and bone metabolism that can occur during these periods. Generally these conditions are due to preexisting disease (*e.g.*, hyperparathyroidism) that is compounded by the alterations in calcium

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and bone metabolism naturally occurring during these reproductive periods.

Although the focus of this review is on human calcium physiology and pathophysiology, the animal literature will be closely considered as well. Many of our models for explaining human physiology are based on these animal studies, particularly since ethical constraints generally prevent all but observational studies from being performed during human pregnancy and fetal development. Where both human and animal data are available, we will point out several significant differences that have been found between the animal and human data. These differences illustrate the difficulty of extrapolating from the animal models in the absence of human data.

The literature reviewed in this paper was obtained from computerized searches of the MEDLINE database, manual searches of *Index Medicus* before 1966, and the bibliographies of individual articles and texts.

II. Maternal Physiology and Pathophysiology During Pregnancy

A. Maternal adaptive goals during pregnancy

Measurements of calcium in ashed human abortuses determined that the normal total accumulation of calcium in a fetus at term is 21 g (range, 13–33 g) (4). Approximately 80% of this calcium accumulates during the third trimester, when the fetal skeleton is rapidly mineralizing (4, 5). Therefore, although maternal adaptations designed to meet the calcium needs of the fetus might begin early in pregnancy, they are most needed in the third trimester. Such adaptations could theoretically involve increased intestinal absorption of calcium, decreased renal excretion of calcium, and increased resorption of calcium from the maternal skeleton. The studies reviewed later in this section indicate that the major adaptive process in human and animal pregnancy is a 2-fold increases in the intestinal absorption of calcium, mediated by increases in 1,25-dihydroxyvitamin D and other mechanisms.

The pregnant rat has typically been used as a model for studying calcium metabolism during pregnancy, but the adaptive strategies of the rat differ importantly from those of the human (Table 1). These differences probably reflect the large litter size (six to 12 fetuses) and the short gestational

TABLE 1. Important differences between calcium physiology of human and rodent pregnancy $% \label{eq:table_state}$

Factor	Human pregnancy	Rat pregnancy
Blood ionized calcium	Stable	Reduced in late pregnancy
PTH	Low to low-normal from early pregnancy	Increased
1,25-D	Increased in early pregnancy	Increased in late pregnancy
Intestinal calcium absorption	Increased; follows rise in 1,25-D	Increased; precedes rise in 1,25-D

1,25-D, 1,25-Dihydroxyvitamin D.

period (22 days) of the rat; the rat must deliver 12 mg of calcium per fetus between day 17 of gestation and term (6).

B. Mineral ions and calcitropic hormones

The changes that occur in human maternal serum calcium, phosphate, and calcitropic hormone levels are schematically depicted in Fig. 1.

1. Calcium. Early studies of blood calcium levels during pregnancy in humans found a significant decrease in the total serum calcium as pregnancy progressed (7, 8). These early results seemed to confirm that the fetus was "draining" the maternal calcium and thereby creating a state of secondary hyperparathyroidism in the mother, as postulated by Albright and Reifenstein (1). The pregnancy-related fall in total serum calcium was later found to be the consequence of a fall in the serum albumin, and, thereby, the albumin-bound fraction of the total calcium (9). The intravascular fluid volume is greatly expanded during pregnancy, leading to the decreased serum albumin and hemodilution of pregnancy. Measurements of the ultrafiltrable fraction of serum calcium (representing complexed and free calcium) showed no significant change over prepregnancy values (10). More recent measurements of serum ionized calcium, using ion-specific electrodes, demonstrated that the mean ionized calcium level was maintained at the nonpregnant level throughout gestation in most cross-sectional (11-13) and longitudinal studies (14-20).

In contrast, the serum total and ionized calcium have been reported to fall during the last several days of pregnancy in the rat (21). Maternal losses of calcium to a litter of rapidly growing fetuses may exceed the maternal capacity to maintain a normal serum calcium level. Indeed, larger litter sizes correlated with lower serum calcium in pregnant rats (22). In white-tailed deer, the corrected serum calcium falls in the last 1 to 2 weeks of gestation (23). Pregnant ewes have a mild decrease in total serum calcium over the last 6 weeks of pregnancy, likely due to the fall in serum albumin (24); moreover, in one study, about 13% of Awassi fat-tail ewes were found to develop signs and biochemical evidence of hypocalcemia in the last month of pregnancy (25). Therefore, data from several animal models suggest that maternal blood calcium regulation may be disrupted by fetal demands in late pregnancy.

2. *Phosphate*. Serum phosphate levels are normal throughout pregnancy in humans and animals, as is the renal tubular reabsorption of phosphate (14, 17, 26–29).

3. *PTH*. The bulk of published human data on PTH levels in pregnancy was obtained from studies that used early-generation PTH RIAs (18, 26, 30–40); some of the more frequently cited studies reported high maternal serum levels of PTH in the latter half of pregnancy (18, 30–34, 39). These data must now be reinterpreted, because it is now known that these PTH RIAs were insensitive and heterogeneously measured multiple different fragments of PTH, most of which were biologically inactive (41, 42).

With the advent of sensitive two-site immunoradiometric (IRMA) PTH assays that accurately determine the level of



FIG. 1. Schematic illustration of the longitudinal changes in calcium, phosphate, and calcitropic hormone levels that occur during human pregnancy. Normal adult ranges are indicated by the *shaded areas*. Data have been compiled from the following sources: total calcium (9), ionized calcium (14–19), phosphate (14, 17, 26, 27), PTH (11, 14–16, 27, 44), 1,25-dihydroxyvitamin D (15, 41, 57–59), calcitonin (14, 34, 36, 37, 86–89), and PTHrP (44, 111). The progression in PTHrP levels has been depicted by a *dashed line* to reflect that the data are less complete.

intact PTH (42), PTH levels have been typically found to be low-normal in the serum of pregnant women in all three trimesters (11, 12, 17, 19, 20, 27, 43). Five prospective, longitudinal studies found that the mean PTH level was in the low-normal range (*i.e.*, <50% of the mean nonpregnant value) during the first trimester but increased steadily to the mid-normal range by the end of pregnancy (14–16, 27, 44). These findings have been independently validated by reports of normal nephrogenous cAMP levels (12, 26, 44) and low to normal PTH-like bioactivity (11) throughout human pregnancy (although this may be confounded by synthesis of nephrogenous cAMP due to the effects of PTHrP). Studies in primates suggest that the parathyroid glands may have less secretory reserve as pregnancy progresses; the incremental PTH response to acute EDTA-induced hypocalcemia in rhesus monkeys decreased across the trimesters (39).

In contrast to humans, rats develop secondary hyperparathyroidism late in pregnancy. Normally, in late pregnancy, both maternal levels of intact immunoreactive (45) and bioactive (46) PTH rise to exceed the normal range, and the maternal ionized and total calcium levels decline slightly (21). The parathyroid gland volume has also been reported to increase during normal rat pregnancy (47, 48). In vitro studies in pregnant rats indicate that the parathyroids secrete more PTH at a given extracellular calcium concentration, when compared with parathyroid cells taken from nonpregnant rats (49). The PTH levels begin to rise earlier in gestation, and peak at higher levels, in pregnant rats fed a modestly calcium-restricted diet (21, 50). This increase in PTH during late pregnancy is critical for normal maternal calcium homeostasis; parathyroidectomized pregnant rats can exhibit signs of tetany in the last 2-4 days of gestation and death during the birthing process (51-54). In parathyroidectomized pregnant rats, dietary intake and weight gain decline, while serum 1,25-dihydroxyvitamin D and intestinal calbindin_{9K}-D levels fall (52-54). Maternal tetany coincides with the time onset of rapid fetal accretion of calcium (6); therefore, the parathyroidectomized pregnant rat has compromised dietary intake and intestinal calcium absorption at the time of peak fetal demand for calcium. The calcium abnormalities can be completely prevented when the rats are fed a high-calcium, low-phosphorus diet. Taken together, these observations indicate that rats (but not humans) normally develop a form of secondary hyperparathyroidism during late pregnancy in response to the fall in the maternal serum calcium level. Rats may be more dependent on PTHmediated bone resorption and PTH-induced 1α -hydroxylase up-regulation during late pregnancy, at a time when the combined calcium need of a litter of fetuses is at its peak.

In summary, immunoreactive and bioactive PTH levels are in the low-normal range during early human pregnancy and are in the mid-normal range at term; in contrast, immunoreactive and bioactive PTH levels in rats are normal in early pregnancy but exceed the normal range in late gestation.

4. 1,25-Dihydroxyvitamin D. Cross-sectional studies have found that the serum level of 1,25-dihydroxyvitamin D more than doubles early in the first trimester in human pregnancy (12, 27, 36, 37, 55–58). Longitudinal studies have found that

the levels of both free and bound 1,25-dihydroxyvitamin D are doubled, and that this increase is maintained until term (15, 20, 41, 57-59). Although clearance of 1,25-dihydroxyvitamin D has not been studied during human pregnancy, in pregnant rats, sheep, and rabbits the increased 1,25-dihydroxyvitamin D levels were due to increased production, and not decreased metabolic clearance, of 1,25-dihydroxyvitamin D (60-63). In vitro measurements in homogenates of maternal kidney from rabbits and guinea pigs show that the renal 1α -hydroxylase may be up-regulated 2- to 5-fold (64, 65). The increase in the 1,25-dihydroxyvitamin D level begins while the PTH level is in the low-normal range in humans (Section II.B.3, above); this may indicate that PTH does not mediate the up-regulation of the maternal renal 1α -hydroxylase during early human pregnancy. Furthermore, parathyroidectomy in pregnant sheep reduces, but does not eliminate, the pregnancy-related increase in 1,25-dihydroxyvitamin D (66). Other potential direct or indirect regulators of the 1α hydroxylase include PTHrP (Section II.B.6, below), estradiol, PRL, and placental lactogen. Estradiol (67), PRL (68, 69), and placental lactogen (69) acutely stimulate the 1α -hydroxylase in vitro, and placental lactogen (but not PRL) raised the serum 1,25-dihydroxyvitamin D levels in hypophysectomized, nonpregnant rats (70). The effect of estradiol on the 1α -hydroxylase has been confirmed in vivo by the observation that estrogen replacement in postmenopausal women increases the free and total serum 1,25-dihydroxyvitamin D level (71). However, an effect of PRL in vivo has not been confirmed, since hyperprolactinemic patients showed no alteration in 1,25-dihydroxyvitamin D levels (72). Further, in pregnant women, the high 1,25-dihydroxyvitamin D levels of pregnancy did not correlate with serum PRL, estrogens, or human placental lactogen (73).

In addition to the renal 1α -hydroxylase, 1α -hydroxylase activity found in maternal decidua, placenta, and fetal kidneys may also add 1,25-dihydroxyvitamin D to the maternal circulation during pregnancy (59, 74-77). To test this hypothesis, [3H]25-hydroxyvitamin D was administered to pregnant rats after bilateral maternal nephrectomy (74, 78). Newly synthesized (i.e., tritiated) 1,25-dihydroxyvitamin D appeared in the maternal circulation of nephrectomized pregnant rats (but not in nonpregnant nephrectomized rats). Although this study indicates that extrarenally produced 1,25-dihydroxyvitamin D can reach the maternal circulation (74), the specific extrarenal sites and the amounts of 1,25dihydroxyvitamin D produced could not be ascertained. Data from the Hannover pig model (autosomal recessive 1α -hydroxylase deficiency) indicate that the amounts contributed by these extrarenal sites may be insignificant. In pregnant sows homozygous for absence of 1α -hydroxylase activity, serum levels of 1,25-dihydroxyvitamin D were very low, comparable to the nonpregnant values (79). The presence of heterozygous fetuses did not increase the circulating level of 1,25-dihydroxyvitamin D in the homozygous sows (79). The same gene controls renal and decidual 1α -hydroxylase activity in this model (77). A single case report of a human patient on chronic hemodialysis found 1,25-dihydroxyvitamin D levels of 10–15 pg/ml during pregnancy; these levels were higher than the nonpregnant level in the same patient, but were far lower than in normal pregnancy (80). It is, therefore, likely that increased maternal production of 1,25-dihydroxyvitamin D is mainly due to increased activity of maternal, renal 1α -hydroxylase and not to large contributions from extrarenal sites.

Again, the pregnant rat model differs somewhat from the human, in that the maternal rise in 1,25-dihydroxyvitamin D level does not occur in rats until the time of fetal skeletal mineralization in late gestation (22, 45, 45, 81, 82), at which time the serum PTH levels rise above normal (22, 45) and serum ionized calcium levels fall (21, 22). Larger litter sizes correlate with higher maternal 1,25-dihydroxyvitamin D levels (22). These studies suggest that the effect of PTH on the renal 1 α -hydroxylase may dominate the production of 1,25-dihydroxyvitamin D during late pregnancy in the rat.

Serum 25-hydroxyvitamin D levels are unchanged in human pregnancy, and 24,25-dihydroxyvitamin D levels are lower in pregnant women than in controls (35). Supplementation with 1000 IU of vitamin D_3 daily after the first trimester in humans did not affect maternal calcium, phosphate, PTH, and 1,25-dihydroxyvitamin D levels; this suggests that the changes in calcitropic hormone levels observed in human pregnancy are not the result of occult vitamin D deficiency (83). Maternal vitamin D deficiency in the rat has been associated with reduced fertility and smaller litter sizes, and up to 20% of pregnant, vitamin D-deficient rats may die of hypocalcemia near term (84, 85).

In summary, free and total 1,25-dihydroxyvitamin D levels rise early in human pregnancy to peak at twice the normal range, while in rats the 1,25-dihydroxyvitamin D level does not rise until late gestation. These increases appear to be due to increased production of 1,25-dihydroxyvitamin D by the maternal kidneys, with possibly small contributions from maternal decidua, placenta, and fetal kidneys. PTH may be less important during pregnancy in humans compared with rats in mediating this rise in 1α -hydroxylase activity.

5. Calcitonin. Serum calcitonin levels in human pregnancy have generally been reported to be higher than nonpregnant values, with at least 20% of values exceeding the normal range (14, 34, 36, 37, 86–89). Several human studies reported that calcitonin levels were not elevated in pregnancy (15, 18, 33, 35); however, these studies were flawed by the use of improper controls. For example, in some of these studies, postpartum measurements in the same women were used as the baseline, and it has since been shown that calcitonin is also elevated in the postpartum period (see Section IV.B.5, below). Similar data from monkeys (39), sheep (77, 90, 91), deer (23), goats (77), and rats (92) have confirmed that the maternal calcitonin level is elevated during pregnancy. No clearance data are available for humans or other animals, but the increased level of calcitonin is generally thought to reflect increased synthesis.

Thyroidal C cells, breast, and placenta are sites of calcitonin synthesis during pregnancy (93, 94). It is not surprising, therefore, that a rise in calcitonin is found in totally thyroidectomized women, most likely due to calcitonin synthesized by the placenta and breast (93, 94). In pregnant rhesus monkeys, acute calcium infusions led to a progressively greater calcitonin response across the trimesters, which may indicate greater secretory reserve of the thyroidal C cells and placenta (39).

It has been speculated that elevated calcitonin protects the maternal skeleton from excessive resorption of calcium, a hypothesis that has been difficult to prove. Indeed, the physiological role of calcitonin in human calcium and skeletal metabolism has not been established (95). No adequate model of experimental calcitonin deficiency has been created, partly because the extrathyroidal sites of calcitonin synthesis were not appreciated at the time. All models used total thyroidectomy with parathyroid gland autotransplantation and thyroid hormone replacement in pregnant goats or rats (53, 91, 96–98). In none of these models was the serum calcitonin or TSH measured to determine whether a calcitonin-deficient, euthyroid state had been attained. Thus, although these models suggested that an intact thyroid gland protected the maternal skeleton from loss of bone mineral during pregnancy, these findings remain to be confirmed by more rigorously controlled models.

In summary, calcitonin levels are increased during pregnancy in humans and animals, partly due to extrathyroidal synthesis in the placenta and breast. The possible role of calcitonin in protecting the maternal skeleton from increased resorption during pregnancy needs more study.

6. PTH-related protein (PTHrP). PTHrP was originally identified in 1987 as the cause of humoral hypercalcemia of malignancy (99). PTHrP has been postulated to be a prohormone, which is processed into several different circulating fragments or hormones, each of which, in turn, may have different functional roles and specific receptors (100). PTHrP has partial homology in its first 13 amino acids to PTH (101-103) and activates the common PTH/PTHrP receptor (42). Amino-terminal forms of PTHrP (PTHrP 1-34, 1-86, or 1-141) resemble PTH in their actions on kidney and bone (104) and can inhibit acetylcholine-induced uterine contractions in the rat (105). Levels of PTHrP decreased acutely in the amnion and myometrium at the time of onset of labor in humans (106). It has been suggested that amino-terminal forms of PTHrP may, therefore, have a role in regulating the onset of labor (106). A midmolecular form of PTHrP stimulates placental calcium transport in the fetus (Section III.C, below), although its possible role in the mother is unclear. The carboxyl-terminal portion of PTHrP, termed "osteostatin," is able to inhibit osteoclastic bone resorption in some in vitro assays (107, 108) and in rats in vivo (109); therefore, this fragment of PTHrP could have a role in protecting the maternal skeleton during pregnancy.

The development of RIAs for PTHrP has concentrated on detecting the PTH-like amino-terminal fragments of PTHrP and has thus far largely ignored the detection of other fragments that might be biologically active. Therefore, no data are available on the levels of midmolecular or carboxylterminal fragments of PTHrP during pregnancy compared with controls. An early RIA that used an antibody to PTHrP 1-34 found no elevation of PTHrP in pregnancy (110). Newer, more sensitive two-site immunoradiometric assays that measure forms of PTHrP that encompass amino acids 1 through 86 have found a significant increase in the maternal PTHrP level, beginning as early as weeks 3 to 13 of human pregnancy (44, 111). This increase is not due to any change in the clearance of PTHrP 1-34, 1-86, or 1-141 during pregnancy, as determined in sheep (112, 113). The increase in amino-terminal PTHrP, by activating the PTH/PTHrP receptor in kidney and bone, may well explain (at least in part) the increase in 1,25-dihydroxyvitamin D and ionized calcium, and the decrease in PTH levels, found during human pregnancy.

Vol. 18, No. 6

The source of PTHrP in the maternal circulation during pregnancy is not established, but several candidate sites are known. PTHrP is produced by the placenta (114), amnion (106), decidua (106), umbilical cord (115), and fetal parathyroid glands (116) and potentially might reach the maternal circulation. PTHrP produced by the breast tissue is detectable in human colostrum (117), and it is produced as early as day 14 of pregnancy by the mammary glands of the rat (118).

Overproduction of PTHrP by the breast might explain the development of hypercalcemia at 24 weeks of gestation in a woman with massive (4.5 kg) mammary hyperplasia of pregnancy, associated with hypercalciuria, hypophosphatemia, and undetectable PTH levels (119). Bilateral mastectomies in the second trimester of that same pregnancy corrected the hypercalcemia and the suppressed PTH level (119).

In summary, PTHrP may be made available to the maternal circulation by several different maternal and fetal sources. PTHrP fragments encompassing amino acids 1-86 are increased in the maternal circulation during pregnancy and may contribute to the elevations in 1,25-dihydroxyvitamin D and blood calcium, and suppression of PTH, noted during pregnancy. The true quantitative importance of PTHrP in maternal physiology needs to be established.

7. Other hormones. Pregnancy induces a dramatic rise in other hormones, including the sex steroids, PRL, and placental lactogen. The possibility that each of these, in turn, may have direct or indirect effects on calcium and bone metabolism during pregnancy has been largely unexplored. There is some evidence to suggest that PRL and placental lactogen may increase the intestinal transport of calcium (70, 120, 121), reduce urinary calcium excretion (122, 123), and stimulate synthesis of PTHrP (124) and 1,25-dihydroxyvitamin D (68, 69). This is discussed in more detail in the relevant sections.

C. Intestinal absorption of calcium

Calcium is absorbed throughout the small intestine, a small portion by active transport in the duodenum and proximal jejunum, and the major portion by passive mechanisms in the distal jejunum and ileum (125). Mineral balance and calcium kinetic studies in humans using stable isotopes of calcium (⁴⁸Ca, ⁴⁴Ca, ⁴²Ca) have consistently found a positive calcium balance and a doubling of the intestinal absorption of calcium during human pregnancy from as early as 12 weeks of gestation (the earliest time point studied) (27, 126, 127). By studying the effect of an oral calcium load on serum calcium and urine calcium excretion, other investigators indirectly confirmed that intestinal calcium absorption must be increased in all trimesters (12, 128). The results of these studies led to speculation that the increase was mediated by 1,25-dihydroxyvitamin D, and this appeared to be confirmed when elevated levels of 1,25-dihydroxyvitamin D were found during human pregnancy (Section II.B.4, above). 1,25Dihydroxyvitamin D probably stimulates intestinal calcium absorption by increasing the synthesis of proteins, including the intestinal vitamin D-dependent calcium-binding protein, calbindin_{9K}-D. Protein and mRNA levels of calbindin_{9K}-D increase in the intestines of mice and rats during pregnancy and plateau when both maternal 1,25-dihydroxyvitamin D levels and the efficiency of intestinal calcium absorption are at peak levels (129–131). Maternal vitamin D deficiency in rodents reduces the rise in the intestinal expression of calbindin_{9K}-D (132, 133), while 1,25-dihydroxyvitamin D administration can restore it (133).

The rise in intestinal absorption of calcium occurs by midpregnancy in rats, before the onset of rapid skeletal mineralization in the fetus (45). The doubling of intestinal absorption persists in parathyroidectomized rats (134) and may, therefore, be independent of PTH regulation. The early increase in intestinal calcium absorption may allow the pregnant mother to accrete calcium (probably in the maternal skeleton), before the peak fetal demand for calcium in late pregnancy. Consistent with this hypothesis, it has been estimated from isotope studies in the pregnant rat that 92% of fetal skeletal calcium content was absorbed from the maternal diet at some point during pregnancy (135). Further, several investigators have found that pregnant rats normally store calcium during the first half of pregnancy (136), such that by the end of pregnancy, the calcium content of the femurs is unchanged (137). Inadequate accretion of calcium early in pregnancy may lead to a net loss of maternal skeletal calcium later in pregnancy. For example, under dietary calcium restriction, pregnant rats (138, 139) and goats (98) have reduced calcium content in their long bones by the end of gestation. Similarly, maternal vitamin D deficiency has been found to cause maternal skeletal demineralization by the end of pregnancy (140).

PRL treatment of pregnant, vitamin D-deficient rats resulted in an increase in the intestinal absorption of calcium; PRL might, therefore, have an effect on the intestine independent of 1,25-dihydroxyvitamin D (120). This is further supported by studies in everted gut sacs of nonpregnant, hypophysectomized rats, where PRL and placental lactogen stimulated the intestinal transport of calcium (70, 121). Also in rats, the increase in duodenal calcium absorption has been found to precede the rise in the 1,25-dihydroxyvitamin D level by 1 week, suggesting that the intestinal effect is not dependent solely on vitamin D (45, 141). Even in the absence of vitamin D, pregnancy in rats is associated with hypertrophy of the small intestine and a doubling of intestinal absorption of calcium (141, 142). Furthermore, rats hypocalcemic from vitamin D deficiency developed a progressive rise in serum calcium levels during pregnancy, despite unchanged serum PTH levels (143). However, an independent effect of PRL on intestinal calcium absorption could not be demonstrated in studies on humans. Hyperprolactinemic patients showed no alteration in the intestinal absorption of calcium (72).

In summary, intestinal calcium absorption is increased 2-fold early in human and rat pregnancy, probably through a 1,25-dihydroxyvitamin D-mediated increase in intestinal calbindin_{9K}-D and other proteins. PRL and placental lactogen (or possibly other factors) may mediate part of the nor-

mal pregnancy-related increase in intestinal calcium absorption. The early rise in intestinal calcium absorption may allow the maternal skeleton to store calcium in advance of the peak fetal demands later in pregnancy. The increased intestinal calcium absorption appears to be a major maternal adaptation to meet the fetal need for calcium.

D. Renal handling of calcium

Pregnancy is associated with an increase in creatinine clearance and glomerular filtration rate (144, 145). The 24-h urine calcium excretion is increased as early as the 12th week of gestation (the earliest time point studied), and averages 300 ± 61 mg in the third trimester (levels in the hypercalciuric range are not uncommon) (12, 14, 20, 27, 146, 147). Since fasting urine calcium values are normal or low, the increase in 24-h urine calcium reflects the increased intestinal absorption of calcium (absorptive hypercalciuria) (12, 28, 44). A similar 2-fold increase in urinary calcium excretion has been observed in the pregnant rat from the second week of gestation (148). Although PRL and placental lactogen have been shown to reduce urinary calcium excretion in nonpregnant rabbits in vivo (122, 123), the effect (if any) of either hormone on the kidneys of pregnant humans and rats must be very modest.

Interestingly, preeclampsia and pregnancy-induced hypertension (PIH) have been associated with hypocalciuria (147, 149–152). Further studies have found the hypocalciuria to be associated with low 1,25-dihydroxyvitamin D levels (149–152), but to be independent of PTH, calcitonin, or ionized calcium levels (147, 149–151). The finding of hypocalciuria prompted a large trial of calcium supplementation in pregnant women, which recently reported no benefit in preventing preeclampsia or PIH (153). These abnormalities in 1,25-dihydroxyvitamin D and urine calcium excretion are, therefore, probably secondary to a primary renal tubular defect occurring in preeclampsia and PIH and are likely not the primary cause of the hypertension (149).

E. Skeletal calcium metabolism

1. Bone formation and resorption. Histomorphometric parameters of both bone formation and osteoclast-mediated resorption are increased during pregnancy in rats (154). Pregnant beagle dogs also show histomorphometric evidence of increased bone turnover in iliac trabecular bone (155). Despite evidence of increased turnover, bone mineral content during pregnancy in rats does not change (137, 140, 154, 156). In contrast, pregnant ewes have a 20% decrease in skeletal calcium content during gestation (157).

Comparable histomorphometric data are not available for human pregnancy, but markers of bone formation and resorption have been assessed. Generally speaking, such indices are more reliable for measuring changes in bone resorption than bone formation (158, 159). Several markers of bone resorption (tartrate-resistant acid phosphatase, deoxypyridinoline/creatinine, pyridinoline/creatinine, and hydroxyproline/creatinine) are low in the first trimester but rise steadily to peak at values up to twice normal in the last trimester (27, 44, 160, 161). In contrast, osteocalcin, a marker of bone formation, is low or undetectable early in gestation and sometimes rises to normal levels by term (15, 27, 161– 163). Other markers of bone formation (procollagen I carboxypeptides, bone-specific alkaline phosphatase) are low in the first trimester and have been found to remain low (44) or rise to normal or above in the last trimester (27, 160). Total alkaline phosphatase rises early in pregnancy due to contributions from the placental fraction, and, therefore, is not a useful marker of bone formation in pregnancy (14, 44).

Taken together, the histomorphometric data from animals, and the changes in the markers of bone formation and resorption in humans, indicate that bone turnover is probably low in the first half of pregnancy, but may be increased in the third trimester. The third trimester increase in bone turnover corresponds to time of the peak rate of calcium transfer to the fetus and may result from mobilization of skeletal calcium stores (which contain 99% of the body's stores of calcium) to help supply the fetus.

2. Bone density. Concerns about fetal radiation exposure have resulted in few studies of changes in maternal bone mass during pregnancy; these studies used techniques that are far less precise or reproducible than the current standard, dual x-ray absorptiometry (DXA) (164, 165). Of the scant data available, an early study used x-ray spectrophotometry of the radius and femur to demonstrate a progressive decrease in trabecular bone density during pregnancy (166). Using more modern techniques, four prospective studies of bone density during pregnancy did not find a significant change in cortical or trabecular bone density, as respectively determined by single photon absorptiometry (SPA) and/or dual-photon absorptiometry (DPA) (28, 146, 167, 168). Another study found a significant decrease in bone mineral density of the femoral neck and radial shaft, but no change in lumbar bone density, by comparing preconception SPA and DPA measurements to those taken 6 weeks postpartum (169). Most recently, crosssectional (170) and longitudinal studies (161, 171) have found a progressive decrease during pregnancy in indices thought to correlate with bone mineral density, as determined by ultrasonographic measurements of the os calcis in all three trimesters.

The majority of retrospective, epidemiological studies of pre- and postmenopausal women have found no association of parity with bone density or fracture risk (172-191). In contrast, several other studies found increased parity to be beneficial, as indicated by a slightly greater lumbar (192, 193), femoral (194), or radial bone density (194-196) and decreased hip fracture risk (197, 198). Four remaining studies linked parity to somewhat decreased lumbar bone density (199–201) or increased hip fracture risk (202). An epidemiological study of healthy women aged 21 to 95 found divergent effects of parity at different anatomical sites. Femoral neck bone mineral density was decreased in parous women by 1.5% per live birth, while lumbar spine bone density was not influenced by parity (203). Among the studies that found no significant association of parity, several reported that a first pregnancy as an adolescent was associated with decreased bone density (178, 190, 195), possibly because the fetal calcium demands of pregnancy reduce the peak bone mass that is eventually achieved in the adolescent. Overall,

many of these epidemiological studies had significant methodological limitations, specifically the difficult problem of retrospectively separating out the effects of parity from those of lactation. Nevertheless, it may be reasonable to conclude from these studies that if parity has either a positive or a negative effect on bone density or fracture risk, it must be only a very modest effect.

Therefore, although changes in serum and urine markers of bone formation and resorption have indicated that bone turnover may be increased in the third trimester, it is impossible to determine from the available bone density data whether there is any acute change in bone mineral during human pregnancy. Further, it is also unknown whether any such acute change has a long-term effect on the calcium content or fragility of the maternal skeleton.

3. Osteoporosis in pregnancy. The rare presentation of idiopathic osteoporosis in a woman of child-bearing age has often been associated with a recent pregnancy, as noted by Albright and Reifenstein (1) and other early case reports (204–206). The exact prevalence of the condition is uncertain. The theory that pregnancy might cause osteoporosis (as proposed by Albright and Reifenstein) was disputed by an early observational study of five women with symptomatic, severe osteoporosis presenting in a first pregnancy (207). In subsequent pregnancies, these women were found to have no worsening of their condition, but the parameters used (new pain or fracture, worsening of osteopenia on plain roentgenograms) were crude and insensitive by methods available today (207). Despite better documentation of the absence of known causes of decreased bone density in more recent case reports (208-212), it has not been possible to exclude the possibility that low peak bone mass and/or an accelerated bone resorptive state preceded the pregnancy and simply became clinically obvious in pregnancy. In addition, some reported cases of osteoporosis in pregnancy have been clearly confounded by the presence of other recognizable causes of secondary osteoporosis, such as chronic heparin, anticonvulsant, or corticosteroid therapy (209, 212). In two documented cases of osteoporosis diagnosed in pregnancy, the female progeny were found at age 10 to also have low bone mineral density (213). This finding suggested that a shared genetic or environmental factor (and not pregnancy) was the cause of osteoporosis in the mothers and daughters. The limited data from bone biopsy typically show no evidence of osteomalacia, but only mild osteoporosis or normal architecture (208, 212). It remains intriguing to speculate that some of these rare cases of osteoporosis presenting in pregnancy may result from excessive resorption of calcium from the maternal skeleton, perhaps in the setting of inadequate intake of calcium, low stores of 25-hydroxyvitamin D, or an excessive rise in PTHrP in the maternal circulation (see also the discussion of osteoporosis in lactation, Section IV.E.3, below). Nevertheless, these rare cases may simply represent idiopathic osteoporosis occurring in pregnant women by mere chance.

A second (also rare) form of pregnancy-associated osteoporosis is a focal, transient osteoporosis of the hip (214–217). Typically these patients present with unilateral or bilateral hip pain, limp, and/or hip fracture in the third trimester (214, 216, 218, 219). Radiolucency of the femoral head and neck was recognized on plain radiographs taken in early reports of this condition (215, 220); and more recently, DXA measurements have shown that the bone density of the symptomatic femoral head and neck is reduced (218). Magnetic resonance imaging (MRI) of the affected femoral head in one patient showed a joint effusion and images suggesting increased water content of the femoral head and marrow cavity (221). Routine serum chemistries are typically normal (222). Alkaline phosphatase and urine hydroxyproline have been reported to be elevated (215, 218, 223); however, the interpretation of these findings is uncertain, since control measurements from normal pregnant women were not compared. Intriguingly, the decreased bone mineral density of the femoral head and neck typically resolves within 2 to 6 months postpartum (214, 218, 219), including the MRI findings (221). Patients generally require only pain relief and continued mobilization for this self-limited condition. The fact that this rare condition is typically localized to one or both femoral heads, and not the rest of the skeleton, suggests that it is not the result of a generalized increase in skeletal resorption. Several theories have been proposed to explain this condition, including femoral venous stasis due to pressure from the pregnant uterus, a form of Sudeck's atrophy, or reflex sympathetic dystrophy (causalgia), ischemia, trauma, viral infections, marrow hypertrophy, immobilization, and fetal pressure on the obturator nerve (214-216, 220). As yet, the etiology of transient osteoporosis of the hip in pregnancy remains unclear; its association with pregnancy may be not causal but incidental. In any case, it appears likely that this disorder is not a manifestation of altered calcitropic hormone levels or mineral balance during pregnancy.

F. Primary hyperparathyroidism

The presentation of primary hyperparathyroidism in pregnancy raises important diagnostic and management considerations. Many cases are asymptomatic, detected by routine prenatal biochemical tests or after the presentation of hypocalcemia in the neonate. Several normal pregnancy-related changes in calcium and PTH physiology (noted above) may obscure the diagnosis of mild primary hyperparathyroidism. These include the fall in total serum calcium, the rise in the corrected serum calcium (111, 224), the fall in the intact PTH level (14–16, 44), and the rise in the 24-h urinary excretion of calcium, often into the hypercalciuric range (14, 27) (see also *Sections II.B and II.D*, above).

Although maternal primary hyperparathyroidism in pregnancy is probably a rare condition (there are no data available on its prevalence), it has been associated in the literature with an alarming rate of adverse outcomes in the fetus, including a 30% rate of spontaneous abortion or stillbirth (225, 226). In the neonatal period, a 50% rate of tetany and a 25% rate of neonatal death has been reported (225, 227). These adverse outcomes are thought to result from suppression of the fetal parathyroid glands; this suppression may occasionally be prolonged for months (228, 229). PTH cannot cross the placenta (230–232); therefore, the fetal parathyroid suppression is thought to result from increased net calcium flux across the placenta to the fetus, facilitated by maternal hypercalcemia. Evidence from animal models has confirmed that acute elevations in maternal serum calcium cause an increase in fetal serum calcium, and a fall in fetal PTH level (233). However, whether chronic maternal hypercalcemia has the same effect on fetal serum calcium, or placental calcium transport, has not been determined.

Surgical correction of primary hyperparathyroidism during the second trimester, to prevent fetal and neonatal complications, has been almost universally recommended (226, 234–236). Several case series have found elective surgery to be well tolerated and to dramatically reduce the rate of adverse events when compared with the earlier cases reported in the literature (234, 235, 237, 238). However, many of the women in those early cases were symptomatic and had nephrocalcinosis or renal insufficiency. Those early case reports may also have reflected reporting bias of adverse fetal and neonatal outcomes. Whether the milder, asymptomatic form of primary hyperparathyroidism commonly seen today has the same risk of adverse fetal or neonatal outcomes has not been determined. In several case reports, mild elevations in maternal serum calcium were followed without operative intervention, and no adverse fetal or neonatal outcome occurred (239, 240). However, in other cases the mild hypercalcemia of both asymptomatic primary hyperparathyroidism and familial hypocalciuric hypercalcemia has been reported to cause neonatal parathyroid suppression and tetany (241-243). Nevertheless, it is probably reasonable to follow cases of asymptomatic primary hyperparathyroidism with mild hypercalcemia conservatively and to reserve surgery in the second trimester for patients that are symptomatic or have more severe hypercalcemia. If surgery is deferred, the neonate must be monitored closely for the development of hypocalcemia.

G. Hypoparathyroidism and pseudohypoparathyroidism

As described earlier (*Section II.B.4*), free and bound 1,25dihydroxyvitamin D levels normally double during human pregnancy in the presence of low-to-normal intact PTH levels, and, therefore, it is likely that PTH does not mediate the pregnancy-related rise in 1,25-dihydroxyvitamin D production. Other hormones of pregnancy, such as estrogen, PTHrP, and perhaps placental lactogen and PRL, may regulate the increased production of 1,25-dihydroxyvitamin D by maternal kidney and decidua. Also, placenta and fetus may contribute to the maternal increase in 1,25-dihydroxyvitamin D.

In multiple case reports, pregnant hypoparathyroid women have been found to have fewer hypocalcemic symptoms, a rise in the serum calcium, and decreased dependence on supplemental calcitriol to maintain a normal serum calcium (244–252). This finding is consistent with a limited role for PTH in the pregnant woman and suggests that an increase in 1,25-dihydroxyvitamin D and/or increased intestinal calcium absorption will occur in the absence of PTH. The literature on hypoparathyroidism in pregnancy is not entirely consistent on this point, since in other case reports the calcitriol dosage was increased for a variety of reasons (some incompletely documented) (253–257). Despite these contrasting views on the natural history of hypoparathyroidism in pregnancy, there is general agreement (244, 245, 248, 253, 254,

258) that in *late* pregnancy and the puerperium, hypercalcemia may result unless the calcitriol is discontinued, or the dosage is decreased below the prepartum requirement. Since this effect is even more pronounced in those who breast-feed, and since PTHrP is found at high concentrations in the breast during late pregnancy and lactation (further discussed in *Section IV.B.6*, below), the pregnancy-related rise in 1,25dihydroxyvitamin D production may be regulated by PTHrP (secreted from the breast) in these hypoparathyroid women.

Calcitriol (rather than vitamin D or calcifediol) has typically been prescribed for hypoparathyroidism in pregnancy, and the dosage needed may range from 0.5–3.0 μ g daily. Chronic maternal hypocalcemia must be avoided because it has been associated with the development of intrauterine hyperparathyroidism and death in the fetus (*Section III.F*, below).

Further illumination of the role of PTH in pregnancy has come from cases of pseudohypoparathyroidism in pregnancy. Pseudohypoparathyroidism is a heterogeneous group of genetic syndromes characterized by hypocalcemia due to PTH resistance (259). Although the data are limited, Breslau and Zerwekh (260) noted a normalization of serum calcium levels in two pregnant women with pseudohypoparathyroidism (probably type 1b). Before pregnancy the patients had hypocalcemia, markedly elevated PTH levels, and low 1,25-dihydroxyvitamin D levels. During four pregnancies (two for each patient), the serum calcium levels were normal, their PTH levels were halved, and the 1,25-dihydroxyvitamin D levels increased 2- to 3-fold. Contributions of 1,25-dihydroxyvitamin D from placental and fetal sources might have accounted for these findings; Zerwekh and Breslau (261) noted elsewhere that the placental production of 1,25-dihydroxyvitamin D was no different between placentas obtained from pseudohypoparathyroid women and controls. Alternatively, it is possible that the hormonal milieu of pregnancy lessened the renal resistance to PTH and PTHrP and thereby increased the formation of 1,25-dihydroxyvitamin D. It is apparent from Breslau's observations that estrogens alone cannot be the explanation for such an improvement during pregnancy, because the same two pseudohypoparathyroid women were not improved by treatment with an oral contraceptive. In any case, calcitriol supplementation in these patients should be monitored carefully and adjusted during pregnancy. The progeny of these pregnancies are also at risk of intrauterine, fetal hyperparathyroidism (262, 263), perhaps because of relative maternal hypocalcemia during pregnancy.

H. Summary

The fetal demand for calcium, which largely occurs during the third trimester, is met by a doubling of free and bound maternal 1,25-dihydroxyvitamin D levels, which, in turn, partly mediate a doubling of the intestinal absorption of calcium. Some of the increased intestinal calcium absorption may be mediated by PRL or other hormones of pregnancy. Further, the increase in 1,25-dihydroxyvitamin D may be largely independent of changes in PTH, since PTH levels are typically low or normal at the time of the increase in 1,25dihydroxyvitamin D. The increased calcium intake and absorption leads to a marked increase in renal calcium excretion (absorptive hypercalciuria). The serum ionized calcium is normal, despite a fall in total serum calcium caused by a reduction in the albumin-bound fraction. Calcitonin and PTHrP are both elevated, particularly in the latter half of gestation, but the physiological importance of these hormones in pregnancy is not known. The typical changes in calcium and calcitropic hormone levels during pregnancy are depicted schematically in Fig. 1.

Bone resorption is increased during late pregnancy, as evidenced by a rise in the levels of serum and urine markers of bone resorption in the third trimester, and this may indicate that maternal skeletal calcium stores are mobilized during the time of rapid fetal accretion of calcium. As noted at the beginning of Section II.E.2, bone density studies during pregnancy have been of insufficient precision to determine whether this increased bone resorption results in significant loss of skeletal calcium during pregnancy or the third trimester. Retrospective epidemiological studies (although not definitive) have generally found no effect of parity on the risk of osteoporosis or fractures in later life. Uncommonly, pregnancy may be associated with osteoporosis and fractures, particularly if the woman enters pregnancy with a low peak bone mass. A distinct disorder, focal, transient osteoporosis of the hip in pregnancy, is not likely due to altered calcitropic hormone levels and calcium physiology.

Primary hyperparathyroidism in pregnancy has been classically associated with adverse fetal or neonatal outcomes, but the milder, asymptomatic form of primary hyperparathyroidism most often seen today may not share such outcomes. Maternal hypoparathyroidism may be improved in pregnancy by increased intestinal absorption of calcium, possibly mediated by increased production of 1,25-dihydroxyvitamin D caused by PTHrP or some other non-PTH factor. A similar improvement in biochemical indices has been seen in pregnant women with pseudohypoparathyroidism. In both hypoparathyroid and pseudohypoparathyroid women, maternal hypocalcemia may adversely affect the fetus and must be avoided.

The pregnant rat model differs from the human condition in several important respects (Table 1). The rat normally develops a form of secondary hyperparathyroidism in the last several days of pregnancy, prompted by a fall in the maternal serum-ionized calcium at the time of rapid fetal accretion of calcium. 1,25-Dihydroxyvitamin D increases late in gestation in rats, approximately 1 week after the rise in intestinal calcium absorption. This indicates that mechanisms independent of 1,25-dihydroxyvitamin D may contribute to the increased intestinal calcium absorption in rats.

III. Fetal-Placental Physiology and Pathophysiology

A. Fetal adaptive goals

With respect to calcium physiology, the fetal-placental unit has two main adaptive goals. One is to provide sufficient calcium to mineralize the skeleton, and the other is to maintain an extracellular level of calcium that is physiologically appropriate for fetal tissues (*i.e.*, for cell membrane stability, blood coagulation, etc). A human fetus typically accumulates 21 g of calcium by term, and 80% of this calcium is accumulated in the third trimester alone, necessitating an average daily transfer of 200 mg calcium (4). Similarly, the fetal rat accretes less than 0.5 mg calcium in the first 17 days of gestation, and about 12 mg calcium in the remaining 5 days of gestation (6). To attain the required amount of calcium and regulate the fetal calcium level, the fetus makes use of the placenta, kidneys, bone, and intestine. The studies reviewed herein will demonstrate that the fetal-placental unit functions relatively independently of the mother, such that it is capable of mineralizing the fetal skeleton and maintaining a normal blood calcium, even in the presence of significant maternal hypocalcemia and vitamin D deficiency. In addition, this section will show that PTHrP is a major regulator of placental calcium transport, while PTHrP and PTH may both act on fetal bone and kidneys to regulate the blood calcium.

Human handling of placental calcium transport must be largely inferred from data that have been obtained from studies in sheep, pigs, rats, and mice. Therefore, it must be emphasized that mice and rats have hemochorial placentas that are structurally very similar to those of humans (264– 267). In contrast, the epitheliochorial placentas of sheep and pigs differ significantly in structure from the human hemochorial placenta, and may, therefore, be functionally different as well (266).

B. Mineral ions and calcitropic hormones

1. *Calcium.* In humans, rodents, sheep, cattle, monkeys, and other mammals, the fetal blood calcium (total and ionized) is maintained at a higher level than in the maternal circulation (268–275). This elevation is mainly due to an increase in the ionized calcium level (274). Ionized calcium is approximately 80% of the total calcium in fetal rodents (276); only a small fraction is bound to albumin.

In fetal rats, there is a progressive rise in total and ionized calcium over the last week of gestation, corresponding to the time of a progressive decline in fetal pH (277–279). Data are lacking on precisely how early in gestation the fetal blood calcium begins to exceed the maternal. In sheep, fetal hypercalcemia has been detected as early as the 35th day of gestation (280, 281). In humans, fetal hypercalcemia was documented at 15–20 weeks of gestation (by fetoscopy) (282) and at delivery of preterm singleton and twin pregnancies (mean gestational age 33 weeks) (283).

Two physiological models could explain fetal hypercalcemia: either the fetus maintains a fixed positive gradient of calcium with respect to the maternal level, or the fetus maintains a high, fixed level of calcium. Evidence from rat and mouse models indicates that the fetus sets its blood calcium at a higher level *independently* of the maternal calcium level. For example, in rats, the fetal blood calcium is unchanged in the presence of severe maternal hypocalcemia due to a calcium-restricted diet (284), vitamin D deficiency (29, 84, 285), or thyroparathyroidectomy (46, 134). The calcium gradient from mother to fetus is increased in these fetuses because the maternal blood calcium is lower. When both the pregnant rat and its fetus are thyroparathyroidectomized, the fetus still maintains a higher blood calcium level than the mother (286, 287). Also, in genetically engineered mice, maternal hypercalcemia due to heterozygous ablation of the calcium-sensing receptor (CaSR) gene does not affect the blood calcium level set by normal fetuses (288). Similarly, heterozygous calcium-sensing receptor knockout fetuses establish a constant, abnormally high fetal blood calcium level, regardless of whether the mother is heterozygous (and therefore hypercalcemic) or normal (288). The apparent "calcium gradient" is lower in offspring of these heterozygous mice, due to maternal hypercalcemia. Finally, acute alterations in the maternal blood calcium of rodents and primates (by calcium, 1,25-dihydroxyvitamin D, calcitonin, PTH, or EDTA infusions) are not reflected by much perturbation in the fetal blood calcium (232, 289–292).

Others have reported a fall in the fetal blood calcium after maternal parathyroidectomy in rats (52–54). The fetal blood calcium was normal between the 12th and 17th day of gestation, but fell during the period of rapid fetal skeletal calcium accretion. Therefore, these data indicate that the ability of the fetal rat to set its blood calcium may break down during the time of rapid accretion of calcium by the skeleton, if the mother has been parathyroidectomized.

In summary, from early pregnancy, mammalian fetuses have higher levels of blood calcium than their mothers, mainly due to an increase in the ionized calcium level. The fetus does not establish a particular calcium gradient with respect to the maternal blood calcium; instead, it establishes a particular fetal blood calcium level, irrespective of the ambient maternal blood calcium level. This ability persists in the presence of significant maternal hypocalcemia of various causes, but may be impaired during the time of rapid accretion of calcium by the skeleton. The physiological importance of fetal hypercalcemia is not known.

2. *Phosphate*. Fetal phosphorus levels are higher than maternal in rats (279) and humans (32, 270, 273). This suggests that phosphate may be actively transported across the placenta, but the regulators of this active transport are unknown (293). PTHrP and PTH do not stimulate placental transport of phosphate in sheep (294); vitamin D may have a role (295).

3. *PTH.* Fetal parathyroid glands of rats and sheep contain PTH mRNA (114, 116), and PTH immunoreactivity is present in human fetal parathyroid glands as early as 10 weeks of gestation (296). These findings indicate that fetal parathyroid glands are capable of synthesizing PTH early in gestation. Furthermore, PTH detected in the fetal blood likely derives from fetal sources alone. Intact PTH does not cross the placenta of nonhuman primates, sheep, and rodents (230–232) and probably does not cross the human placenta.

The following evidence indicates that fetal parathyroid glands appear to contribute to calcium homeostasis, by secretion of PTH or PTHrP. Fetal thyroparathyroidectomy in sheep and fetal decapitation in rats caused hypocalcemia (52, 297, 298), and mice lacking the PTH/PTHrP receptor gene are hypocalcemic *in utero* (299). PTH can be regulated by the ambient fetal blood calcium, since EDTA-induced fetal hypocalcemia has been found to induce a rise in fetal PTH levels in rats (300), cattle (275), and rhesus monkeys (301), although another study in rhesus monkeys found no fetal PTH re-

sponse (271). Removal of a maternal parathyroid adenoma was followed by a rise in amniotic fluid PTH levels and a decline in the amniotic fluid calcium level during a human pregnancy (302). Since maternal PTH cannot cross the placenta, the findings in this case have been interpreted to indicate that fetal PTH secretion can be influenced by the maternal blood calcium (302).

In fetal humans and other animals, immunoreactive PTH blood levels have been found to be undetectable or very low (*i.e.*, <0.5 pmol/liter) with respect to maternal PTH level near the end of gestation (17, 32, 35, 38, 43, 110, 124, 268, 275, 303–310). Little information is available on PTH levels earlier in gestation. One study in fetal rats found that the PTH level declined in the last several days of gestation as the serum ionized calcium rose (277), while two cross-sectional studies in preterm humans found that the fetal PTH level was not lower than the maternal PTH level (283, 308).

In summary, the available evidence suggests that the fetal parathyroids are capable of synthesizing PTH. Since blood levels of PTH have been typically found to be low in late gestation at a time when the fetal blood calcium is high, other factors must determine the fetal blood calcium level. The precise role of PTH in normal fetal calcium homeostasis will be clarified by ablating the PTH gene in mice.

4. 1,25-Dihydroxyvitamin D. Although maternal vitamin D deficiency reduces fertility and litter size in the rat (84, 85), evidence from several animal models indicates that 1,25-dihydroxyvitamin D is not necessary for normal fetal calcium and bone metabolism. In pregnant rats, sheep, and pigs that were hypocalcemic due to severe vitamin D deficiency, the fetuses maintained completely normal blood calcium and phosphate levels and had fully mineralized skeletons at term, as determined by total weight, ash weight, and calcium content of femurs (29, 84, 284, 285, 311). Each of these studies is limited by the possibility that low levels of vitamin D might have reached the fetus.

Further evidence that 1,25-dihydroxyvitamin D is not needed for normal fetal calcium and bone homeostasis comes from the 1 α -hydroxylase-deficient Hannover pig model, in which the fetuses of homozygous 1,25-dihydroxyvitamin Ddeficient sows also maintained completely normal blood calcium and phosphate levels and fully mineralized their skeletons (79). Nephrectomy of fetal rats did not affect fetal blood calcium or phosphate levels when measured 48 h later, even though fetal 1,25-dihydroxyvitamin D levels fell (286). Also in fetuses of vitamin D-deficient rats, normal levels of calbindin_{28K}-D and calbindin_{9K}-D were found in the placenta, intestine, and other tissues (132, 311, 312). In addition, fetal mice that lack the gene encoding the receptor for 1,25dihydroxyvitamin D are born with normal skeletons (313, 314).

Some data from humans lend support to the observation that 1,25-dihydroxyvitamin D is not needed for normal fetal calcium and bone metabolism. At term, the cord blood calcium and skeletal mineralization is completely normal in the offspring of vitamin D-deficient mothers (315–317). It is only in the first or second week after birth that hypocalcemia develops; skeletal demineralization and other rachitic changes are typically not detectable until 1 or 2 months of age (see *Section V.E*, below).

These observations of a minimal effect of vitamin D deficiency on fetal calcium and skeletal metabolism do not mean that 1,25-dihydroxyvitamin D is inactive or has no role in fetal life. In rats, the receptor for 1,25-dihydroxyvitamin D appears on day 13 of gestation in the mesenchyme that will subsequently condense to form the skeletal tissues, and by day 17 of gestation it is expressed in proliferating and hypertrophic chondrocytes, and osteoblasts of limb buds and the vertebral column (318). The widespread expression of the vitamin D receptor early in fetal skeletal development suggests an important role for its ligand in fetal bone development, but evidence for this postulated role has not yet been found. Further studies manipulated the 1,25-dihydroxyvitamin D level in fetal animals to test the role of this hormone. Infusion of antibody to 1,25-dihydroxyvitamin D decreased the ovine fetal blood calcium level (66). 1,25-Dihydroxyvitamin D given to pregnant guinea pigs increased the fetal calcium and phosphate levels (319). Bilateral nephrectomy in fetal sheep resulted in reduced ionized and total calcium and increased phosphate and PTH levels; these changes could be reversed by administration of 1,25-dihydroxyvitamin D to the fetus (295). Since these changes could be attributable to uremia and not loss of the renal 1α -hydroxylase enzyme, additional fetuses underwent bilateral ureteral sectioning alone. This surgical procedure allowed urine to drain into the fetal peritoneal cavity while retaining functional kidneys in situ. In these fetuses, ureteral sectioning had no effect on fetal calcium or calcitropic hormone levels. Thus, at least in the absence of normal renal function, 1,25-dihydroxyvitamin D may have a substantial influence on fetal mineral ion homeostasis.

1,25-Dihydroxyvitamin D does not readily cross the placenta in rats (320); consequently, circulating levels of 1,25dihydroxyvitamin D in the fetus are largely derived from fetal sources. The fetal kidneys and placenta possess the 1α -hydroxylase enzyme and convert 25-hydroxyvitamin D to the active form (1, 25-dihydroxyvitamin D) (75, 76). The contribution of the fetal kidneys must be significant, since fetal nephrectomy reduced the fetal 1,25-dihydroxyvitamin D levels in sheep and rats (66, 286). Fetal blood levels of 1,25-dihydroxyvitamin D are typically lower than maternal levels in humans (37, 56, 304, 321), but umbilical artery levels of 1,25-dihydroxyvitamin D are slightly higher than umbilical venous levels, confirming the contribution of the fetal kidneys (37). 25-Hydroxyvitamin D levels have been found to be roughly equal to maternal levels (37, 56); this is not surprising since 25-hydroxyvitamin D readily crosses the placenta in rats (322). Levels of 24,25-dihydroxyvitamin D correlate with, but are typically lower than, maternal levels at term in humans (283, 307, 321).

In summary, evidence from animal models indicates that deficiency of 1,25-dihydroxyvitamin D impairs neither fetal skeletal formation and calcification nor the ability of the fetus to maintain a normal blood calcium. Although these data suggest a limited role for 1,25-dihydroxyvitamin D in the fetus, fetal production of 1,25-dihydroxyvitamin D and the vitamin D receptor mandate a continued search for fetal roles for 1,25-dihydroxyvitamin D readily crosses the placenta and can be 1α -hydroxylated by the fetal kidneys. However, 1,25-dihydroxyvitamin D does not cross the placenta, and fetal blood levels of 1,25-dihydroxyvitamin D are low.

5. *Calcitonin*. Immunoreactive calcitonin can be detected in human fetal thyroid glands from as early as the 15th week of gestation (323), and fetal calcitonin levels are maintained at higher levels than maternal (35, 37, 86, 88, 89, 269, 273, 304, 324). Maternal calcitonin cannot cross the placenta (325). The increased fetal levels of calcitonin are thought to reflect increased synthesis, but the metabolism and clearance of calcitonin have not been studied in fetal animals.

Several acute experimental perturbations suggest a role for calcitonin in fetal calcium homeostasis. Infusion of calcitonin antiserum to fetal rats at day 21.5 of gestation slightly increased the fetal blood calcium 1 h later (326), while fetal injection of calcitonin caused hypocalcemia and hypophosphatemia (327). However, fetal thyroidectomy with subsequent T_4 replacement did not affect the fetal blood calcium in sheep, indicating that fetal thyroidal C cells alone may not affect the regulation of the blood calcium level (298). Therefore, the precise role of calcitonin in fetal calcium homeostasis and skeletal metabolism has not yet been established.

6. PTHrP. Studies of PTH bioactivity in human umbilical cord blood (as determined by an in vitro cytochemical bioassay) found high PTH-like bioactivity, while immunoreactive PTH was simultaneously found to be undetectable or low (38, 303, 328). Subsequently, it has been recognized that human cord blood PTHrP levels are significantly higher than the simultaneous maternal levels at term (43, 310). When both PTH 1-84 and PTHrP 1-86 were simultaneously measured by two-site immunoradiometric assays [and expressed in equivalent units (picomoles/liter)], human cord blood PTHrP levels were 2–4 pmol/liter, up to 15-fold higher than the levels of PTH (0.2–0.5 pmol/liter) (43, 110, 124). It has yet to be confirmed that PTHrP accounts for the high PTH-like bioactivity in human cord blood; however, studies in fetal pigs (329) and sheep (116, 330) found that the levels of PTHrP and PTH-like bioactivity were tightly correlated in late gestation and the neonatal period.

As noted earlier (*Section II.B.6*), PTHrP may be a prohormone that is processed into separate circulating fragments, each of which may have different functional roles and receptors (100). Although the structures of these fragments have been deduced from studies of tumor cell lines transfected with the PTHrP gene, it has yet to be determined which of these fragments normally circulate in fetal life. Full-length PTHrP has twice the molecular weight of PTH; since PTH cannot cross the placenta, PTHrP probably does not either. PTHrP 1–86 did not cross the placentas of sheep and goats (113); however, the possibility that smaller, biologically active fragments of PTHrP might cross the placenta has not been evaluated.

PTHrP is produced in many sites throughout the developing embryo and fetus, including the fetal parathyroid glands (116, 331), skeletal growth plate (332, 333), trophoblast cells of the placenta (114, 331), amnion (106, 334), chorion (334), umbilical cord (115), and many other organs. All of

these sites may contribute to the circulating level of PTHrP in the fetus and may thereby be relevant to fetal calcium and bone metabolism. Since venous umbilical PTHrP levels were higher than umbilical arterial levels in pigs, the placenta may be an important source of systemically circulating PTHrP in the fetus (329). Due to local production of PTHrP by the umbilical cord (115), the level of PTHrP in cord blood might not accurately reflect the systemic level of PTHrP, but this has not been tested.

PTHrP has multiple possible roles during embryonic and fetal development (335). PTHrP gene-ablated mice have abnormalities of chondrocyte differentiation (336) and aberrant breast development (337). PTHrP may also be an important regulator of the fetal blood calcium. PTHrP levels correlate with the fetal ionized calcium levels in pigs (329). In genetically engineered mice, homozygous ablation of the PTHrP gene results in a fetal blood calcium no higher than that of the mother (299). In sheep, fetal parathyroidectomy causes hypocalcemia that can be reversed by PTH or PTHrP infusion (297, 298, 338). Since PTH normally circulates at low or undetectable levels in the fetus near term (Section III.B.3, above), it is possible that the hypocalcemic effect of fetal parathyroidectomy is at least partly due to the loss of PTHrP produced by the parathyroids. In the next section (III.C), the unique role of PTHrP in stimulating placental calcium transport will be discussed.

In summary, PTHrP is produced by diverse fetal tissues and circulates in fetal blood at levels higher than adult levels. PTHrP appears to regulate the fetal blood calcium as well as fetal-placental calcium transport.

C. Fetal-placental calcium transport

Calcium is actively transported across the placenta (339, 340). The site of active transport is likely at the fetus-facing basement membrane of the syncytiotrophoblast cells in the human and at the trophoblast cells and the basal surface of the endoderm of the intraplacental yolk sac in rodents (341, 342). The active transport of calcium may be mediated by a Ca^{2+} -ATPase present at these same sites (339, 342). This enzyme's activity can be inhibited by dinitrophenol, ouabain, quercetin, and antibody to the human erythrocyte plasma membrane calcium pump (339, 342). Calcium-binding proteins in the placenta and yolk sac are also thought to be involved in active placental calcium transport. The placental calbindin9K-D mRNA and protein levels increase over the last week of gestation in rats (129, 343) and mice (130, 133) and are unaffected by maternal vitamin D deficiency (132, 311). Transplacental transport of calcium is generally found to be a one-way process, *i.e.*, fetal-to-maternal flow of calcium is typically less than 1% of the forward (maternal-to-fetal) flow (344, 345). In rhesus monkeys, backflux was reported to be 80% of the forward flow (345); it is not certain whether this represented a true species difference or methodological differences. It has not been determined when active transport of calcium begins in gestation, due to technical difficulties involved in studying placental physiology early in gestation. However, active transport of calcium must be underway by the third trimester in humans, which is the time of rapid skeletal mineralization and peak fetal calcium requirement. 1. Maternal hormones. Maternal hormones might influence the fetal-placental calcium transport process by raising or lowering the ambient maternal calcium level, and by direct effects on the placenta. However, several lines of evidence from animal experiments indicate that fetal-placental calcium transport and net maternal-fetal calcium transfer are maintained relatively independently of maternal hypocalcemia or hormone deficiencies. In pregnant sheep, maternal hypocalcemia due to parathyroidectomy or dietary calcium restriction did not affect the rate of fetal-placental calcium transfer as directly assessed in placental perfusion experiments (297, 346); in addition, the fetal blood calcium, phosphate, PTH, and 1,25-dihydroxyvitamin D levels were all unchanged by maternal hypocalcemia (284, 347). The finding of a "normal" rate of calcium transfer across the placenta indicates that the fetal-placental unit must be working harder to extract the normal amount of calcium from a reduced amount of maternal calcium presented to the placenta. Indeed, the following observation from intact fetal rats confirmed that the rate of placental calcium transfer is up-regulated in response to parathyroidectomy-related chronic maternal hypocalcemia. In this experiment, a maternal calcium infusion caused a marked, acute rise in the blood calcium of fetuses from parathyroidectomized rats, but had no effect on the fetuses of normal rats (348). Such an up-regulation in placental calcium transfer may compensate for a low ambient maternal blood calcium and permit a normal amount of calcium to be transferred by the end of gestation.

Several additional studies have examined only indirectly the effect of other maternal hormone deficiencies on placental calcium transfer. In these studies, net fetal accumulation of calcium at term was used as an index of placental calcium transfer during pregnancy in vitamin D-deficient rats (311), thyroidectomized, T₄-supplemented ("calcitonin-deficient") sheep (349, 350), and in sheep that received daily administration of PRL and/or bromocriptine (351). Since placental calcium transport was not directly assessed in these studies, conclusions cannot be drawn about the effect of maternal vitamin D, calcitonin, and PRL deficiency on placental calcium transport.

2. *Fetal hormones.* The role of 1,25-dihydroxyvitamin D in fetal-placental calcium transport has not been thoroughly studied. Receptors for 1,25-dihydroxyvitamin D are present in the placentas of humans, rats, and sheep and might, therefore, have a role in placental calcium physiology (352–355). In placental perfusion models in rats, guinea pigs, and sheep, pharmacological doses of 1,25-dihydroxyvitamin D or 1 α -cholecalciferol increased the fetal blood calcium, transport of calcium across the placenta, and mineral content of ashed fetuses (319, 356, 357). Also, prior nephrectomy of fetal sheep reduced calcium transfer in the placental perfusion model, and this effect could be partly restored by administering 1,25-dihydroxyvitamin D (339). Thus, the evidence indicates a possible role for 1,25-dihydroxyvitamin D in fetal calcium homeostasis, but this role is not yet well defined.

The role of fetal calcitonin is even less well established. In intact fetal sheep, calcitonin has been found to reduce the PTHrP-mediated increases in the apparent rate of calcium transfer (349, 358). In contrast, fetal thyroidectomy with sub-

sequent T_4 replacement did not alter placental calcium transfer in sheep, indicating that loss of fetal calcitonin does not perturb the placental calcium pump (298).

The role of the parathyroid gland in fetal calcium metabolism has been extensively studied. Fetal thyroparathyroidectomy in sheep and fetal decapitation in rats resulted in a lower fetal blood calcium, such that the maternal-fetal calcium gradient was obliterated (297, 298, 359). In addition, when the thyroparathyroidectomized sheep or rat fetuses were removed so that the placentas could be artificially perfused in situ, active transport of calcium across these experimentally perfused placentas was found to be reduced (297, 298, 359, 360). These findings indicate that the parathyroid glands have a critical role in maintaining the fetal blood calcium and the active transport of calcium across the placenta. Infusion of autologous blood from fetuses with intact parathyroid glands restored calcium transport across the perfused placentas of thyroparathyroidectomized fetal sheep (298). However, PTH failed to restore the active transport of calcium in fetal sheep or rats under these conditions (338, 359, 361).

The placenta may be able to transport calcium actively to some extent even in the absence of fetal parathyroid glands. When rat dam and fetus were both thyroparathyroidectomized, the fetus maintained a higher blood calcium level than the dam (286, 287). Although calcium transport was not measured, the fact that relative fetal hypercalcemia was maintained may indicate that some capability for active transport of calcium persists after fetal parathyroidectomy.

In the studies of thyroparathyroidectomized fetal sheep, synthetic PTHrP molecules of amino acid lengths 1-141, 1-86, and 67-86 were found to stimulate placental calcium transport in the experimentally perfused placentas (338, 361-363). These results suggested that PTHrP, perhaps produced by the parathyroid glands, stimulates active transport of calcium across the placenta. In contrast, PTHrP 1-34, which contains only the PTH-like amino-terminus, failed (like PTH) to stimulate calcium transport in this model. Studies in genetically engineered mice support the hypothesis that PTHrP stimulates placental calcium transport. A reduction in blood calcium to the maternal level and reduced placental transfer of calcium has been found in homozygous PTHrP-gene knockout fetal mice (299). The placental transfer of calcium was acutely increased in the homozygous fetuses by treatment with PTHrP 1-86 or PTHrP 67-86, but not by PTHrP 1-34 or intact PTH (299). These data suggest that PTHrP increases placental calcium transport by acting through a receptor distinct from the PTH/PTHrP receptor, since the PTH/PTHrP receptor is stimulated equally by amino-terminal PTH and PTHrP. This hypothesis is further supported by the studies of fetal mice homozygous for deletion of the PTH/PTHrP receptor gene. These mice are hypocalcemic, but placental calcium transport in these fetuses is increased (299).

These data on the effects of PTHrP in parathyroidectomized fetal sheep and genetically engineered mice are not supported by the following observations in fetal rats. In the perfused placentas obtained from intact or decapitated fetal rats, active transport of calcium was found to be stimulated slightly by amino-terminal PTH or PTHrP, but not by fragments of PTHrP that do not contain the amino terminus (359, 360). This could reflect a true species difference or methodological differences.

Taken together, the studies in genetic knockout mice, and those in thyroparathyroidectomized fetal sheep, suggest that in fetal life, PTHrP is necessary to maintain the normal fetal hypercalcemia and at least part of the active transport of calcium across the placenta. This transport is regulated in part by a midmolecular portion of PTHrP acting on a novel (as yet uncloned) receptor distinct from the PTH/PTHrP receptor. Recent evidence suggests that the structure of this midmolecular fragment of PTHrP may encompass amino acids 38-94 (100, 364), but this has yet to be confirmed by RIAs of fetal blood. The studies in sheep suggested that the parathyroid glands secrete the PTHrP that controls placental calcium transport; however, direct evidence to support this hypothesis is not yet available. Finally, the physiological importance of the actions of fetal 1,25-dihydroxyvitamin D and calcitonin require further exploration.

D. Renal handling of calcium and the amniotic fluid

The fetal kidneys may play a role in regulating the fetal blood calcium level, by adjusting the relative reabsorption and excretion of calcium and phosphate by the renal tubules in response to the filtered load and other factors, such as PTHrP and/or PTH. The fetal kidneys may also participate by synthesizing 1,25-dihydroxyvitamin D. However, little hard data are available on fetal kidney function and its relative importance in regulating the fetal blood calcium. As noted previously, nephrectomy in fetal lambs resulted in hypocalcemia, hyperphosphatemia, and reduced placental calcium transfer; these effects were attributed to loss of renal production of 1,25-dihydroxyvitamin D (295, 339). The fetal renal tubular function may be under the control of PTHrP or PTH in fetal life, since thyroparathyroidectomy in fetal sheep resulted in an increase in fractional excretion of calcium by the fetal kidneys and reduced phosphate excretion (365, 366). These effects were reversed by treatment with amino-terminal fragments of either PTH or PTHrP (365, 366). Thyroparathyroidectomy also reduced placental calcium transport in fetal sheep (Section III.C., above); therefore, the hypocalcemia in thyroparathyroidectomized fetuses likely results from the combined effects of reduced influx of calcium across the placenta, increased excretion of calcium by the fetal kidneys, and possible effects of the loss of PTHrP and/or PTH on the fetal skeleton and renal 1α -hydroxylase.

In fetal life, renal excretion of calcium does not necessarily represent a permanent loss of calcium for the fetus. Fetal urine excretion is probably the major source of fluid and solute in the amniotic fluid, while fetal swallowing is likely the major pathway for clearance of amniotic fluid and is a pathway by which excreted calcium can be made available again to the fetus (367). The volume and composition of amniotic fluid have been used as indirect measures of fetal renal function. The amniotic fluid ionized calcium level has been found to be constant between 14–15 weeks of gestation and term in humans, while the total calcium and phosphate levels decline over the same interval (13). This may indicate that renal excretion of calcium is equally balanced by fetal

swallowing and intestinal reabsorption of calcium. However, other sources may contribute to the amniotic fluid, including secretions from the respiratory tract, and exchange of fluid and/or solute across the fetal skin, fetal membranes (amnion, chorion and chorionic plate), and umbilical cord (367). Little is known about the relative contribution of these sites to the volume and composition of the amniotic fluid, and hence uncertainty remains about how accurately the amniotic fluid composition reflects renal function. Nevertheless, amniotic fluid represents a pathway by which excreted calcium may be recirculated to the fetus.

E. Skeletal calcium metabolism

The fetal skeleton may well have two interdependent roles - substantial growth during late fetal life and participation in fetal calcium homeostasis. Several lines of evidence indicate that the fetal skeleton participates in fetal calcium homeostasis and that skeletal calcium may be mobilized in response to reduced transfer of calcium from mother to fetus. Maternal hypocalcemia due to thyroparathyroidectomy or calcitonin infusion increased the basal level of bone resorption in subsequently cultured fetal rat bones (368, 369). These effects were blocked by prior fetal decapitation, which is thought to mimic the effects of thyroparathyroidectomy (368, 369). Further, the fetal parathyroid glands enlarge in response to maternal hypocalcemia in the rat (47, 370), and fetal femur length and mineral ash content are subsequently reduced (53). Additional, recent observations in genetically engineered mice suggest a role for the skeleton in fetal calcium homeostasis. The ionized calcium of fetal mice that lack the PTH/PTHrP receptor gene is lower than that of fetal mice that lack the PTHrP gene, despite the fact that placental calcium transport is supranormal in PTH/PTHrP receptor knockout fetuses and subnormal in PTHrP knockout fetuses (299). Lack of bone responsiveness to amino-terminal PTH and PTHrP may well, therefore, contribute to the hypocalcemia in mice without PTH/PTHrP receptors.

Intact fetal parathyroid glands are needed for normal skeletal development, since fetal thyroparathyroidectomy in sheep caused decreased ash content and rachitic changes in the fetal skeleton by term (371, 372). These effects could be partly reversed or prevented by fetal calcium and phosphate infusions; thus, much of the effect of fetal parathyroidectomy was caused by a decrease in blood levels of calcium and phosphate (372). However, although bone formation parameters were corrected by the calcium and phosphate infusion, bone resorption parameters remained abnormal (reduced resorption cavities, reduced osteoclast numbers). Therefore, functioning fetal parathyroids (and, therefore, parathyroid gland-produced PTH and/or PTHrP) are required for normal fetal bone resorption and mineralization.

PTHrP is clearly important for growth plate development, because absence of PTHrP (in the PTHrP gene-knockout mouse) results in a chondrodysplasia characterized by accelerated differentiation of growth cartilage and adjacent endochondral bone (336). This effect of PTHrP must be mediated by the PTH/PTHrP receptor, because mice homozygous for the absence of the PTH/PTHrP receptor have a similar skeletal phenotype (373). However, these gene knockout experiments could not determine whether systemically delivered PTHrP adds to the likely role of PTHrP produced in the growth plate itself.

In summary, normal mineralization of the fetal skeleton requires intact fetal parathyroid glands and adequate delivery of calcium to the fetal circulation. The fetal skeleton can participate in the regulation of fetal calcium homeostasis, probably through actions of PTHrP and/or PTH. In addition to its effects on regulating placental calcium transport, PTHrP is required for normal skeletal development. Evidence from experimental vitamin D deficiency and the vitamin D receptor knockout indicates that 1,25-dihydroxyvitamin D may not be required by the developing skeleton.

F. Fetal response to maternal hyper- or hypoparathyroidism

As discussed earlier (*Section II.F*), in humans, maternal hypercalcemia due to primary hyperparathyroidism may suppress the fetal parathyroid glands, since hypocalcemia can be present at birth (225, 235). The fetal parathyroid glands may also be suppressed when the mother has hypercalcemia due to familial hypocalciuric hypercalcemia (241–243, 374).

On the other hand, maternal hypoparathyroidism in human pregnancy has been associated with the development of intrauterine, fetal hyperparathyroidism. This condition is characterized by fetal parathyroid gland hyperplasia, generalized skeletal demineralization, subperiosteal bone resorption, bowing of the long bones, osteitis fibrosa cystica, rib and limb fractures, and low birth weight (246, 375-378). Spontaneous abortion, stillbirth, and neonatal death have also been associated with this condition (379-381). Similar skeletal findings have been reported in the fetuses and neonates of women with pseudohypoparathyroidism (262, 263), renal tubular acidosis (382), and chronic renal failure (383). Although these skeletal changes have been interpreted to indicate fetal hyperparathyroidism, no serum measurements of intact PTH (or PTHrP) have been reported for this condition, and the serum calcium level has been generally reported to be normal.

G. Integrated fetal calcium homeostasis

Previous sections have demonstrated that the fetus maintains a blood calcium higher than the maternal level, and that the placenta and bone, and perhaps fetal kidney and intestine, transport calcium into and out of the bloodstream. However, the need for fetal hypercalcemia and the mechanisms by which it is maintained are not fully understood.

The need for an increased fetal blood calcium level is uncertain. It may well not be necessary for normal accretion of calcium by the developing skeleton, since, despite obliteration of the normal fetal-maternal calcium gradient (299), homozygous PTHrP-null fetal mice do not have a deficit in skeletal mineral content, as assessed by alizarin red staining and ash mineral content (336, 384).

In adult life, the CaSRs on the parathyroid glands and kidneys set the ambient serum calcium level, mainly by controlling the synthesis and secretion of PTH by the parathyroid glands and regulating renal tubular handling of calcium (385, 386). Inactivating or loss-of-function mutations of this receptor raise the set point for PTH secretion and increase renal calcium retention; these actions lead to hypercalcemia (385, 386). If the parathyroid gland CaSR were responsible for the elevated fetal calcium level, the set point would have to be different in fetal life, and one would expect to find increased or inappropriately normal PTH levels in the presence of the increased fetal calcium level. Instead, PTH in humans is normally low or undetectable at term (Section V.B.3), and the fetal blood calcium and PTH level are negatively correlated (387). Therefore, it is likely that neither the parathyroid CaSR nor PTH is responsible for the high fetal blood calcium; instead, the normally functioning CaSR suppresses the parathyroids in response to a high fetal calcium that is maintained by other processes. Further evidence that factors other than PTH derived from the parathyroids have a role in regulating the fetal blood calcium come from studies of thyroparathyroidectomized rat fetuses, which maintain a higher blood calcium than their simultaneously thyroparathyroidectomized mothers (286, 287). Such factors might include PTHrP (derived from the placenta, parathyroids, and other tissues), among other factors that might still be undiscovered.

That PTHrP has a role in maintaining the fetal blood calcium is suggested by the finding of a reduction in the fetal calcium to a level equal to that of the mother, in the PTHrP gene knockout fetuses (299). PTH levels are sharply increased in the PTHrP-null fetus (388), indicating that, in the absence of PTHrP, the parathyroids respond to regulate the fetal calcium level. Indeed, other evidence indicates that eliminating the amino-terminal actions of PTH, in addition to those of PTHrP, has a greater effect on reducing the fetal blood calcium than removing the PTHrP gene. The ionized calcium level of PTH/PTHrP receptor-null fetuses is *lower* than the ambient maternal level, despite the presence of a supranormal rate of placental calcium transfer (299).

Since the ionized calcium level of PTHrP-null fetuses is not higher than the maternal calcium level, this might indicate that PTH cannot make up for lack of PTHrP (including the effect of PTHrP to stimulate placental calcium transport); alternatively, in the absence of PTHrP, the fetal parathyroid CaSR may then control the regulation of the blood calcium, by stimulating PTH and setting the ionized calcium at the normal adult level. Further work is needed to determine whether either of these proposed mechanisms is correct.

Thus, an integrated model of normal fetal calcium homeostasis proposes that the fetal blood calcium is set at a level higher than maternal through the actions of PTHrP (among other potential factors). The parathyroid CaSR responds appropriately to this increased level of calcium and suppresses the parathyroids. 1,25-Dihydroxyvitamin D synthesis and secretion are, in turn, suppressed due to the effects of low PTH and high blood calcium and phosphate. PTHrP may be autonomously produced by the placenta and other tissues, or its production may be regulated. This model further proposes that the fetal blood calcium is maintained not only by flux of calcium across the placenta from the mother, but by contributions from fetal skeleton and kidney.

This tentative model needs to be tested further, and the physiological benefit of fetal hypercalcemia requires clarification.

H. Summary

The fetal-placental unit has adapted to rapidly extract calcium from the maternal blood stream in sufficient amounts to mineralize the fetal skeleton in late gestation. Fetal blood calcium and active transport of calcium across the placenta are regulated relatively independently of the levels of maternal calcium and calcitropic hormones. The fetus has a higher blood calcium than the ambient maternal calcium level. PTH and 1,25-dihydroxyvitamin D, the traditional calcitropic hormones, are present at low levels in the fetal circulation and may have a limited role in fetal calcium physiology. Calcitonin levels are elevated in the fetal circulation, but the role of calcitonin in fetal calcium homeostasis is uncertain. PTHrP is critical for maintaining normal fetal hypercalcemia and active transport of calcium across the placenta, although it is likely that other factors regulate placental calcium transport, as well. A midmolecular form of PTHrP stimulates placental calcium transport, through actions on a receptor that is distinct from the cloned PTH/PTHrP receptor.

The fetal-placental unit has a remarkable ability to meet its needs irrespective of maternal calcium or vitamin D levels. However, maternal hypercalcemia due to primary hyperparathyroidism or familial benign hypercalcemia can suppress the fetal parathyroid glands. In turn, maternal hypocalcemia due to hypoparathyroidism and pseudohypoparathyroidism (and other causes) can cause fetal parathyroid gland enlargement and increased resorption in the fetal skeleton.

The fetus does not set a calcium gradient against the maternal circulation; instead, the fetus sets its blood calcium at a particular level independently of the maternal value. The physiological role of this relative hypercalcemia is uncertain. The normal setting of this level requires the presence of a normal calcium-sensing receptor. The fetal blood calcium is not simply determined by the influx of calcium across the placenta. Rather, other important factors include fluxes of calcium in and out of the developing skeleton, and (probably to a lesser extent) renal tubular reabsorption and excretion of calcium, and reabsorption of calcium from swallowed amniotic fluid.

IV. Maternal Physiology and Pathophysiology During Lactation

A. Maternal adaptive goals during lactation

Albright and Reifenstein (1) reported that maternal losses of calcium during 9 months of lactation are 4-fold higher than the losses occurring during pregnancy. More specifically, the typical daily loss of calcium in breast milk has been estimated to range from 280–400 mg (389, 390), although daily losses as great as 1000 mg calcium have been reported (391). Accurate estimates of the calcium content of breast milk are complicated because the content varies within and between feedings (392, 393), between breasts of the same person (393), and among different mothers and ethnic groups (392). Although the calcium content of milk is lower at 6 months compared with 3 months postpartum (390, 392, 394, 395), the volume of milk produced at 6 months tends to be greater (390), and therefore the daily maternal loss of calcium may be greater when lactation extends to 6 months and beyond (390).

In a classic study in 1931, Donelson *et al.* (396) carefully measured dietary calcium intake and losses of calcium in feces, urine, and breast milk in a group of lactating women who exclusively pumped breast milk and did not permit their infants to suckle. Despite adequate vitamin D and calcium intake (in the form of cod liver oil and yeast), calcium balance was negative in these women throughout lactation (396). A similar study of three lactating women found a negative calcium balance (net loss of 1 g calcium) during the interval of greatest milk production, despite ingestion of supplemental calcium and phosphorus; only during the time of lessened milk production (weaning) did the calcium balance return to normal (391).

To compensate for the calcium requirements of lactation, the maternal adaptations could, in theory, include increased intestinal absorption of calcium, renal conservation of calcium, and increased resorption of calcium from the skeleton. In fact, the studies reviewed in this section indicate that a temporary demineralization of the skeleton is the main mechanism by which lactating humans and animals meet these calcium requirements. This demineralization does not appear to be mediated by PTH or 1,25-dihydroxyvitamin D, but may be mediated by PTHrP in the setting of a fall in estrogen levels.

B. Mineral ions and calcitropic hormones

The changes that occur in maternal calcium, phosphate, and calcitropic hormone levels during lactation, weaning, and postweaning are schematically depicted in Fig. 2.

1. *Calcium.* From the earliest measurements of total calcium to the newer reports of ionized calcium determinations in lactation, the blood calcium of lactating humans has been found to be normal or slightly increased (7, 397–399). More recently, with larger sample sizes, it has been shown that the mean ionized calcium level of exclusively lactating women is higher than that of normal controls (400, 401). Also, mothers nursing twins have been found to have significantly higher total calcium levels than mothers nursing singletons (402). Furthermore, occasionally substantial hypercalcemia may develop during lactation and resolve only at weaning (403).

In contrast, the data from lactating rats are conflicting: two reports found lactating rats to be slightly hypocalcemic compared with nonlactating controls (50, 404); another found normal calcium levels that could be decreased by feeding a low calcium diet (405), while a fourth found mild hypercalcemia (406). More intensive lactation (as determined by the relative rate of weight gain in the pups of a litter or by the number of pups in a litter) correlated with a lower maternal serum calcium (22, 50, 407, 408). Deer have also been found to have higher corrected serum calcium levels during lactation than after weaning (23). With abrupt weaning, the serum calcium of lactating rats typically rebounds into the hypercalcemic range (81, 409).

2. *Phosphate*. Serum phosphate levels are typically higher during lactation in humans, and, in some cases, exceed the



FIG. 2. Schematic illustration of the longitudinal changes in calcium, phosphate, and calcitropic hormone levels that occur during lactation and postweaning in humans. Normal adult ranges are indicated by the *shaded areas*. Data have been compiled from the following sources: total calcium (7, 397–399), ionized calcium (400, 401), phosphate (14, 398, 401, 410–412), PTH (27, 160, 398–401, 410, 411), 1,25-dihydroxyvitamin D (27, 58, 160, 397, 398, 410, 414), calcitonin (14, 397, 402, 420), and PTHrP (399–401, 411, 435). The progression in PTHrP levels has been depicted by a *dashed line* to reflect that the data are less complete.

normal range (14, 28, 398, 401, 410–412); similar results have been reported for lactating rats (50) and deer (23). Reabsorption of phosphate by the kidneys may be increased, although few measurements of tubular reabsorption of phosphate have been made (27, 160, 410, 411), and the data are conflicting. The increased serum phosphate levels may, therefore, reflect the combined effects of increased flux of phosphate into the blood from diet and from skeletal resorption (*Sections IV.E.1* and *IV.E.2*, below) in the setting of decreased renal phosphate excretion (*Section IV.D*, below).

3. PTH. Lactation had been described as a state of secondary hyperparathyroidism by Albright and Reifenstein (1) and, as with pregnancy, early PTH assay results appeared to confirm this hypothesis (413) [although some early assays reported normal levels (30, 55, 397, 414)]. However, intact PTH, as determined by a two-site IRMA, has been found to be reduced 50% or more in lactating women in both cross-sectional (399, 401) and longitudinal studies (27, 160, 398, 400, 411, 415-417). The intact PTH level remains low compared with that of nonlactating postpartum women, rising to normal (160), or above normal (27, 398, 410) after weaning. This postweaning increase in intact PTH level may be sustained for 2-3 months (27, 398). This elevation corresponds to the time when bone mineral is restored to the skeleton (see discussion on bone density during lactation, Section IV.E.2, below) (27, 410).

PTH, measured with a two-site IRMA, rises during lactation in rats (45, 49, 50). Under the stress of a low calcium diet, urine cAMP levels sharply increase (405), and PTH levels (N-terminal assay) rise even higher (21, 50). Similar effects have been seen in dams nursing larger litters (50, 408). However, a functioning parathyroid gland is not necessary to provide calcium to the milk of the lactating rat (418). After weaning in rats, PTH levels were found to fall within 6 h (as determined by an N-terminal assay) and were normal by 24–48 h (21).

These data suggest that PTH levels are likely subnormal or normal during lactation in humans; the original hypothesis of hyperparathyroidism in lactation has not been substantiated. The metabolism of lactating rats, on the other hand, may represent a functionally hyperparathyroid state, in response to a fall in maternal blood calcium during lactation.

4. 1,25-Dihydroxyvitamin D. In contrast to the high 1,25-dihydroxyvitamin D levels of pregnancy that contribute to the doubling of intestinal calcium absorption, within days of parturition, maternal free and bound 1,25-dihydroxyvitamin D levels fall to normal (20, 55, 58, 73, 160). The levels remain normal throughout lactation and postweaning (27, 58, 160, 397, 398, 410, 414, 417), although one study reported that the levels rose above the normal range when lactation continued beyond 6 months (414), and another found an elevation after weaning (398). Women nursing twins had higher 1,25-dihydroxyvitamin D levels than women nursing singletons (402). Serum 25-hydroxyvitamin D levels are typically unchanged by lactation (410).

In contrast to humans, lactating rats have elevated levels of 1,25-dihydroxyvitamin D (22, 81), and more intense lactation correlated with higher levels (22, 407). The lactational rise in 1,25-dihydroxyvitamin D seen in rats was prevented by a small increase in the calcium content of the diet and was exacerbated by a low-calcium diet (419). Parathyroidectomy in the lactating rat caused a 70% decrease in the level of 1,25-dihydroxyvitamin D (81). Therefore, 1,25-dihydroxyvitamin D levels are elevated in rats in response to a fall in blood ionized calcium and a rise in serum PTH.

5. *Calcitonin*. In lactating humans, high calcitonin levels that do not vary with suckling have been reported (14, 420), while normal levels 6 weeks to 6 months postpartum have been found by others (397, 402, 417). In these reports the serum calcium was not different between lactating women and controls. Nursing of twins resulted in higher calcitonin and corrected serum calcium levels compared with nursing of singletons, although the levels remained in the normal range (402). Calcitonin is secreted into breast milk at concentrations 45 times that of maternal plasma (93); its functional role within the breast is unknown. Since a rise in serum calcitonin during lactation persists in totally thyroidectomized women, the breast may be an important extrathyroidal source of calcitonin during lactation (93).

In sheep, despite constancy of serum calcium levels, calcitonin levels fall to nonpregnant levels at parturition, rise over the period of lactation, and fall again at weaning (324). A similar rise in serum calcitonin has been found in deer, which also manifest a mild increase in the corrected serum calcium during lactation (23). In contrast, the data from lactating rats are conflicting. Investigators have reported low calcitonin levels in the presence of mild lactational hypercalcemia (406), while others have found high calcitonin levels until weaning in the presence of a normal or low serum calcium (421).

In the "calcitonin-deficiency" model previously described (*Section II.B.5*), rats made calcitonin-deficient during pregnancy lost more mineral content of their femurs than normal after 3 weeks of lactation (96, 97). Similarly, calcitonin-deficient goats fed a calcium-deficient diet lost more bone mineral by day 60 of lactation than control animals (77). Lactating, thyroparathyroidectomized rats had a prompt (1.8 mg/ dl) rise in blood calcium after eating, while lactating control (intact) rats had no change in blood calcium (422). These findings indicate that in some species, during lactation, calcitonin protects the maternal skeleton from excessive resorption and regulates the maternal blood calcium level, particularly in response to meals. The relevance of these findings to humans has not been determined.

6. *PTHrP*. PTHrP has been detected in breast milk of humans and other animals at concentrations exceeding 10,000 times the level in the blood of patients with hypercalcemia of malignancy or normal human controls (110, 117). The primary role of PTHrP in the mammary glands or milk is not clear. A paracrine action of PTHrP in mammary tissue has been suggested because PTHrP concentrations in the milk are positively correlated with total milk calcium content of the human (423) and cow (424) [although no such correlations were found in rats (118, 425)], and administration of bromocriptine reduces both the PTHrP and calcium level in the milk of goats (426). However, a direct effect of PTHrP on the transport of calcium into the breast and breast milk has not been established. PTHrP has been found to have vasodilatory effects on mammary vessels and, therefore, may regulate mammary blood flow (427, 428). PTHrP has also been shown to have an essential role in mammary development (337).

PTHrP immunoreactivity in milk and PTHrP mRNA levels in mammary tissue have been observed to rise over the first few days postpartum in rats (118). Suckling induces PTHrP (mRNA and protein) locally in rat mammary glands (425, 429), and this response appears to be mediated by PRL and not oxytocin (429) and is blocked by bromocriptine (429). Milking a mammary gland in goats caused a marked increase in the PTHrP concentration in milk from that gland, but not in the milk of the contralateral (unstimulated) mammary gland (430). This result suggests that the synthesis and/or secretion of PTHrP by the mammary glands is under the control of local factors rather than the systemic level of PRL alone (430). Further, in rats, PTHrP is higher in milk at day 21 of lactation, after the PRL level has fallen, indicating that factors other than PRL may stimulate continued production of PTHrP by the mammary glands (431).

Studies in lactating rats have found that suckling induces phosphaturia and an increase in nephrogenous cAMP, which persists in parathyroidectomized, lactating rats (432). This result suggests that PTHrP (but not PTH) is released into the maternal circulation in physiologically relevant amounts in response to suckling. A similar study in cows found a milking-induced phosphaturia that could be blocked by (Tyr)³⁴bovine PTH(7–34)-NH₂, a PTH/PTHrP receptor antagonist (433). Further, a significant venous-arterial concentration gradient of PTHrP was found across the mammary gland of the goat (426), consistent with PTHrP reaching the maternal circulation from the mammary glands. However, chronic, passive immunization of the lactating mouse with antibodies to amino-terminal PTHrP, sufficient to eliminate PTHrP bioactivity in the breast tissue, failed to affect maternal calcium or phosphate levels (434). Thus, PTHrP is not the sole determinant of mineral homeostasis in lactating rats. Perhaps PTH, which is elevated in lactating rats (Section IV.B.3), can substitute for PTHrP in this circumstance.

Although one less sensitive single-site assay found normal levels of PTHrP in maternal blood during human lactation (110), studies that used a more sensitive RIA, or two-site IRMAs, have found PTHrP levels to be significantly higher than in nonpregnant controls (399–401, 411, 435). In addition, a small rise in the systemic level of PTHrP can be demonstrated after suckling (400, 436). PTHrP was not elevated in the first 3 days postpartum, at which time lactation is not fully established (437); PTHrP immunoreactivity rises steadily in the breast milk over the first few days postpartum and subsequently declines as lactation wanes (400, 431).

Amino-terminal fragments of PTHrP and PTH are equipotent in activating the human PTH/PTHrP receptor (104), and it is possible that fluctuations in secretion of PTHrP into the maternal circulation from the lactating breast cause resorption of calcium from the maternal skeleton, renal tubular resorption of calcium, and suppression of PTH. In support of this hypothesis, PTHrP levels have been found to correlate negatively with PTH levels and positively with the ionized calcium levels of lactating women (400, 401). Also, PTHrP

levels correlate with loss of bone mineral density during lactation in humans (435). Furthermore, aparathyroid or hypoparathyroid women have been found, while lactating, to be able to activate the 1 α -hydroxylase enzyme to synthesize low-to-normal levels of 1,25-dihydroxyvitamin D and maintain normal serum calcium levels while not receiving supplemental calcitriol (244, 245, 258). This is consistent with PTHrP reaching the maternal circulation in amounts sufficient to allow stimulation of 1,25-dihydroxyvitamin D synthesis and maintenance of normal (or slightly increased) maternal serum calcium. This consistent finding has resulted in the recommendation that calcitriol supplementation be reduced and the ionized calcium carefully followed in the postpartum period of hypoparathyroid women who plan to breast-feed (254).

PTH-independent hypercalcemia can occur during lactation and may be PTHrP-mediated. A 21-yr-old woman was reported to develop hypercalcemia and hypercalciuria during lactation in the presence of low PTH, low 1,25-dihydroxyvitamin D, and high PTHrP levels (438). These abnormalities resolved within 2 weeks of weaning, except that the elevated PTHrP level declined more gradually (438). A second woman had lactational hypercalcemia associated with low PTH levels and bone biopsy evidence of active cellular resorption, consistent with hyperparathyroidism (439). Another case of PTHrP-mediated hypercalcemia was reported to occur in a lactating woman with mammary hyperplasia and fully resolved only after a reduction mammoplasty was performed (440).

In summary, PTHrP is present at very high concentrations in breast milk and at higher than normal concentrations in the maternal circulation during lactation. PTHrP may affect maternal calcium metabolism, particularly by increasing skeletal resorption of calcium and phosphate, and renal tubular reabsorption of calcium. It may, thereby, be at least partially responsible for the slight increase in ionized calcium that occurs normally in lactating women and may contribute to the occasional occurrences of hypercalcemia and osteoporosis in lactation (*Section IV.E.3*, below).

C. Intestinal absorption of calcium

Intestinal absorption of calcium in lactating humans falls from the high levels of pregnancy to control levels (12, 28, 126–128, 412, 441), coinciding with the corresponding fall in 1,25-dihydroxyvitamin D levels to normal (Section IV.B.4, above). Even when women are assigned to different levels of dietary calcium intake, a lactational increase in efficiency of intestinal absorption of calcium cannot be demonstrated (412). Intestinal absorption of calcium does increase slightly in lactating women whose menses have resumed (412). In some women, dietary intake of calcium may be increased during lactation, such that the total amount of calcium absorbed is increased, even though the efficiency of intestinal absorption is not (412). However, other studies have indicated that such dietary calcium supplementation is of uncertain benefit, since it will increase the urine calcium excretion without affecting the calcium content of breast milk or maternal skeletal losses of calcium (160, 442, 443) (see discussion in Section IV.E.2, below). After weaning, there is

an increase in intestinal absorption of calcium (412), which may facilitate restoration of calcium to the maternal skeleton.

The adaptations in lactating rats differ from those in humans. These may reflect the effect of greater calcium demands due to the larger litter size and far shorter lactational period. Rats demonstrate a 2-fold increase in duodenal calcium absorption in everted gut sacs, similar to the levels found during pregnancy (141, 444); 1,25-dihydroxyvitamin D levels are also elevated (81). Other factors, in addition to vitamin D, may influence calcium absorption, since, even in the setting of vitamin D deficiency, lactating rats exhibited an increase in duodenal calcium absorption and raised their serum calcium (141, 444). This finding suggests that PRL or some other factor present during lactation might stimulate intestinal calcium absorption. The increased intestinal calcium absorption also persisted in the presence of ovariectomy on the postpartum day 2 (445). At weaning, rats normally experience a rebound hypercalcemia (81, 409), and this is still detected in the presence of severe vitamin D deficiency (143), consistent with an increase in intestinal absorption of calcium. However, this rebound hypercalcemia could also be explained by enhanced release of calcium from the skeleton due to active bone resorption. Despite the persistent increase in dietary absorption, rats appear to also mobilize calcium from the skeleton to meet the demands of lactation (Sections IV.E.1 and IV.E.2, below).

Therefore, in summary, intestinal calcium absorption is not increased during lactation in humans, despite calcium requirements similar to those in pregnancy. In contrast, intestinal absorption of calcium is increased in rodents throughout lactation by increases in 1,25-dihydroxyvitamin D and probably other unknown mechanisms.

D. Renal handling of calcium

In humans, the glomerular filtration rate falls from the elevated level of pregnancy, and renal excretion of calcium is typically reduced to levels as low as 50 mg/24 h (12, 14, 27, 28, 44, 128, 146, 410). The low urine calcium in the setting of high calcium in blood suggests that tubular reabsorption of calcium might be increased to account for the reduction in calcium excretion. The reduction in calcium excretion appears to persist after weaning during the period of restoration of bone density to the skeleton (410). Renal calcium excretion has been shown to be similarly reduced in the lactating rat (446).

E. Skeletal calcium metabolism

1. Bone formation and resorption. Bone turnover is increased during lactation in rats, as indicated by changes in histomorphometric parameters of bone (136, 156, 447). Beagle dogs also show histomorphometric evidence of increased bone turnover in iliac trabecular bone (155), proximal femur (448), and lumbar vertebrae (448) during lactation.

In humans, definitive histomorphometric data are lacking, and, therefore, serum and urine markers of bone formation and resorption have been used to assess bone turnover. Markers of bone resorption (tartrate-resistant acid phosphatase, deoxypyridinoline/creatinine, hydroxyproline/ creatinine) are elevated 2- to 3-fold during lactation and are higher than the levels attained in the third trimester (27, 28, 160, 161, 400, 410, 415, 449). Although markers of bone formation have been reported in a few studies to be generally normal (27, 162), most studies have found such markers to be high during lactation and increased over the levels attained during the third trimester (28, 160, 161, 400, 410, 415, 449). Total alkaline phosphatase falls immediately postpartum due to loss of the placental fraction, but may still remain above normal due to the elevation in the bone-specific fraction (161, 162).

Overall, the results of these studies in humans of markers of bone formation and resorption are consistent with the histomorphometric data from animal models, indicating that bone turnover is significantly increased during lactation.

2. Bone density. Within 21 days of parturition, lactating rats lose about 35% of bone mineral, primarily from trabecular sites (primarily femurs, tibias, and lumbar vertebrae), as determined by a variety of methods that determined total or trabecular ash weight, ash mineral content, and changes in bone mineral density (2, 50, 139, 140, 405, 447, 450). These losses can be increased to 43% or more of trabecular bone mineral by consumption of a low calcium diet beginning at parturition (50, 405, 408, 451, 452). These changes are sufficient to adversely affect the mechanical properties of bone (strength, stiffness, toughness, and ductility) (408).

The loss of skeletal calcium can be worsened by larger litter sizes (408). The loss is similar in vitamin D-deficient and vitamin D-replete rats (137), although another study found a 2-fold greater loss of calcified bone in vitamin D-deficient rats (154). Moreover, vitamin D-replete rats regain the lost mineral after weaning while vitamin D-deficient rats do not; thus vitamin D is needed to restore the lost mineral (137). The lactational loss of bone mineral also persists in the absence of the maternal parathyroid glands (453, 454) and is not affected by ovariectomy, adrenalectomy, or immediate pregnancy (455), unless the animals are simultaneously fed a low calcium (0.1%) diet (445). Therefore, estrogen (and perhaps estrogen deficiency), adrenal hormones, 1,25-dihydroxyvitamin D, and PTH are not needed to mobilize skeletal calcium during lactation. On the basis of these observations, Brommage and DeLuca (455) first proposed that the putative hypercalcemia of malignancy factor, later identified to be PTHrP, might be the mediator of skeletal bone resorption during lactation.

The earliest longitudinal study of bone mineral density during human lactation followed 10 women with serial measurements of the femoral shaft, using a ²⁴¹Am source, and found a mean loss of 2.2% of bone mineral content over 100 days of lactation (456). More recent studies have followed women with serial measurements of bone density during lactation (by SPA, DPA, or DXA), and a fall of 3–8.0% in bone mineral content has been reported after 2 to 6 months of lactation at trabecular sites (lumbar spine, hip, femur, and distal radius), with smaller losses at cortical sites (28, 146, 160, 166, 169, 400, 410, 415, 417, 449, 456–463); this subject has recently been reviewed in detail elsewhere (390). In contrast, bottle-feeding, postpartum controls do not lose bone density at the lumbar spine over the same interval and may show a

net gain in bone density (415, 417, 458, 462, 464). A 15% decrease in cortical bone density determined by SPA of the distal radius was found in teenaged mothers who consumed calcium and phosphate at levels below the recommended daily allowance. This result suggests that poor maternal nutrition might worsen the skeletal changes during lactation (459, 460). This was confirmed in a follow-up study, in which lactating adolescents who consumed calcium in excess of 1600 mg daily had no change in bone mineral density, compared with lactating adolescents who consumed 900 mg calcium daily and lost 10% of bone mineral content over the same interval (465). These teenaged mothers had not yet reached their peak bone mass, which might have been a factor in the responsiveness of the skeleton to dietary calcium supplementation.

Other evidence suggests that skeletal calcium will be preferentially resorbed and that supplementing the diet with calcium will not prevent resorption. For example, consumption of a calcium supplement by lactating Gambian women caused a sharp increase in urinary calcium excretion but had no effect on lactational bone mineral loss when compared with women who consumed less than the recommended daily allowance of calcium (442, 443). In a randomized clinical intervention trial that studied the effect of consuming dietary calcium in excess of the recommended daily allowance (2.4 g daily), lactating women still lost 6.3% of bone mineral density at the lumbar spine and up to 8% from the radius and ulna, as determined by DXA (160). Furthermore, in a preliminary report, the lactational decrease in lumbar spine bone mineral density was not influenced by maternal calcium intake but was negatively and significantly correlated with the breast milk output (466). Therefore, loss of bone mineral from the maternal skeleton appears to be a normal consequence of lactation and may not be preventable by raising the calcium intake above the recommended dietary allowance.

It is not clear whether this loss of bone mineral content is simply due to relative estrogen deficiency of lactation or a more complex, possibly humorally mediated, mechanism. Several studies have suggested that estrogen withdrawal and the intensity and duration of lactation are independent factors in determining the rate and magnitude of bone loss during lactation (218, 416, 462, 464, 467). For example, early resumption of menses (464) or an oral contraceptive (462) can reduce the skeletal losses, but bone density may continue to decrease during extended lactation, even after menses have resumed (218, 467). No published study has adequately addressed the relative role of estrogen withdrawal during lactation in a definitive way, since no study has thus far manipulated estrogen independently of lactation. However, it is evident from studying the effects of acute estrogen deficiency induced by GnRH agonist therapy in young women (given for such diverse conditions as endometriosis, uterine leiomyomata, and premenstrual syndrome) that estrogen deficiency alone is unlikely to account for all of the changes in skeletal calcium metabolism seen during lactation (Table 2). After 6 months of GnRH agonist therapy, the bone mineral content of trabecular-rich sites in the axial skeleton alone is affected and is typically reduced by only 2-4%, as determined by DXA (468–478). At the same time, the serum calcium and

TABLE 2. Comparison of the changes in bone density, calcitropic hormones, minerals, and markers of bone resorption that occur during 6 months of lactation and 6 months of GnRH agonist therapy

	Lactation	GnRH Agonist
Serum calcium	Increased	Increased
Serum phosphate	Increased	Increased
PTH	Decreased	Decreased
1,25-Dihydroxyvitamin D	Normal	Decreased
24-h Urine Ca	Decreased	Increased
Urinary Ca/Cr	Decreased	Increased
Urine HP/Cr	Increased	Increased
BMD changes (DXA)	3–8% at trabecular-rich sites; less at cortical	2–4% at trabecular-rich sites
Recovery of BMD at 6 months	? Complete	? Complete

BMD, Bone mineral density; Ca, calcium; Cr, creatinine, HP, hydroxyproline.

phosphate are increased (469-471, 473, 479), 24-h urine calcium excretion and urinary calcium/creatinine ratio are increased (469-473, 478, 479), but the levels of intact PTH and 1,25-dihydroxyvitamin D are low (469, 473, 479). Therefore, in lactation, the greater losses of bone mineral density (at both trabecular and cortical sites), the normal 1,25-dihydroxyvitamin D levels, and the reduced urinary calcium excretion may be due to the effects of other factors (such as PTHrP) in addition to the effects of estrogen withdrawal. In one recent study of lactating women, higher PTHrP levels were found to correlate with loss of bone mineral density at the lumbar spine and femoral neck, even after accounting for the effects of estradiol levels, PTH, and breast-feeding status (435). In this same study, the high PTHrP levels correlated with breast-feeding status, high PRL levels, and lower estradiol levels (435). Further support for a role of PTHrP in mediating the skeletal changes that occur during lactation comes from the finding that PTHrP is also elevated in the serum of patients with hyperprolactinemia due to pituitary adenomas (401, 480). In these patients, PTHrP correlated positively with the serum PRL and calcium and was negatively correlated with the serum PTH and bone density of the lumbar spine.

The bone density losses of lactation are substantially reversed during weaning, such that the maternal skeleton is able to provide for the calcium requirements of lactation with few, if any, long-term consequences (160, 410, 417, 467). Also, the losses are regained quickly enough that women who breast-feed for at least 6 months, but have a second pregnancy within 18 months, do not have lower bone density after the second pregnancy (463, 481). Only one study has thus far reported failure of the bone density to return to baseline (415). Compared with the studies of GnRH therapy, the longitudinal, prospective studies of lactating women were of insufficient power, however, to eliminate the possibility of modest failure to completely restore bone mass after weaning. The reversibility of bone density losses has also been found in the lactating rat model, where the loss of 35% of trabecular bone mineral content is completely restored at weaning (2, 140, 452).

In the long-term, the consequences of lactation-induced depletion of bone mineral content appear clinically unimportant. The vast majority of epidemiological studies of preand postmenopausal women have found that a history of lactation has no adverse effect (172, 174–181, 183, 188–191, 195, 197, 202, 482) or a protective effect (184, 187, 203, 483–487) on peak bone mass, bone density, or hip fracture risk. Only five studies suggest that a history of lactation correlates with reduced radial (196, 488–490) or lumbar (199) bone mineral content. Thus, there appears to be little or no long-term harm caused by the temporary demineralization of the skeleton during lactation.

A similar pattern of decreasing bone density of the lumbar spine during lactation with recovery at weaning has been observed in another primate species, the African green monkey (491). The bone mineral density of the lumbar spine in these monkeys has been observed to fall by about 12% from the baseline value at parturition during 20 weeks of lactation; during an additional 20 weeks of observation after weaning, the bone mineral density increased but did not fully recover to baseline. This model may more closely resemble the human condition, but has not yet been thoroughly studied.

In summary, bone mineral density decreases of 3–8% at trabecular sites occur in the first 6 months of lactation in humans. By comparison, lactating rats lose about 35%; less dramatic losses have been reported in African green monkeys. In all cases, the lost bone mineral content is largely restored after weaning, and there appears to be little or no long-term increased risk of fracture or decreased bone density in women who have lactated. The loss of bone mineral density during lactation may be due to the combined effects of relative estrogen deficiency and PTHrP-induced skeletal resorption; both of these factors may correlate with more intense and prolonged lactation.

3. Osteoporosis of lactation. Rarely, osteoporosis presents during lactation; like the osteoporosis of pregnancy, this may represent a coincidental, unrelated disease. Alternatively, it may represent a continuum of the condition that may present in pregnancy and an exacerbation of the normal degree of skeletal demineralization during lactation (208, 223, 438, 492). In some of these cases, low peak bone mass may have preceded lactation (and pregnancy). Typically these women present 1 to 6 months postpartum with vertebral crush fractures, bone pain, loss of height, and rarely hypercalcemia; the osteopenia may resolve subsequently (438, 492). Bone biopsies show features indistinguishable from normal (223) or may show evidence of increased resorption (492). When measured, PTH is typically normal or reduced, and 1,25-dihydroxyvitamin D is also normal (438, 492). As for the osteoporosis of pregnancy, the difficult diagnostic question remains: when did the condition start? Given the preceding discussion, the possibility arises that excessive PTHrP release from the lactating breast into the maternal circulation could cause excessive bone resorption, osteoporosis, and fractures in these cases (438, 493). PTHrP was high in one case of lactational osteoporosis and was found to remain elevated for months after weaning (438). However, the extent to which PTHrP contributes to the reduction of bone density during lactation has yet to be established.

F. Hypoparathyroidism and pseudohypoparathyroidism

Calcitriol requirements of hypoparathyroid women fall early in the postpartum period, especially if the woman breast-feeds, and hypercalcemia may occur if the calcitriol dosage is not substantially reduced (244, 245, 248, 253–255, 258, 494). As noted earlier in the discussion of hypoparathyroidism in pregnancy (*Section II.G*), this may result from the activation of the renal 1α -hydroxylase by PTHrP.

The management of pseudohypoparathyroidism has been less well documented. Since these patients are likely resistant to the renal actions of PTHrP, and the placental sources of 1,25-dihydroxyvitamin D are lost at parturition, the calcitriol requirements can be expected to return to prepregnancy levels and should be unchanged by lactation.

G. Summary

Even when dietary intake of calcium exceeds the recommended daily intake, the calcium demands of lactation in humans are preferentially met by increased skeletal resorption of calcium and probably increased renal conservation of calcium, but not by increased intestinal absorption of calcium. Serum calcium is slightly increased or normal, while phosphate is high normal or frankly elevated. These increases reflect calcium and phosphate entering the circulation from bone in increased amounts and reabsorption of calcium and phosphate by the kidney. PTHrP levels are extremely high in breast milk and increased in the maternal circulation, while PTH levels are generally low during lactation. 1,25-Dihydroxyvitamin D levels are typically normal throughout lactation but may be higher in women nursing twins. Markers of bone resorption and formation are increased, and bone density has been found to reversibly decrease during lactation. These lactational decreases in bone density may not adversely affect the skeleton in the longterm, although occasionally the normal lactation-induced decrease in bone density may be excessive, leading to fractures and a clinical diagnosis of osteoporosis.

Increasing the dietary intake of calcium does not consistently prevent the loss of skeletal calcium from occurring. Relative estrogen deficiency and elevation in PTHrP levels may both play a role in the lactational loss of skeletal calcium. PTH and 1,25-dihydroxyvitamin D elevations occurring after weaning may indicate a role for these hormones in providing the mineral needed to restore the skeleton.

In contrast to the findings in humans, skeletal losses of calcium are far greater in lactating rats, and hypocalcemia may ensue with more intense lactation. In that setting, the dietary absorption of calcium is increased, and both PTH and 1,25-dihydroxyvitamin D levels are elevated. These striking differences may limit the usefulness of the lactating rat as a model for understanding the human condition (Table 3).

Although critical experimental data are missing, a tentative hypothesis for explaining calcium physiology in lactation starts with the dominant role of PRL to suppress gonadotropins (and, therefore, estrogen) and to stimulate PTHrP synthesis. Estrogen withdrawal then leads to bone resorption. If PTHrP were not available, the bone resorption would be expected to suppress PTH levels, which would cause loss of calcium in the urine, decrease in 1,25-dihydroxyvitamin D levels and intestinal calcium absorption, and diminution of net bone resorption (*i.e.*, similar to the changes seen in GnRH therapy, Fig. 3). Such actions would act against the provision of calcium for milk production. In this context, the production of PTHrP can be viewed as the provision of a PTH surrogate, but one that is not under negative feedback control by calcium. Thus, even with low levels of PTH, 1,25-dihydroxyvitamin D levels are then maintained, urinary calcium is reabsorbed, and bone resorption continues (Fig. 3).

In this formulation, the bone resorption initiated by estrogen withdrawal is not a disease, but an intelligent way to assure a supply of calcium for milk, independent of dietary vagaries. At weaning, young bones are apparently able to restore bone mass by as yet uncertain mechanisms. In this context, menopausal osteoporosis can be viewed as an evolutionary accident — a property of estrogen withdrawal that is constructive during the reproductive years, but destructive in the setting of older bones and prolonged estrogen withdrawal.

V. Neonatal Physiology and Pathophysiology

A. Neonatal adaptive goals

As described earlier, the fetus maintains a higher blood calcium than the mother. This level of blood calcium appears to be dependent on the input of calcium across the placenta, an intact PTH/PTHrP receptor, and fetal PTHrP to maintain the fetal blood calcium and regulate placental calcium transport. After the umbilical cord has been cut and the placental calcium infusion (and placental sources of PTHrP) abruptly lost, the neonate becomes dependent on intestinal calcium intake and skeletal calcium stores to maintain a normal blood calcium at a time of continued skeletal growth. Further, the kidney, which postnatally produces urine unavailable for reingestion, takes on a major role in maintaining calcium and, particularly, phosphate homeostasis. If intestinal calcium intake is inadequate, continued growth and mineralization of the skeleton will be compromised. In adjusting to the loss of the placental calcium pump, PTH and 1,25-dihydroxyvitamin D become more important, while PTHrP appears to be less involved in neonatal calcium homeostasis. Therefore, the adaptive goals of the neonate are to quickly turn on PTH and 1,25-dihydroxyvitamin D synthesis, which,

TABLE 3. Important differences between calcium physiology of humans and rodents during lactation

Factor	Human lactation	Rat lactation
Blood ionized calcium PTH 1,25-D Intestinal calcium	Stable or slightly increased Low to low-normal Normal Normal	Reduced Increased Increased Increased
absorption Skeletal calcium losses	3-8%	30-35%

1,25-D, 1,25-Dihydroxyvitamin D.



FIG. 3. Schematic illustration of the responses to acute estrogen deficiency alone (e.g., GnRH analog therapy) vs. lactation. Acute estrogen deficiency increases skeletal resorption and raises the blood calcium; in turn, PTH is suppressed and renal calcium losses are increased. During lactation, the combined effects of PTHrP (secreted by the breast) and estrogen deficiency increase skeletal resorption and raise the blood calcium, but calcium is directed into breast milk. Although PTH is suppressed during lactation, renal calcium losses are lower than normal.

in turn, up-regulate intestinal calcium absorption, and regulate skeletal and renal handling of calcium and phosphate.

B. Mineral ions and calcitropic hormones

The changes that occur in neonatal calcium, phosphate, and calcitropic hormone levels over the first 4 days after birth are schematically depicted in Fig. 4.

1. *Calcium.* In rodents, ionized calcium and total calcium fall to about 60% of the high fetal values within 6–12 h of birth (269, 278). Thereafter, the ionized calcium rises over the succeeding 12–36 h to about 80% of the fetal value, and then slowly falls over the following week to normal adult levels (276). In contrast, after reaching a nadir at 6–12 h after birth, the total calcium gradually rises to adult levels over the succeeding 7–14 days (269, 276). The total calcium changes largely reflect alterations in the serum albumin, such that, by the end of the first 2 weeks of life, the albumin-bound fraction is approximately 50% of the total calcium (in fetal life it accounts for only about 20%) (276). Neonatal lambs differ in that the blood calcium may remain at the higher fetal value over the first several months of postnatal life (330).

Although data from umbilical cord and neonatal blood levels are less complete in humans, the progression in ionized and total calcium values appears to be similar to that of rats (268, 270, 495). The ionized calcium in normal neonates has been reported to fall from the umbilical cord level of 1.45 mmol/liter to a mean of 1.20 mmol/liter by 24 h after birth (496). Babies delivered by elective cesarean section were found to have lower blood calcium and higher PTH levels at birth compared with babies delivered by spontaneous vaginal delivery (387). This finding suggests that the mode of delivery can affect early neonatal calcium homeostasis, a variable that has not been controlled in most studies of neonatal calcium physiology.



FIG. 4. Schematic illustration of the longitudinal changes in calcium, phosphate, and calcitropic hormone levels that occur during the neonatal period in humans. Normal adult ranges are indicated by the *shaded areas*. Data have been compiled from the following sources: total calcium (268, 270, 495), ionized calcium (268, 270, 495, 496), phosphate (32, 270, 273), PTH (17, 43, 110, 124, 268, 272, 305–307, 387, 495, 497–501), 1,25-dihydroxyvitamin D (11, 37, 40, 56, 304, 321), calcitonin (11, 86, 272, 500), and PTHrP (43, 110, 124, 310). The progression in PTHrP levels has been depicted by a *dashed line* to reflect that it is speculative.

2. *Phosphate*. Phosphate initially rises over the first 24 h of postnatal life in humans (270) and then gradually declines. The rise in phosphate corresponds to the early postnatal period when the parathyroid glands are still suppressed; phosphate declines as PTH secretion increases (see *Section*, *V.B.3*).

3. PTH. As discussed earlier (Section III.B.3), in fetal life PTH is present at low levels in the circulation and is apparently synthesized at low levels by the parathyroid glands. Based on measurements taken only at birth and 24 h of age in humans, the intact PTH level has been found to rise briskly after birth to within or near the normal adult range (17, 387, 497-501). However, these studies did not ascertain how soon the PTH level begins to rise postnatally and whether the peak level is attained by 24 h or even later. A more detailed time course for PTH was obtained using earlier generation PTH assays; as discussed earlier, these assays were less reliable, and measured fragments that accumulated in the circulation much more slowly than intact PTH. With these caveats in mind, data from these older assays found that PTH levels remained low over the first 12-24 h in humans and did not reach peak levels until 48 h or later (268, 272, 305-307).

Regardless of the exact time course, it is apparent that the increase in PTH levels follows the early postnatal drop in the serum ionized calcium and precedes the subsequent rise in ionized calcium (268, 272, 306, 495) and 1,25-dihydroxyvitamin D (Section V.B.4, below). The fall in ionized calcium is probably a consequence of the parathyroid gland suppression seen in late gestation (Section III.B.3, above) combined with the sudden loss of placental calcium delivery; the subsequent rise in PTH and the ionized calcium represents the progressive recovery from this suppression. During the first 48 h, the parathyroid glands have been found to respond sluggishly to more severe falls in the ionized calcium, such as that caused by exchange transfusion with citrated blood (268, 272, 306, 502). The degree of responsiveness to acute hypocalcemia appears to increase with postnatal age (306, 502); however, the interpretation of these data are limited by the nature of the older PTH assays from which they were obtained.

4. 1,25-Dihydroxyvitamin D. In humans, 1,25-dihydroxyvitamin D rises to adult levels over the first 48 h of postnatal life, likely in response to the rise in PTH (11, 40). Phosphate levels are high at this time and would tend to inhibit a rise in 1,25-dihydroxyvitamin D. Although in humans, rats, sheep, and pigs the fetal blood calcium and skeletal mineralization are unaffected by vitamin D deficiency, in the neonatal period deficiency of vitamin D can become manifest (29, 79, 84, 284, 285). This is further discussed in *Section V.E*, below.

5. *Calcitonin.* Data from humans have found a rise in the serum calcitonin, 2- to 10-fold over cord blood levels, over the first 24–48 h of life (11, 272, 500). Infants that are premature, asphyxiated, or hypocalcemic have the highest postnatal calcitonin levels (272); consequently, it has been suggested that hypercalcitoninemia may cause neonatal hypocalcemia (86). Consistent with this hypothesis, hypocalcemic preterm infants had higher calcitonin levels than normocalcemic, preterm control infants (503). However, the same study found

that the calcitonin levels of hypocalcemic term infants did not differ from those of normocalcemic term infants (503). Therefore, the influence of calcitonin on neonatal calcium homeostasis may vary with gestational age at birth; or, alternatively, have little role in calcium homeostasis. The physiological effect of calcitonin administration to neonates (human or animal) has not been directly tested.

Further evidence suggests that calcitonin levels are not significantly regulated by calcium, or responsive to the serum calcium level, in the neonatal period. In infants of normal and diabetic pregnancies, calcitonin levels increased after birth irrespective of the rate of fall in serum calcium, indicating that the postnatal surge in serum calcitonin may not be the main mechanism of postnatal hypocalcemia (500). Similarly, in very-low-birth-weight, preterm infants, spontaneous hypocalcemia provoked a rise in PTH but no change in calcitonin levels, and a calcium infusion to treat the hypocalcemia reduced the PTH level but did not affect the calcitonin level (504).

After the early surge in calcitonin levels over the first 48 h of life, calcitonin gradually falls to adult levels over the first month after birth in sheep (324) and humans (86).

6. PTHrP. PTHrP secretion from placenta, amnion, and umbilical cord is lost at birth; secretion from the parathyroid glands is also apparently lost, since PTHrP circulates at low to undetectable levels (with currently available assays) during normal adult life in humans and animals (104, 116, 335, 493, 505). The loss of placental sources of PTHrP, in addition to loss of the placental calcium pump, may be one of the reasons that the ionized and total calcium levels fall dramatically at birth. However, a preliminary report in neonatal (1- to 3 day-old) rat pups has found persistently high PTHrP levels, in the range of 20-40 pmol/liter (506). Studies in neonatal lambs have shown a progressive decline in PTHrP immunoreactivity in the parathyroid glands, and serum bioactivity attributable to PTHrP, over the first several months after birth, corresponding to the delayed fall in blood calcium that occurs in this species (116, 330). As noted earlier (Section III.C), indirect evidence from fetal sheep suggests that the fetal parathyroid gland may be an important source of PTHrP for normal regulation of the blood calcium and placental calcium transport. However, it is not known whether PTHrP synthesis occurs in the parathyroid glands of neonatal humans, or is turned off at some point in postnatal development.

Although immunoreactive PTHrP in milk can be absorbed into the bloodstream of suckling neonates (and might account for high levels of PTHrP in neonatal plasma) (113, 507), PTHrP may not have a dramatic effect on neonatal mineral homeostasis, since blocking its activity in milk by passive immunization of the mother had no dramatic effect on mouse pup blood calcium, weight gain, or femur mass (434). These negative results, however, might result from compensatory changes in neonatal PTH, calcitonin, and 1,25-dihydroxyvitamin D, which were not measured in this study. Passive immunization of the 2-day-old neonatal mouse with antibodies to amino-terminal PTHrP did not affect neonatal calcium homeostasis (508), but again, compensations may have obscured the role of PTHrP. These findings are consistent

Vol. 18, No. 6

with the observations described above from humans and rats, that PTH dominates the regulation of calcium homeostasis by 48 h of postnatal life. However, since there are no human data, and limited animal data on PTHrP levels in the neonatal period, an important role for PTHrP in calcium homeostasis after birth has not been ruled out.

C. Intestinal absorption of calcium

Calcium is absorbed in the gastrointestinal tract by both active and passive mechanisms (125, 509). Active intestinal transport occurs mainly in the duodenum and is regulated by 1,25-dihydroxyvitamin D, partly through its stimulation of calbindin_{9K}-D. Passive transport of calcium appears to be an unregulated, nonsaturable process that occurs throughout the small intestine; the rate of passive transfer of calcium into intestinal cells is directly proportional to the intraluminal concentration of calcium.

In newborn rat pups, intestinal calcium absorption is largely a passive, nonsaturable process that is not dependent on vitamin D (510-512). The high lactose content of milk has been shown to specifically increase the efficiency of intestinal calcium absorption, and net bioavailability of dietary calcium, through effects on paracellular diffusion in the distal small bowel (513-515). As the pups mature, receptors for 1,25-dihydroxyvitamin D begin to appear in intestinal cells (516), and the mucosal levels of calbindin_{9K}-D increase sharply (517). Around the same time, vitamin D-dependent active transport of calcium becomes noticeable (512), while passive transfer of calcium into the intestinal cells declines (510, 511). By the time of weaning, the intestine is less permeable to passive absorption of calcium, and active transport has become the dominant means by which calcium is transferred into the intestinal mucosa (510-512).

Data from newborn humans is less complete; therefore, the timing of the development of vitamin D-dependent calcium transport in humans is not known. However, normal term and preterm infants exhibit a similar postnatal increase in the efficiency of intestinal calcium absorption (518–520). Studies from preterm infants indicate that passive, non-vitamin Ddependent absorption of calcium may be the dominant route of calcium transfer (518, 521). In addition, the lactose content of breast milk has been shown to increase the efficiency of intestinal calcium absorption in human infants (522, 523). Postnatal supplementation of the preterm infant (mean gestational age, 32 weeks) with vitamin D markedly increased the absorption of calcium by 2-4 weeks after birth, as compared with similar preterm infants that were not supplemented (520). However, the intestinal absorption of vitamin D has been shown to undergo a postnatal age-dependent increase (524); this may limit the efficacy of vitamin D supplementation in the newborn.

In summary, data from newborn rats and humans indicate that intestinal calcium absorption is largely a passive, nonvitamin D-dependent process until near the time of weaning. The high lactose content of milk increases the efficiency of passive absorption of calcium. The normal postnatal maturation of the neonatal intestine may affect the ability of preterm humans to accrete sufficient calcium for skeletal mineralization and to regulate their blood calcium.

D. Renal handling of calcium

Little data are available from animal or human studies on the renal handling of calcium in the first few days of postnatal life (509). In humans, urinary excretion of calcium is low over the first few days of life and rises over the succeeding 2 weeks (498, 525, 526). This rise in renal calcium excretion probably reflects the concurrent 2-fold rise in glomerular filtration rate over the first 2 weeks after birth (525, 527). It may also reflect a greater filtered load of calcium as a consequence of the gradually rising serum calcium, 1,25-dihydroxyvitamin D, and PTH levels and gradually declining serum calcitonin (Section V.B, above). The responsiveness of the renal proximal tubules to PTH may increase over the same time period. In addition to evidence from animal studies presented earlier (Section III.D), data in preterm and term human infants indicate that the renal tubules are responsive to exogenously administered PTH, as evidenced by a rise in urinary cAMP (528, 529); further, this response to exogenous PTH increases with postnatal age (277, 528, 529).

E. Skeletal calcium metabolism

In humans that receive adequate dietary calcium and phosphate, the neonatal skeleton continues to accrete calcium at a rate of about 150 mg/kg per day, similar to the rate of the late-term fetus (5, 530–532). Having abruptly lost the placental calcium pump, the neonate becomes completely dependent on intestinal intake of calcium and phosphate to continue to meet the demands of the developing skeleton. In turn, the blood calcium will be maintained by calcium obtained from skeletal stores, absorbed by the intestine, and reabsorbed by the kidneys. The effects of vitamin D deficiency during gestation become manifest during the neonatal period because intestinal calcium transport becomes necessary for supply of calcium to the skeleton. As noted earlier, from studies of maternal vitamin D deficiency in rats (29, 79, 84), pigs (79, 285), and sheep (284), the blood calcium is typically normal at birth and the skeleton is fully mineralized, but after birth the neonate will begin to manifest the complications of vitamin D deficiency. In vitamin D-deficient rats and pigs, a slight decrease in blood calcium was detected by 3 days after birth, but it was only after 14 days of neonatal life that absence of 1,25-dihydroxyvitamin D resulted in significant, but still modest, hypocalcemia (84, 285, 311). Thereafter, rat pups failed to thrive and manifested rachitic skeletal growth plates and decreased longitudinal growth (84, 285). These effects became more pronounced after weaning (84, 285). Further, in vitamin D receptor knockout mice, hypocalcemia and rachitic changes in the skeletons developed only after the first three postnatal weeks (313, 314).

The development of vitamin D dependence is similar for humans. In human vitamin D deficiency, hypocalcemia appears late in the first or second week of life, and rickets develops after 2–3 months (316, 317). Vitamin D supplementation during pregnancy is associated with higher neonatal serum calcium levels and a reduced incidence of neonatal hypocalcemia (83, 533, 534). Neonates with rachitic changes in the skull (craniotabes) have lower 25-hydroxyvitamin D levels than those without craniotabes (535), and fewer neonates develop craniotabes if the mothers received a vitamin D supplement during pregnancy (534). Infants in China with lower cord blood 25-hydroxyvitamin D levels were found to have fewer wrist ossification centers at 3–5 days of postnatal age (536); furthermore, the crown-heel lengths of infants born to vitamin D-insufficient Pakistani women were found to correlate positively with the maternal serum calcium and negatively with the maternal PTH level (537). These findings suggest that fetal vitamin D deficiency may be manifest earlier in postnatal life than previously believed (and perhaps even during fetal life) or may be a marker for more global nutritional deficiency.

Although parathyroidectomy of rats at birth results in hypocalcemia and hyperphosphatemia (538, 539), by the time of weaning the neonatal rats still have normal body weight, normal shape and trabecular content of developing long bones, and normal calcium and phosphorus content of bone ash (539). These observations are similar to the finding of normal skeletons in experimental models of vitamin D deficiency, and in the vitamin D receptor knockout mice. These findings suggest that factors other than PTH and vitamin D may be required for normal accretion of calcium by the skeleton in the first several weeks after birth, while the infant is nursing. The lactose content of milk (discussed in *Section V.C*) and the high levels of PTHrP in milk may be important.

Premature infants are prone to develop metabolic bone disease of prematurity, a form of rickets precipitated by loss of the placental calcium pump at a time when the skeleton is accreting calcium at a peak rate (315, 316). These premature infants are born with a bone mineral content appropriate for gestation age (as assessed by SPA of the radius), but if untreated, their radial bone mineral content fails to increase as appropriate for their gestational age (540–544). By 2–3 months of age, the physical and radiological signs of rickets of prematurity may develop. These include craniotabes, rachitic rosary, chest deformity, osteopenia, pathological fractures, metaphyseal stippling, and flaring and widening of the epiphyses of long bones (315).

Rickets of prematurity is not due to vitamin D deficiency. 25-Hydroxyvitamin D and 1,25-dihydroxyvitamin D levels are typically normal, and vitamin D supplementation of the mother during pregnancy, or the infant postnatally, does not prevent its development (315, 545, 546). Rickets of prematurity appears to be the consequence of inadequate calcium and phosphate intake to meet the demands of the mineralizing neonatal skeleton. Special oral or parenteral formulas that are high in calcium and phosphorus content will correct the demineralization process and allow normal skeletal accretion of these minerals (315, 316, 544, 545, 547).

In summary, continued mineralization of the neonatal skeleton is dependent on adequate vitamin D stores and intact intestinal calcium absorption. Preterm infants are prone to develop a form of metabolic bone disease if the immature intestine cannot absorb calcium efficiently enough to make up for the loss of the placental calcium pump.

F. Neonatal response to maternal hyper- or hypoparathyroidism

Although the fetal blood calcium is set independently of the maternal level in utero, and PTH does not cross the placenta, it is clear from more than 100 reported cases in humans that maternal hyperparathyroidism adversely affects the neonate (11, 225, 227, 229, 235, 237). Premature labor and stillbirth may result from unrecognized maternal hyperparathyroidism (225, 235). Typically, the parathyroid glands remain suppressed after birth, and complications of neonatal hypocalcemia, tetany, permanent childhood hypoparathyroidism, and even death may result. The mechanism of the prolonged suppression is not known, but is probably due to increased flux of calcium across the placenta when the mother is hypercalcemic (233). However, this hypothesis has not been tested experimentally. A similar suppression has also been observed in normal infants of women with hypercalcemia due to familial hypocalciuric hypercalcemia, who typically have similar elevations in the serum calcium without an increase in serum PTH level (241-243, 374).

Similarly, maternal hypoparathyroidism in humans has been associated in the neonate with parathyroid gland hyperplasia, generalized skeletal demineralization, subperiosteal bone resorption, bowing of the limbs, fractures of ribs and long bones, and low birth weight (246, 375-378). Stillbirth and neonatal death have also been associated with this condition (379-381). The serum calcium level of the neonate has been reported to be normal in most cases, while the PTH level (older assays) has been found to be elevated (263, 376, 548). The skeletal findings generally resolve over the first several months after birth, but acute interventions may be required to raise or lower the blood calcium in the neonate. In addition, subtotal parathyroidectomy, with or without parathyroid autotransplantation and cryopreservation, may be required to control more severe, autonomous disease (549).

Maternal hypocalcemia of any cause may result in parathyroid gland hyperplasia and hyperparathyroidism in the fetus and neonate (376, 382, 383). In women with pseudohypoparathyroidism, children who do not inherit the genetic disorder are usually normal at birth, although transient neonatal hyperparathyroidism has been reported in some cases (262, 263). Furthermore, children who did inherit the condition may also be normal at birth but may gradually develop the full biochemical features of pseudohypoparathyroidism over the first several years of life (550).

G. Neonatal hypocalcemia

Neonatal hypocalcemia typically presents as seizures, starting between 4–28 days of age (551, 552). The preterm infant is particularly prone to hypocalcemia, having lost the placental calcium pump at a time when the skeleton is rapidly accreting calcium, and the intestinal calcium absorption mechanism is relatively immature. In addition to prematurity, other causes of neonatal hypocalcemia include congenital hypoparathyroidism, magnesium deficiency, maternal diabetes, vitamin D deficiency or resistance, and hyperphosphatemia (Table 4) (552). In many cases the etiology of the

TABLE 4. Causes of neonatal hypocalcemia

Prematurity
Maternal diabetes
Congenital hypoparathyroidism
Maternal hypercalcemia
Magnesium deficiency
PTH resistance
Maternal vitamin D deficiency
Resistance to vitamin D
Anticonvulsants
Hyperphosphatemia
Citrated blood transfusion
Phototherapy
Respiratory alkalosis
Alkali therapy

hypocalcemia is unknown. One study reported that three hypocalcemic, otherwise unremarkable, neonates had high PTH and serum phosphate levels and a subnormal phosphaturic response to PTH infusion; this "neonatal pseudohypoparathyroidism" completely resolved by 6 months of age (551). In addition, the higher phosphate content of infant formula has been associated with increased serum phosphate and decreased serum ionized calcium levels in formula-fed infants, as compared with breast-fed neonates (553).

When older PTH assays were used, preterm infants, infants of diabetic pregnancies, and hypocalcemic neonates were typically found to have even lower or undetectable levels of PTH than normal neonates and took a day or two longer to manifest the rise in PTH (272, 305, 554). This was interpreted to indicate that in preterm infants, and in infants of diabetic pregnancies, the parathyroid glands are less able to regulate the blood calcium and prevent hypocalcemia. However, more recent studies have failed to confirm the earlier impression that parathyroid function is abnormal in preterm infants (17, 504).

Neonatal hypocalcemia can occur as a complication of maternal diabetes in pregnancy in up to 50% of cases (555, 556), although tight control of the maternal glucose during pregnancy reduces the incidence of neonatal hypocalcemia (557). The cause of hypocalcemia in these infants is likely to be multifactorial. As noted above, parathyroid gland secretion may be blunted during the first few days of life in both normocalcemic and hypocalcemic infants of diabetic pregnancies (272, 554, 558). Neonatal hypomagnesemia, which results from maternal wasting of magnesium in association with glycosuria during pregnancy, correlates with the severity of neonatal hypocalcemia and may be the major factor contributing to the sluggish neonatal parathyroid function (557, 559, 560). However, this hypothesis has not been tested by correcting the hypocalcemia through magnesium replacement alone. Although occult vitamin D deficiency has been suggested as a cause of hypocalcemia in these infants (561), supplementation of infants of diabetic pregnancies with vitamin D at 2, 24, 48, and 120 h after birth did not reduce the magnitude or incidence of hypocalcemia as compared with infants that were not supplemented (562).

In addition to the blunted parathyroid function and hypomagnesemia, it is also possible that alterations in maternal calcium homeostasis due to diabetes might predispose the neonate to become hypocalcemic. Although two cross-sectional studies in humans found no effect of maternal diabetes on maternal levels of calcium, phosphate, PTH, and calcitonin (563, 564), a longitudinal study found that pregnant diabetic women had lower 1,25-dihydroxyvitamin D, total serum calcium, and ionized calcium levels in the third trimester, as compared with nondiabetic pregnant women (565). Furthermore, experimental models of diabetes in pregnant rats demonstrate marked maternal hypercalciuria (566), reduced maternal-fetal transfer of calcium and magnesium (567), a 12-fold reduction in placental calbindin_{9K}-D levels (567), and reduced calcium content of fetal ash (567). Collectively, the human and animal data indicate that maternal diabetes can affect maternal calcium homeostasis and reduce the placental transfer of calcium. However, whether these effects predispose to the development of hypocalcemia in infants of diabetic pregnancies has not been determined.

H. Summary

In the early neonatal period, the neonate is challenged by the loss of the placental calcium pump and manifests a quick transition, from an environment in which PTHrP plays an important role to a PTH- and 1,25-dihydroxyvitamin D-controlled neonatal milieu. This is reflected in a rapid fall in total and ionized calcium over the first 6 h of life. The calcium level gradually corrects over the following 48 h, after PTH secretion by the parathyroid gland increases, and 1,25-dihydroxyvitamin D levels ascend to adult values. Serum phosphate persists at high levels until the rising PTH levels and increasing renal responsiveness to PTH permit a phosphaturia. Calcitonin levels may remain elevated for several weeks; the physiological importance of this elevation is uncertain. These changes in calcium and calcitropic hormone levels are summarized in Fig. 4. In contrast to the rapid changes in calcitropic hormone levels, intestinal calcium absorption changes gradually from a passive to an active, 1,25dihydroxyvitamin D-mediated process over the first weeks of life.

Although human and animal fetuses develop remarkably normally in the presence of maternal calcium, PTH, and vitamin D deficiency, the resulting neonates demonstrate abnormalities that are consequences of the prior abnormal maternal calcium homeostasis. Maternal hyperparathyroidism and hypoparathyroidism during pregnancy can be manifest as disturbances of neonatal calcium and bone metabolism. In addition, maternal diabetes during pregnancy can predispose to neonatal hypocalcemia, probably as a consequence of fetal hypomagnesemia induced by maternal renal wasting of magnesium.

When data from fetal and neonatal humans and animals are compared, it is apparent that PTHrP has a significant role in fetal calcium homeostasis and circulates at higher levels than PTH. In contrast, there is as yet no evidence of an important role for PTHrP in normal postnatal calcium homeostasis. PTHrP is normally found at low to undetectable levels in the adult, but the postnatal time point at which PTHrP is lost is not known.

VI. Discussion and Conclusions

Maternal adaptations of calcium homeostasis differ between pregnancy and lactation (Fig. 5). The pregnant woman manifests a 2-fold increase in intestinal calcium absorption that is mediated partly by a 2-fold increase in free and total 1,25-dihydroxyvitamin D levels and partly by mechanisms that are independent of vitamin D. In addition, the increased dietary intake of calcium is offset by increased renal losses of calcium. Skeletal stores of calcium do not appear to be mobilized to any significant degree during pregnancy. PTH levels are low or low-normal for much of pregnancy, and thus the concept of "physiological hyperparathyroidism of pregnancy" is invalid. The increased intestinal calcium absorption results in a gradual, slight increase in the corrected serum calcium and ionized calcium and a marked increase in renal calcium excretion (absorptive hypercalciuria). Bone mineral stores may be increased early in pregnancy in anticipation of the peak fetal demand of the third trimester; by term, the maternal skeleton has no apparent deficit of bone mineral. In contrast, the lactating woman does not increase intestinal calcium absorption, but increases bone turnover and renal tubular reabsorption of calcium to provide adequate calcium for the breast milk. Again, the serum ionized calcium is increased slightly, while the serum phosphate may be frankly elevated. These increases reflect the increased skeletal resorption and decreased renal excretion of these minerals. The lactation-associated fall in estrogen levels, along with secretion of PTHrP, leads to bone resorption. This, combined with PTHrP-mediated calcium reabsorption from the urine, leads to suppression of PTH. Lactation causes a loss of 3-8% of bone mineral content that is restored after weaning; this reversible loss of bone mineral does not appear to adversely affect the skeleton in the long term.

Estrogen deficiency due to menopause is associated with significant, essentially irreversible losses of calcium from the aging skeleton; these losses cause osteoporosis and increase the risk of fractures. During lactation, the effect of estrogen deficiency, in association with PTHrP, increases skeletal resorption to provide calcium for the milk; the younger skeleton appears capable of restoring calcium losses after weaning. In this sense, the occurrence of osteoporosis at menopause may be regarded as the unfortunate consequence of outliving normal ovarian function and inducing permanent estrogen deficiency without the normal bone-restorative factors that are also present during lactation and weaning.

The regulatory mechanisms that direct the adaptive processes that occur in pregnancy and lactation are by no means fully elucidated. The potential roles of reproductive hormones of pregnancy in calcium homeostasis (estrogen, PRL, CG, placental lactogen, etc.) have not been adequately explored, and it is also possible that other calcitropic factors, perhaps specific to pregnancy and lactation, remain to be identified. The results of studies from animal models of pregnancy and lactation must be interpreted carefully, given that there are significant differences between the adaptive strategies for calcium and bone homeostasis seen in humans, rodents, and other animals (Tables 1 and 3). For example, the pregnant and lactating rat both develop secondary hyperparathyroidism, and the lactating rat can lose up to 35% of skeletal calcium before restoring it after weaning. The lactating rat also increases the intestinal absorption of calcium, whereas lactating women do not.

The rare disorders of osteoporosis in pregnancy and lactation may represent chance occurrences of idiopathic osteoporosis, or they may represent a spectrum of one common condition whose time of presentation is determined by the prepregnancy bone mass and the rate of bone resorption subsequent to conception. It seems likely that estrogen deficiency combines with the actions of PTHrP to stimulate the loss of calcium from the skeleton during lactation. The mechanism by which calcium is restored to the skeleton after weaning is not known, but if understood, might be adapted to a bone-restorative therapy for osteoporosis. The postpartum restoration of normal estrogen levels (recognized by the resumption of normal menses) is clearly an important factor,



but probably not the only one, since part of the restoration of calcium to the skeleton occurs only with weaning.

Fetal calcium homeostasis in late gestation appears to be largely regulated by PTHrP, which stimulates placental calcium transport, resorbs bone, and may stimulate renal calcium reabsorption. PTH can also resorb bone, stimulate renal calcium reabsorption, and stimulate synthesis of 1,25-dihydroxyvitamin D. However, since PTH and 1,25-dihydroxyvitamin D are found at very low levels in the fetal circulation during late gestation, their roles in fetal life are less well defined. The relative importance of PTH and PTHrP earlier in fetal life are unknown. The fetus sets its blood calcium level irrespective of the ambient maternal blood calcium level. The usefulness to the fetus of a blood calcium higher than the mother's is not understood, as it does not appear to be necessary for full mineralization of the skeleton to occur. It is possible that the high fetal calcium levels act as a safety margin at birth, allowing the newborn to experience a postnatal fall in ionized calcium without tetany or convulsions. Alternatively, a higher ionized calcium may be useful for cellular functioning under fetal conditions (low pO₂, low pH, for example) that differ from those in later life. In any case, the fetus is able to maintain a normal blood calcium and fully mineralize its skeleton in the setting of significant maternal calcium and vitamin D deficiencies.

A changeover to a PTH- and 1,25-dihydroxyvitamin Ddriven environment occurs during the first 48 h of postnatal life, accompanied by loss of hypercalcemia and high PTHrP levels. Placental calcium transfer may suppress PTH synthesis by the fetal parathyroid glands, and loss of the placenta may, therefore, stimulate PTH synthesis in the neonatal parathyroid glands. Since PTHrP is normally undetectable in the adult circulation, secretion of PTHrP into the circulation by fetal tissues must be lost postnatally; the time of this occurrence has not been determined. The rising PTH and 1,25dihydroxyvitamin D levels mobilize skeletal calcium to maintain the blood calcium level and increase the efficiency of intestinal calcium absorption to meet the continued demands of the mineralizing skeleton. Preterm infants are compromised by the loss of the placental calcium pump at a gestational age when rapid accretion of calcium by the skeleton normally occurs. They cannot increase their bone mineral content postnatally unless given high amounts of calcium and phosphate parenterally or in their diet. Infants of hypercalcemic mothers may have suppressed parathyroid function in the first month of life, while infants of hypocalcemic mothers may have enlarged, overactive (and occasionally autonomous) parathyroid glands that caused significant skeletal demineralization in utero. Although increased mortality has been associated with both conditions, in most instances the disturbance in neonatal calcium homeostasis is self-limited.

As stated, the adaptive strategies of the pregnant woman differ from the lactating woman, in the face of similar calcium demands. The fetal and neonatal adaptations differ, largely because the former utilizes a placental calcium pump, while the source of calcium for the latter is the intestine. If more fully understood, these adaptive mechanisms might be exploited further to treat disorders of calcium and bone metabolism in later life, such as osteoporosis.

References

- 1. Albright F, Reifenstein EC 1948 Parathyroid Glands and Metabolic Bone Disease. Williams & Wilkins, Baltimore
- Chesney RW, Specker BL, Mimouni F, McKay CP 1992 Mineral metabolism during pregnancy and lactation. In: Coe FL, Favus MJ (eds) Disorders of Bone and Mineral Metabolism. Raven Press, New York, pp 383–393
- Martin MC, Taylor RN, Kitzmiller JL 1994 The endocrinology of pregnancy. In: Greenspan FS, Baxter JD (eds) Basic & Clinical Endocrinology. Appleton & Lange, Norwalk, CT, pp 525–550
- 4. Givens MH, Macy IC 1933 The chemical composition of the human fetus. J Biol Chem 102:7–17
- Trotter M, Hixon BB 1974 Sequential changes in weight, density, and percentage ash weight of human skeletons from an early fetal period through old age. Anat Rec 179:1–18
- Comar CL 1956 Radiocalcium studies in pregnancy. Ann NY Acad Sci 64:281–298
- Mull JW, Bill AH 1934 Variations in serum calcium and phosphorus during pregnancy. Am J Obstet Gynecol 27:510–517
- Oberst WF, Plass ED 1932 The variations in serum calcium, protein, and inorganic phosphorus in early and late pregnancy, during parturition and the puerperium, and in non-pregnant women. J Clin Invest 11:123–127
- Pitkin RM, Gebhardt MP 1977 Serum calcium concentrations in human pregnancy. Am J Obstet Gynecol 127:775–778
- Kerr C, Loken HF, Glendening MB, Gordon CS, Page EW 1962 Calcium and phosphorus dynamics in pregnancy. Am J Obstet Gynecol 83:2–8
- Davis OK, Hawkins DS, Rubin LP, Posillico JT, Brown EM, Schiff I 1988 Serum parathyroid hormone (PTH) in pregnant women determined by an immunoradiometric assay for intact PTH. J Clin Endocrinol Metab 67:850–852
- Gertner JM, Coustan DR, Kliger AS, Mallette LE, Ravin N, Broadus AE 1986 Pregnancy as state of physiologic absorptive hypercalciuria. Am J Med 81:451–456
- Cruikshank DP, Pitkin RM, Reynolds WA, Williams GA, Hargis GK 1980 Calcium-regulating hormones and ions in amniotic fluid. Am J Obstet Gynecol 136:621–625
- Dahlman T, Sjoberg HE, Bucht E 1994 Calcium homeostasis in normal pregnancy and puerperium. A longitudinal study. Acta Obstet Gynecol Scand 73:393–398
- Seki K, Makimura N, Mitsui C, Hirata J, Nagata I 1991 Calciumregulating hormones and osteocalcin levels during pregnancy: a longitudinal study. Am J Obstet Gynecol 164:1248–1252
- Rasmussen N, Frolich A, Hornnes PJ, Hegedus L 1990 Serum ionized calcium and intact parathyroid hormone levels during pregnancy and postpartum. Br J Obstet Gynaecol 97:857–859
- 17. Saggese G, Baroncelli GI, Bertelloni S, Cipolloni C 1991 Intact parathyroid hormone levels during pregnancy, in healthy term neonates and in hypocalcemic preterm infants. Acta Paediatr Scand 80:36–41
- Pitkin RM, Reynolds WA, Williams GA, Hargis GK 1979 Calcium metabolism in normal pregnancy: a longitudinal study. Am J Obstet Gynecol 133:781–790
- Frolich A, Rudnicki M, Fischer-Rasmussen W, Olofsson K 1991 Serum concentrations of intact parathyroid hormone during late human pregnancy: a longitudinal study. Eur J Obstet Gynecol Reprod Biol 42:85–87
- Seely EW, Brown EM, DeMaggio DM, Weldon DK, Graves SW 1997 A prospective study of calciotropic hormones in pregnancy and post partum: reciprocal changes in serum intact parathyroid hormone and 1,25-dihydroxyvitamin D. Am J Obstet Gynecol 176: 214–217
- 21. Garner SC, Peng TC, Toverud SU 1988 Modulation of serum parathyroid hormone and ionized calcium concentrations during reproduction in rats fed a low calcium diet. J Bone Miner Res 3:319–323
- 22. Boass A, Garner SC, Schultz VL, Toverud SU 1997 Regulation of serum calcitriol by serum ionized calcium in rats during pregnancy and lactation. J Bone Miner Res 12:909–914
- 23. Chao CC, Brown RD, Deftos LJ 1985 Metabolism of calcium and

phosphorus during pregnancy and lactation in white-tailed deer. Acta Endocrinol (Copenh) 109:269-275

- Allen WM, Sansom BF 1986 Metabolic disorders. In: Howard JL (ed) Current Veterinary Therapy: Food Animal Practice. W.B. Saunders, Philadelphia, vol 2:311–322
- 25. Elias E, Shainkin-Kestenbaum R 1990 Hypocalcaemia and serum levels of inorganic phosphorus, magnesium parathyroid and calcitonin hormones in the last month of pregnancy in Awassi fat-tail ewes. Reprod Nutr Dev 30:693–699
- Gillette ME, Insogna KL, Lewis AM, Baran DT 1982 Influence of pregnancy on immunoreactive parathyroid hormone levels. Calcif Tissue Int 34:9–12
- Cross NA, Hillman LS, Allen SH, Krause GF, Vieira NE 1995 Calcium homeostasis and bone metabolism during pregnancy, lactation, and postweaning: a longitudinal study. Am J Clin Nutr 61:514–523
- Kent GN, Price RI, Gutteridge DH, Allen JR, Rosman KJ, Smith M, Bhagat CI, Wilson SG, Retallack RW 1993 Effect of pregnancy and lactation on maternal bone mass and calcium metabolism. Osteoporos Int 3 [Suppl 1]:44–47
- Brommage R, DeLuca HF 1984 Placental transport of calcium and phosphorus is not regulated by vitamin D. Am J Physiol 246:F526– F529
- Cushard Jr WG, Creditor MA, Canterbury JM, Reiss E 1972 Physiological hyperparathyroidism in pregnancy. J Clin Endocrinol Metab 34:767–771
- 31. Watney PJ, Rudd BT 1974 Calcium metabolism in pregnancy and in the newborn. J Obstet Gynaecol Br Commonw 81:210–219
- Reitz RE, Daane TA, Woods JR, Weinstein RL 1977 Calcium, magnesium, phosphorus, and parathyroid hormone interrelationships in pregnancy and newborn infants. Obstet Gynecol 50:701– 705
- Cruikshank DP, Pitkin RM, Reynolds WA, Williams GA, Hargis GK 1980 Altered maternal calcium homeostasis in diabetic pregnancy. J Clin Endocrinol Metab 50:264–267
- Drake TS, Kaplan RA, Lewis TA 1979 The physiologic hyperparathyroidism of pregnancy. Is it primary or secondary? Obstet Gynecol 53:746–749
- Hillman LS, Slatopolsky E, Haddad JG 1978 Perinatal vitamin D metabolism. IV. Maternal and cord serum 24,25-dihydroxyvitamin D concentrations. J Clin Endocrinol Metab 47:1073–1077
- Whitehead M, Lane G, Young O, Campbell S, Abeyasekera G, Hillyard CJ, MacIntyre I, Phang KG, Stevenson JC 1981 Interrelations of calcium-regulating hormones during normal pregnancy. Br Med J 283:10–12
- Wieland P, Fischer JA, Trechsel U, Roth HR, Vetter K, Schneider H, Huch A 1980 Perinatal parathyroid hormone, vitamin D metabolites, and calcitonin in man. Am J Physiol 239:E385–E390
- Allgrove J, Adami S, Manning RM, O'Riordan JL 1985 Cytochemical bioassay of parathyroid hormone in maternal and cord blood. Arch Dis Child 60:110–115
- Reynolds WA, Williams GA, Pitkin RM 1981 Calcitropic hormone responsiveness during pregnancy. Am J Obstet Gynecol 139:855– 862
- 40. **Steichen JJ, Tsang RC, Gratton TL, Hamstra A, DeLuca HF** 1980 Vitamin D homeostasis in the perinatal period: 1,25-dihydroxyvitamin D in maternal, cord, and neonatal blood. N Engl J Med 302:315–319
- Verhaeghe J, Bouillon R 1992 Calciotropic hormones during reproduction. J Steroid Biochem Mol Biol 41:469–477
- Potts Jr JT, Bringhurst FR, Gardella T, Nussbaum S, Segre G, Kronenberg HM 1995 Parathyroid hormone: physiology, chemistry, biosynthesis, secretion, metabolism, and mode of action. In: DeGroot LJ (ed) Endocrinology. W.B. Saunders, Philadelphia, pp 920–966
- 43. Thiébaud D, Janisch S, Koelbl H, Hanzal E, Jacquet AF, Leodolter S, Burckhardt P, Pecherstorfer M 1993 Direct evidence of a parathyroid related protein gradient between the mother and the newborn in humans. Bone Miner 23:213–221
- 44. Gallacher SJ, Fraser WD, Owens OJ, Dryburgh FJ, Logue FC, Jenkins A, Kennedy J, Boyle IT 1994 Changes in calciotrophic hormones and biochemical markers of bone turnover in normal human pregnancy. Eur J Endocrinol 131:369–374

- Quan-Sheng D, Miller SC 1989 Calciotrophic hormone levels and calcium absorption during pregnancy in rats. Am J Physiol 257: E118–E123
- Bourdeau A, Manganella G, Thil-Trubert CL, Sachs C, Cournot G 1990 Bioactive parathyroid hormone in pregnant rats and fetuses. Am J Physiol 258: E549–E554
- Sinclair JG 1942 Fetal rat parathyroids as affected by changes in maternal serum calcium and phosphorus through parathyroidectomy and dietary control. J Nutr 23:141–152
- Rucart G, Gagnon PM 1959 Modifications des parathyroides sous l'influence de la gestation. C R Seances Soc Biol Fil 153:1102–1103
- Schultz VL, Boass A, Garner SC, Toverud SU 1997 Altered regulation of parathyroid hormone secretion by calcium in pregnant and lactating rats. J Bone Miner Res 12:903–908
- Garner SC, Peng TC, Hirsch PF, Boass A, Toverud SU 1987 Increase in serum parathyroid hormone concentration in the lactating rat: effects of dietary calcium and lactational intensity. J Bone Miner Res 2:347–352
- 51. Bodansky M, Duff VB 1939 Regulation of the level of calcium in the serum during pregnancy. J Am Med Assoc 112:223–229
- 52. Garel JM, Gilbert M, Besnard P 1981 Fetal growth and 1,25dihydroxyvitamin D₃ injections into thyroparathyroidectomized pregnant rats. Reprod Nutr Dev 21:961–968
- Chalon S, Garel JM 1985 Plasma calcium control in the rat fetus. I. Influence of maternal hormones. Biol Neonate 48:313–322
- 54. Gilbert M, Besnard P, Garel JM 1980 Effects of maternal parathyroid glands and vitamin D_3 metabolites on fetal growth and fetal liver glycogen stores in thyro-parathyroidectomized pregnant rats. Biomedicine 32:93–99
- Lund B, Selnes A 1979 Plasma 1,25-dihydroxyvitamin D levels in pregnancy and lactation. Acta Endocrinol (Copenh) 92:330–335
- Fleischman AR, Rosen JF, Cole J, Smith CM, DeLuca HF 1980 Maternal and fetal serum 1,25-dihydroxyvitamin D levels at term. J Pediatr 97:640–642
- Bikle DD, Gee E, Halloran B, Haddad JG 1984 Free 1,25-dihydroxyvitamin D levels in serum from normal subjects, pregnant subjects, and subjects with liver disease. J Clin Invest 74:1966–1971
- Wilson SG, Retallack RW, Kent JC, Worth GK, Gutteridge DH 1990 Serum free 1,25-dihydroxyvitamin D and the free 1,25-dihydroxyvitamin D index during a longitudinal study of human pregnancy and lactation. Clin Endocrinol (Oxf) 32:613–622
- 59. Gray TK, Lowe W, Lester GE 1981 Vitamin D and pregnancy: the maternal-fetal metabolism of vitamin D. Endocr Rev 2:264–274
- Paulson SK, Ford KK, Langman CB 1990 Pregnancy does not alter the metabolic clearance of 1,25-dihydroxyvitamin D in rats. Am J Physiol 258:E158–E162
- Delvin EE, Gilbert M, Pere MC, Garel JM 1988 In vivo metabolism of calcitriol in the pregnant rabbit doe. J Dev Physiol 10:451–459
- 62. Ross R, Halbert K, Tsang RC 1989 Determination of the production and metabolic clearance rates of 1,25-dihydroxyvitamin D₃ in the pregnant sheep and its chronically catheterized fetus by primed infusion technique. Pediatr Res 26:633–638
- 63. **Ross R, Dorsey J, Ellis K** 1990 Progressive increases in 1,25-dihydroxyvitamin D₃ production rate in multiple ovine pregnancy are independent of changes in the metabolic clearance rate. Pediatr Res 27:192A (Abstract)
- 64. Kubota M, Ohno J, Shiina Y, Suda T 1982 Vitamin D metabolism in pregnant rabbits: differences between the maternal and fetal response to administration of large amounts of vitamin D₃. Endocrinology 110:1950–1956
- Fenton E, Britton HG 1980 25-hydroxycholecalciferol 1 alpha-hydroxylase activity in the kidney of the fetal, neonatal and adult guinea pig. Biol Neonate 37:254–256
- 66. Ross R, Care AD, Robinson JS, Pickard DW, Weatherley AJ 1980 Perinatal 1,25-dihydroxycholecalciferol in the sheep and its role in the maintenance of the transplacental calcium gradient. J Endocrinol 87:17P–18P
- 67. **Baksi SN, Kenny AD** 1978 Acute effect of estradiol on the renal vitamin D hydroxylases in Japanese quail. Biochem Pharmacol 27:2765–2768
- Spanos E, Colston KW, Evans IM, Galante LS, Macauley SJ, MacIntyre I 1976 Effect of prolactin on vitamin D metabolism. Mol Cell Endocrinol 5:163–167

- Spanos E, Brown DJ, Stevenson JC, MacIntyre I 1981 Stimulation of 1,25-dihydroxycholecalciferol production by prolactin and related peptides in intact renal cell preparations *in vitro*. Biochim Biophys Acta 672:7–15
- Takeuchi K, Morikawa H, Ueda Y, Mochizuki M 1988 Studies on the effects of placental lactogen on calcium metabolism during pregnancy]. Nippon Naibunpi Gakkai Zasshi 64:1175–1186
- Cheema C, Grant BF, Marcus R 1989 Effects of estrogen on circulating free and total 1,25-dihydroxyvitamin D and on the parathyroid-vitamin D axis in postmenopausal women. J Clin Invest 83:537–542
- Kumar R, Abboud CF, Riggs BL 1980 The effect of elevated prolactin levels on plasma 1,25-dihydroxyvitamin D and intestinal absorption of calcium. Mayo Clin Proc 55:51–53
- Reddy GS, Norman AW, Willis DM, Goltzman D, Guyda H, Solomon S, Philips DR, Bishop JE, Mayer E 1983 Regulation of vitamin D metabolism in normal human pregnancy. J Clin Endocrinol Metab 56:363–370
- Weisman Y, Vargas A, Duckett G, Reiter E, Root AW 1978 Synthesis of 1,25-dihydroxyvitamin D in the nephrectomized pregnant rat. Endocrinology 103:1992–1996
- 75. Tanaka Y, Halloran B, Schnoes HK, DeLuca HF 1979 In vitro production of 1,25-dihydroxyvitamin D₃ by rat placental tissue. Proc Natl Acad Sci USA 76:5033–5035
- Weisman Y, Sapir R, Harell A, Edelstein S 1976 Maternal-perinatal interrelationships of vitamin D metabolism in rats. Biochim Biophys Acta 428:388–395
- 77. Glorieux FH, Arabian A, Delvin EE 1995 Pseudo-vitamin D deficiency: absence of 25-hydroxyvitamin D 1 alpha-hydroxylase activity in human placenta decidual cells. J Clin Endocrinol Metab 80:2255–2258
- 78. Gray TK, Lester GE, Lorenc RS 1979 Evidence for extra-renal 1α -hydroxylation of 25-hydroxyvitamin D₃ in pregnancy. Science 204:1311–1313
- 79. Lachenmaier-Currle U, Harmeyer J 1989 Placental transport of calcium and phosphorus in pigs. J Perinat Med 17:127–136
- Turner M, Barre PE, Benjamin A, Goltzman D, Gascon-Barre M 1988 Does the maternal kidney contribute to the increased circulating 1,25-dihydroxyvitamin D concentrations during pregnancy? Miner Electrolyte Metab 14:246–252
- Pike JW, Parker JB, Haussler MR, Boass A, Toverud SV 1979 Dynamic changes in circulating 1,25-dihydroxyvitamin D during reproduction in rats. Science 204:1427–1429
- Halloran BP, Barthell EN, DeLuca HF 1979 Vitamin D metabolism during pregnancy and lactation in the rat. Proc Natl Acad Sci USA 76:5549–5553
- Delvin EE, Salle BL, Glorieux FH, Adeleine P, David LS 1986 Vitamin D supplementation during pregnancy: effect on neonatal calcium homeostasis. J Pediatr 109:328–334
- Halloran BP, De Luca HF 1981 Effect of vitamin D deficiency on skeletal development during early growth in the rat. Arch Biochem Biophys 209:7–14
- Halloran BP, DeLuca HF 1979 Vitamin D deficiency and reproduction in rats. Science 204:73–74
- Samaan NA, Anderson GD, Adam-Mayne ME 1975 Immunoreactive calcitonin in the mother, neonate, child and adult. Am J Obstet Gynecol 121:622–625
- Silva OL, Titus-Dillon P, Becker KL, Snider RH, Moore CF 1981 Increased serum calcitonin in pregnancy. J Natl Med Assoc 73: 649–652
- Kovarik J, Woloszczuk W, Linkesch W, Pavelka R 1980 Calcitonin in pregnancy (letter). Lancet 1:199–200
- Woloszczuk W, Kovarik J, Pavelka P 1981 Calcitonin in pregnant women and in cord blood. Gynecol Obstet Invest 12:272–276
- Garel JM, Savajol H, Barlet JP 1976 Plasma immunoreactive calcitonin levels in pregnant ewes and their lambs. Biol Neonate 28:207–218
- Barlet JP, Garel JM 1975 Physiological role of calcitonin in pregnant goats and ewes. In: Talmage RV, Owen M, Parsons A (eds) Calcium Regulating Hormones: Proceedings of the Fifth Parathyroid Conference, Oxford, U.K., July 21–26, 1974. Excerpta Medica, Amsterdam, pp 119–121

- Garel JM, Jullienne A 1977 Plasma calcitonin levels in pregnant and newborn rats. J Endocrinol 75:373–382
- Bucht E, Telenius-Berg M, Lundell G, Sjoberg HE 1986 Immunoextracted calcitonin in milk and plasma from totally thyroidectomized women. Evidence of monomeric calcitonin in plasma during pregnancy and lactation. Acta Endocrinol (Copenh) 113:529– 535
- 94. Balabanova S, Kruse B, Wolf AS 1987 Calcitonin secretion by human placental tissue. Acta Obstet Gynecol Scand 66:323–326
- Deftos LJ 1993 Calcitonin. In: Favus MJ (ed) Primer on the Metabolic Bone Diseases and Disorders of Mineral Metabolism. Raven Press, New York, pp 70–76
- Taylor TG, Lewis PE, Balderstone O 1975 Role of calcitonin in protecting the skeleton during pregnancy and lactation. J Endocrinol 66:297–298
- Lewis P, Rafferty B, Shelley M, Robinson CJ 1971 A suggested physiological role of calcitonin: the protection of the skeleton during pregnancy and lactation. J Endocrinol 49:9–10
- Barlet JP 1974 Role physiologique de la calcitonine chez la chèvre gestante ou allaitante. Ann Biol Anim Biochim Biophys 14:447–457
- de Papp AE, Stewart AF 1993 Parathyroid hormone-related protein: a peptide of diverse physiologic functions. Trends Endocrinol Metab 4:181–187
- 100. Orloff JJ, Reddy D, de Papp AE, Yang KH, Soifer NE, Stewart AF 1994 Parathyroid hormone-related protein as a prohormone: posttranslational processing and receptor interactions. Endocr Rev 15: 40–60
- 101. Moseley JM, Kubota M, Diefenbach-Jagger H, Wettenhall RE, Kemp BE, Suva LJ, Rodda CP, Ebeling PR, Hudson PJ, Zajac JD, Martin TJ 1987 Parathyroid hormone-related protein purified from a human lung cancer cell line. Proc Natl Acad Sci USA 84:5048–5052
- 102. Mangin M, Webb AC, Dreyer BE, Posillico JT, Ikeda K, Weir EC, Stewart AF, Bander NH, Milstone L, Barton DE, Francke U, Broadus AE 1988 Identification of a cDNA encoding a parathyroid hormone-like peptide from a human tumor associated with humoral hypercalcemia of malignancy. Proc Natl Acad Sci USA 85: 597–601
- 103. Suva LJ, Winslow GA, Wettenhall RE, Hammonds RG, Moseley JM, Diefenbach-Jagger H, Rodda CP, Kemp BE, Rodriguez H, Chen EY, Hudson PJ, Martin TJ, Wood WI 1987 A parathyroid hormone-related protein implicated in malignant hypercalcemia: cloning and expression. Science 237:893–896
- Martin TJ, Moseley JM 1995 Parathyroid hormone-related protein. In: DeGroot LJ (ed) Endocrinology. W.B. Saunders, Philadelphia, pp 967–977
- 105. Barri ME, Abbas SK, Care AD 1992 The effects in the rat of two fragments of parathyroid hormone-related protein on uterine contractions *in situ*. Exp Physiol 77:481–490
- 106. Ferguson II JE, Gorman JV, Bruns DE, Weir EC, Burtis WJ, Martin TJ, Bruns ME 1992 Abundant expression of parathyroid hormonerelated protein in human amnion and its association with labor. Proc Natl Acad Sci USA 89:8384–8388
- 107. Fenton AJ, Kemp BE, Hammonds Jr RG, Mitchelhill K, Moseley JM, Martin TJ, Nicholson GC 1991 A potent inhibitor of osteoclastic bone resorption within a highly conserved pentapeptide region of parathyroid hormone-related protein; PTHrP[107–111]. Endocrinology 129:3424–3426
- 108. Fenton AJ, Kemp BE, Kent GN, Moseley JM, Zheng MH, Rowe DJ, Britto JM, Martin TJ, Nicholson GC 1991 A carboxyl-terminal peptide from the parathyroid hormone-related protein inhibits bone resorption by osteoclasts. Endocrinology 129:1762–1768
- 109. Cornish J, Callon KE, Nicholson GC, Reid IR 1997 Parathyroid hormone-related protein-(107–139) inhibits bone resorption *in vivo*. Endocrinology 138:1299–1304
- Khosla S, Johansen KL, Ory SJ, O'Brien PC, Kao PC 1990 Parathyroid hormone-related peptide in lactation and in umbilical cord blood. Mayo Clin Proc 65:1408–1414
- 111. **Bertelloni S, Baroncelli GI, Pelletti A, Battini R, Saggese G** 1994 Parathyroid hormone-related protein in healthy pregnant women. Calcif Tissue Int 54:195–197
- 112. Ratcliffe WA, Abbas SK, Care AD 1993 Clearance of exogenous parathyroid hormone-related protein in pregnant, non-pregnant and fetal sheep, goats and pigs. J Endocrinol 138:459–465

December, 1997

- 113. **Ratcliffe WA, Thompson GE, Abbas SK, Care AD** 1992 Studies on the metabolic clearance of parathyroid hormone-related protein in pregnant, nonpregnant and fetal animals. In: Cohn DV, Gennari C, Tashjian Jr AH (eds) Calcium Regulating Hormones and Bone Metabolism: Basic and Clinical Aspects. Elsevier Science Publishers, New York, pp 69–75
- 114. Senior PV, Heath DA, Beck F 1991 Expression of parathyroid hormone-related protein mRNA in the rat before birth: demonstration by hybridization histochemistry. J Mol Endocrinol 6:281– 290
- 115. Ferguson JE, Seaner R, Bruns DE, Redick JA, Mills SE, Jüppner H, Segre GV, Bruns ME 1994 Expression of parathyroid hormonerelated protein and its receptor in human umbilical cord: evidence for a paracrine system involving umbilical vessels. Am J Obstet Gynecol 170:1018–1024
- MacIsaac RJ, Caple IW, Danks JA, Diefenbach-Jagger H, Grill V, Moseley JM, Southby J, Martin TJ 1991 Ontogeny of parathyroid hormone-related protein in the ovine parathyroid gland. Endocrinology 129:757–764
- 117. Budayr AA, Halloran BP, King JC, Diep D, Nissenson RA, Strewler GJ 1989 High levels of a parathyroid hormone-like protein in milk. Proc Natl Acad Sci USA 86:7183–7185
- 118. Rakopoulos M, Vargas SJ, Gillespie MT, Ho PW, Diefenbach-Jagger H, Leaver DD, Grill V, Moseley JM, Danks JA, Martin TJ 1992 Production of parathyroid hormone-related protein by the rat mammary gland in pregnancy and lactation. Am J Physiol 263: E1077–E1085
- 119. Van Heerden JA, Gharib H, Jackson IT 1988 Pseudohyperparathyroidism secondary to gigantic mammary hypertrophy. Arch Surg 123:80–82
- Pahuja DN, DeLuca HF 1981 Stimulation of intestinal calcium transport and bone calcium mobilization by prolactin in vitamin D-deficient rats. Science 214:1038–1039
- 121. **Mainoya JR** 1975 Effects of bovine growth hormone, human placental lactogen and ovine prolactin on intestinal fluid and ion transport in the rat. Endocrinology 96:1165–1170
- 122. Burstyn PG, Lloyd IJ, McKillop W 1975 The effect of human placental lactogen on the renal excretion of calcium in the rabbit. IRCS J Med Sci 3:30
- 123. **Burstyn PG, McKillop W, Lloyd IJ** 1974 The effects of prolactin on the renal excretion of water, sodium, potassium and calcium in the rabbit. J Int Res Commun 2:1474
- 124. Dvir R, Golander A, Jaccard N, Yedwab G, Otremski I, Spirer Z, Weisman Y 1995 Amniotic fluid and plasma levels of parathyroid hormone-related protein and hormonal modulation of its secretion by amniotic fluid cells. Eur J Endocrinol 133:277–282
- Bringhurst FR 1995 Calcium and phosphate distribution, turnover, and metabolic actions. In: DeGroot LJ (ed) Endocrinology. W.B. Saunders, Philadelphia, pp 1015–1043
 Heaney RP, Skillman TG 1971 Calcium metabolism in normal
- 126. **Heaney RP, Skillman TG** 1971 Calcium metabolism in normal human pregnancy. J Clin Endocrinol Metab 33:661–670
- 127. Kent GN, Price RI, Gutteridge DH, Rosman KJ, Smith M, Allen JR, Hickling CJ, Blakeman SL 1991 The efficiency of intestinal calcium absorption is increased in late pregnancy but not in established lactation. Calcif Tissue Int 48:293–295
- 128. Kent GN, Price RI, Gutteridge DH, Allen JR, Blakeman SL, Bhagat CI, St.John A, Barnes MP, Smith M, Evans DV 1991 Acute effects of an oral calcium load in pregnancy and lactation: findings on renal calcium conservation and biochemical indices of bone turnover. Miner Electrolyte Metab 17:1–7
- 129. Delorme AC, Marche P, Garel JM 1979 Vitamin D-dependent calcium-binding protein. Changes during gestation, prenatal and postnatal development in rats. J Dev Physiol 1:181–194
- Delorme AC, Danan JL, Ripoche MA, Mathieu H 1982 Biochemical characterization of mouse vitamin D-dependent calcium-binding protein. Evidence for its presence in embryonic life. Biochem J 205:49–57
- Bruns ME, Vollmer S, Wallshein V, Bruns DE 1981 Vitamin Ddependent calcium-binding protein. Immunochemical studies and synthesis by placental tissue *in vitro*. J Biol Chem 256:4649–4653
- 132. Marche P, Delorme A, Cuisinier-Gleizes P 1978 Intestinal and placental calcium-binding proteins in vitamin D-deprived or -supplemented rats. Life Sci 23:2555–2561

- 133. **Bruns ME, Wallshein V, Bruns DE** 1982 Regulation of calciumbinding protein in mouse placenta and intestine. Am J Physiol 242:E47–E52
- 134. **Ibrahim MM, Thomas ML, Forte LR** 1984 Maternal-fetal relationships in the parathyroidectomized rat. Intestinal calcium transport, serum calcium, immunoreactive parathyroid hormone and calcitonin. Biol Neonate 46:89–97
- 135. Wasserman RH, Comar CL, Nold MM, Lengemann FW 1957 Placental transfer of calcium and strontium in the rat and rabbit. Am J Physiol 189:91–97
- 136. Ellinger GM, Duckworth J 1952 Skeletal changes during pregnancy and lactation in the rat: effect of different levels of dietary calcium. Br J Nutr 6:235–253
- Halloran BP, DeLuca HF 1980 Skeletal changes during pregnancy and lactation: the role of vitamin D. Endocrinology 107:1923–1929
 Bawden JW, Wolkoff AS 1964 Distribution of ⁴⁵Ca during preg-
- Bawden JW, Wolkoff AS 1964 Distribution of ⁴⁵Ca during pregnancy under conditions of calcium deficiency in rats. J Dent Res 43:563–567
- 139. **Rasmussen P** 1977 Calcium deficiency, pregnancy, and lactation in rats. Some effects on blood chemistry and the skeleton. Calcif Tissue Res 23:87–94
- 140. **Miller SC, Halloran BP, DeLuca HF, Jee WS** 1982 Role of vitamin D in maternal skeletal changes during pregnancy and lactation: a histomorphometric study. Calcif Tissue Int 34:245–252
- 141. Halloran BP, DeLuca HF 1980 Calcium transport in small intestine during pregnancy and lactation. Am J Physiol 239:E64–E68
- 142. Brommage R, Baxter DC, Gierke LW 1990 Vitamin D-independent intestinal calcium and phosphorus absorption during reproduction. Am J Physiol 259:G631–G638
- 143. Thomas ML, Forte LR 1982 Serum calcium and parathyroid hormone during the reproductive cycle in normal and vitamin Ddeficient rats. Endocrinology 110:703–707
- 144. **Gaboury CL, Woods LL** 1995 Renal reserve in pregnancy. Semin Nephrol 15:449–453
- 145. **Baylis C** 1994 Glomerular filtration and volume regulation in gravid animal models. Baillieres Clin Obstet Gynaecol 8:235–264
- 146. Allen J, Kent N, Price R, Gutteridge D, Blakeman S, Rosman K, Bhagat C, Smith M 1990 Calcium and phosphate metabolism in human pregnancy and lactation. Bone Miner 10:S317
- 147. Pedersen EB, Johannesen P, Kristensen S, Rasmussen AB, Emmertsen K, Moller J, Lauritsen JG, Wohlert M 1984 Calcium, parathyroid hormone and calcitonin in normal pregnancy and preeclampsia. Gynecol Obstet Invest 18:156–164
- 148. **Chef R** 1969 Métabolisme du calcium chez la ratte en gestation. Etude cinetique par le 45Ca. C R Seances Soc Biol Fil 163:541–545
- 149. Frenkel Y, Barkai G, Mashiach S, Dolev E, Zimlichman R, Weiss M 1991 Hypocalciuria of preeclampsia is independent of parathyroid hormone level. Obstet Gynecol 77:689–691
- 150. August P, Marcaccio B, Gertner JM, Druzin ML, Resnick LM, Laragh JH 1992 Abnormal 1,25-dihydroxyvitamin D metabolism in preeclampsia. Am J Obstet Gynecol 166:1295–1299
- 151. Seely EW, Wood RJ, Brown EM, Graves SW 1992 Lower serum ionized calcium and abnormal calciotropic hormone levels in preeclampsia. J Clin Endocrinol Metab 74:1436–1440
- 152. Lalau JD, Jans I, el Esper N, Bouillon R, Fournier A 1993 Calcium metabolism, plasma parathyroid hormone, and calcitriol in transient hypertension of pregnancy. Am J Hypertens 6:522–527
- 153. Levine RJ, Hauth JC, Curet LB, Sibai BM, Catalano PM, Morris CD, DerSimonian R, Esterlitz JR, Raymond EG, Bild DE, Clemens JD, Cutler JA 1997 Trial of calcium to prevent preeclampsia. N Engl J Med 337:69–76
- 154. Marie PJ, Cancela L, Le Boulch N, Miravet L 1986 Bone changes due to pregnancy and lactation: influence of vitamin D status. Am J Physiol 251: E400–E406
- 155. **Fukuda S, Iida H** 1993 Histomorphometric changes in iliac trabecular bone during pregnancy and lactation in beagle dogs. J Vet Med Sci 55:565–569
- 156. Warnock GM, Duckworth J 1944 Changes in the skeleton during gestation and lactation in the rat. Biochem J 38:220–224
- 157. Braithwaite GD, Glascock RF, Riazuddin S 1970 Calcium metabolism in pregnant ewes. Br J Nutr 24:661–670
- 158. Delmas PD 1993 Markers of bone formation and resorption. In:

Favus MJ (ed) Primer on the Metabolic Bone Diseases and Disorders of Mineral Metabolism. Raven Press, New York, pp 108–112

- 159. **Calvo MS, Eyre DR, Gundberg CM** 1996 Molecular basis and clinical application of biological markers of bone turnover. Endocr Rev 17:333–368
- 160. Cross NA, Hillman LS, Allen SH, Krause GF 1995 Changes in bone mineral density and markers of bone remodeling during lactation and postweaning in women consuming high amounts of calcium. J Bone Miner Res 10:1312–1320
- 161. Yamaga A, Taga M, Minaguchi H, Sato K 1996 Changes in bone mass as determined by ultrasound and biochemical markers of bone turnover during pregnancy and puerperium: a longitudinal study. J Clin Endocrinol Metab 81:752–756
- 162. Rodin A, Duncan A, Quartero HW, Pistofidis G, Mashiter G, Whitaker K, Crook D, Stevenson JC, Chapman MG, Fogelman I 1989 Serum concentrations of alkaline phosphatase isoenzymes and osteocalcin in normal pregnancy. J Clin Endocrinol Metab 68:1123–1127
- Karlsson R, Eden S, Eriksson L, von Schoultz B 1992 Osteocalcin 24-hour profiles during normal pregnancy. Gynecol Obstet Invest 34:197–201
- 164. Chesnut CH 1992 The imaging and quantitation of bone by radiographic and scanning methodologies. In: Coe FL, Favus MJ (eds) Disorders of Bone and Mineral Metabolism. Raven Press, New York, pp 443–454
- 165. Genant HK, Engelke K, Fuerst T, Glüer CC, Grampp S, Harris ST, Jergas M, Lang T, Lu Y, Majumdar S, Mathur A, Takada M 1996 Noninvasive assessment of bone mineral and structure: state of the art. J Bone Miner Res 11:707–730
- Lamke B, Brundin J, Moberg P 1977 Changes of bone mineral content during pregnancy and lactation. Acta Obstet Gynecol Scand 56:217–219
- Christiansen C, Rodbro P, Heinild B 1976 Unchanged total body calcium in normal human pregnancy. Acta Obstet Gynecol Scand 55:141–143
- 168. Sowers M, Crutchfield M, Jannausch M, Updike S, Corton G 1991 A prospective evaluation of bone mineral change in pregnancy. Obstet Gynecol 77:841–845
- Drinkwater BL, Chesnut III CH 1991 Bone density changes during pregnancy and lactation in active women: a longitudinal study. Bone Miner 14:153–160
- 170. Paparella P, Giorgino R, Maglione A, Lorusso D, Scirpa P, Del Bosco A, Mancuso S 1995 Maternal ultrasound bone density in normal pregnancy. Clin Exp Obstet Gynecol 22:268–278
- 171. Gambacciani M, Spinetti A, Gallo R, Cappagli B, Teti GC, Facchini V 1995 Ultrasonographic bone characteristics during normal pregnancy: longitudinal and cross-sectional evaluation. Am J Obstet Gynecol 173:890–893
- 172. van Hemert AM, Vandenbroucke JP, Birkenhager JC, Valkenburg HA 1990 Prediction of osteoporotic fractures in the general population by a fracture risk score. A 9-year follow-up among middle-aged women. Am J Epidemiol 132:123–135
- 173. Johansson C, Mellstrom D, Milson I 1993 Reproductive factors as predictors of bone density and fractures in women at the age of 70. Maturitas 17:39–50
- 174. Kritz-Silverstein D, Barrett-Connor E, Hollenbach KA 1992 Pregnancy and lactation as determinants of bone mineral density in postmenopausal women. Am J Epidemiol 136:1052–1059
- 175. Armamento-Villareal R, Villareal DT, Avioli LV, Civitelli R 1992 Estrogen status and heredity are major determinants of premenopausal bone mass. J Clin Invest 90:2464–2471
- 176. Alderman BW, Weiss NS, Daling JR, Ure CL, Ballard JH 1986 Reproductive history and postmenopausal risk of hip and forearm fracture. Am J Epidemiol 124:262–267
- 177. Walker AR, Richardson B, Walker F 1972 The influence of numerous pregnancies and lactations on bone dimensions in South African Bantu and Caucasian mothers. Clin Sci 42:189–196
- 178. Sowers MR, Clark MK, Hollis B, Wallace RB, Jannausch M 1992 Radial bone mineral density in pre- and perimenopausal women: a prospective study of rates and risk factors for loss. J Bone Miner Res 7:647–657
- 179. Johnell O, Gullberg B, Kanis JA, Allander E, Elffors L, Dequeker J, Dilsen G, Gennari C, Lopes Vaz A, Lyritis G, Mazzuoli G,

Miravet L, Passeri M, Perez Cano R, Rapado A, Ribot C 1995 Risk factors for hip fracture in European women: the MEDOS Study. Mediterranean Osteoporosis Study. J Bone Miner Res 10:1802–1815

- Melton III LJ, Bryant SC, Wahner HW, O'Fallon WM, Malkasian GD, Judd HL, Riggs BL 1993 Influence of breastfeeding and other reproductive factors on bone mass later in life. Osteoporos Int 3:76–83
- 181. Cox ML, Khan SA, Gau DW, Cox SA, Hodkinson HM 1991 Determinants of forearm bone density in premenopausal women: a study in one general practice. Br J Gen Pract 41:194–196
- 182. Picard D, Ste-Marie LG, Coutu D, Carrier L, Chartrand R, Lepage R, Fugere P, D'Amour P 1988 Premenopausal bone mineral content relates to height, weight and calcium intake during early adulthood. Bone Miner 4:299–309
- 183. Bauer DC, Browner WS, Cauley JA, Orwoll ES, Scott JC, Black DM, Tao JL, Cummings SR 1993 Factors associated with appendicular bone mass in older women. The Study of Osteoporotic Fractures Research Group. Ann Intern Med 118:657–665
- 184. Kreiger N, Kelsey JL, Holford TR, O'Connor T 1982 An epidemiologic study of hip fracture in postmenopausal women. Am J Epidemiol 116:141–148
- Kreiger N, Gross A, Hunter G 1992 Dietary factors and fracture in postmenopausal women: a case-control study. Int J Epidemiol 21: 953–958
- 186. Ribot C, Tremollieres F, Pouilles JM, Albarede JL, Mansat M, Utheza G, Bonneu M, Bonnissent P, Ricoeur C 1993 Risk factors for hip fracture. MEDOS study: results of the Toulouse Centre. Bone 14[Suppl 1]:S77–S80
- Cumming RG, Klineberg RJ 1993 Breastfeeding and other reproductive factors and the risk of hip fractures in elderly women. Int J Epidemiol 22:684–691
- 188. Johnell O, Nilsson BE 1984 Life-style and bone mineral mass in perimenopausal women. Calcif Tissue Int 36:354–356
- 189. Sowers MR, Wallace RB, Lemke JH 1985 Correlates of mid-radius bone density among postmenopausal women: a community study. Am J Clin Nutr 41:1045–1053
- 190. Sowers M, Wallace RB, Lemke JH 1985 Correlates of forearm bone mass among women during maximal bone mineralization. Prev Med 14:585–596
- 191. Sinigaglia L, Varenna M, Binelli L, Gallazzi M, Calori G, Ranza R 1996 Effect of lactation on postmenopausal bone mineral density of the lumbar spine. J Reprod Med 41:439–443
- 192. Laitinen K, Valimaki M, Keto P 1991 Bone mineral density measured by dual-energy X-ray absorptiometry in healthy Finnish women. Calcif Tissue Int 48:224–231
- 193. Stevenson JC, Lees B, Devenport M, Cust MP, Ganger KF 1989 Determinants of bone density in normal women: risk factors for future osteoporosis? Br Med J 298:924–928
- 194. Nilsson BE 1969 Parity and osteoporosis. Surg Gynecol Obstet 129:27–28
- 195. Fox KM, Magaziner J, Sherwin R, Scott JC, Plato CC, Nevitt M, Cummings S 1993 Reproductive correlates of bone mass in elderly women. Study of Osteoporotic Fractures Research Group. J Bone Miner Res 8:901–908
- Goldsmith NF, Johnston JO 1975 Bone mineral: effects of oral contraceptives, pregnancy, and lactation. J Bone Joint Surg [Am] 57:657–668
- 197. Hoffman S, Grisso JA, Kelsey JL, Gammon MD, O'Brien LA 1993 Parity, lactation and hip fracture. Osteoporos Int 3:171–176
- 198. Paganini-Hill A, Chao A, Ross RK, Henderson BE 1991 Exercise and other factors in the prevention of hip fracture: the Leisure World study. Epidemiology 2:16–25
- Lissner L, Bengtsson C, Hansson T 1991 Bone mineral content in relation to lactation history in pre- and postmenopausal women. Calcif Tissue Int 48:319–325
- Biberoglu KO, Yildiz A, Kandemir O 1993 Bone mineral density in Turkish postmenopausal women. Int J Gynaecol Obstet 41:153– 157
- 201. Parra-Cabrera S, Hernandez-Avila M, Tamayo-y-Orozco J, L-pez-Carrillo L, Meneses-González F 1996 Exercise and reproductive factors as predictors of bone density among osteoporotic women in Mexico City. Calcif Tissue Int 59:89–94

- 202. Fujiwara S, Kasagi F, Yamada M, Kodama K 1997 Risk factors for hip fracture in a Japanese cohort. J Bone Miner Res 12:998–1004
- 203. Hreshchyshyn MM, Hopkins A, Zylstra S, Anbar M 1988 Associations of parity, breast-feeding, and birth control pills with lumbar spine and femoral neck bone densities. Am J Obstet Gynecol 159:318–322
- 204. Jones OV 1953 Crush fracture of the dorsal spine in eclampsia. J Obstet Gynaecol Br Emp 60:259–262
- Nordin BE, Roper A 1955 Postpregnancy osteoporosis: a syndrome? Lancet 1:431–434
- 206. Jackson WP 1958 Osteoporosis of unknown cause in younger people: idiopathic osteoporosis. J Bone Joint Surg [Br] 40:420-441
- Dent CE, Friedman M 1965 Pregnancy and idiopathic osteoporosis. Q J Med 34:341–357
- 208. Smith R, Stevenson JC, Winearls CC, Woods CG, Wordsworth BP 1985 Osteoporosis of pregnancy. Lancet 1:1178–1180
- Dunne F, Walters B, Marshall T, Heath DA 1993 Pregnancy associated osteoporosis. Clin Endocrinol (Oxf) 39:487–490
- Rillo OL, Di Stefano CA, Bermudez J, Maldonado Cocco JA 1994 Idiopathic osteoporosis during pregnancy. Clin Rheumatol 13:299– 304
- 211. Chung HC, Lim SK, Lee MK, Lee MH, Huh KB 1988 Pregnancyassociated osteoporosis. Yonsei Med J 29:286–294
- 212. Smith R, Athanasou NA, Ostlere SJ, Vipond SE 1995 Pregnancyassociated osteoporosis. Q J Med 88:865–878
- 213. Carbone LD, Palmieri GM, Graves SC, Smull K 1995 Osteoporosis of pregnancy: long-term follow-up of patients and their offspring. Obstet Gynecol 86:664–666
- 214. Goldman GA, Friedman S, Hod M, Ovadia J 1994 Idiopathic transient osteoporosis of the hip in pregnancy. Int J Gynaecol Obstet 46:317–320
- 215. Longstreth PL, Malinak LR, Hill Jr CS 1973 Transient osteoporosis of the hip in pregnancy. Obstet Gynecol 41:563–569
- Brodell JD, Burns Jr JE, Heiple KG 1989 Transient osteoporosis of the hip of pregnancy. Two cases complicated by pathological fracture. J Bone Joint Surg [Am] 71:1252–1257
- 217. Guerra JJ, Steinberg ME 1995 Distinguishing transient osteoporosis from avascular necrosis of the hip. J Bone Joint Surg [Am] 77:616-624
- Funk JL, Shoback DM, Genant HK 1995 Transient osteoporosis of the hip in pregnancy: natural history of changes in bone mineral density. Clin Endocrinol (Oxf) 43:373–382
- 219. Lose G, Lindholm P 1986 Transient painful osteoporosis of the hip in pregnancy. Int J Gynaecol Obstet 24:13–16
- Curtiss Jr PH, Kincaid WE 1959 Transitory demineralization of the hip in pregnancy: a report of three cases. J Bone Joint Surg [Am] 41:1327–1333
- 221. Takatori Y, Kokubo T, Ninomiya S, Nakamura T, Okutsu I, Kamogawa M 1991 Transient osteoporosis of the hip. Magnetic resonance imaging. Clin Orthop 271:190–194
- Beaulieu JG, Razzano CD, Levine RB 1976 Transient osteoporosis of the hip in pregnancy. Clin Orthop 115:165–168
- 223. **Gruber HE, Gutteridge DH, Baylink DJ** 1984 Osteoporosis associated with pregnancy and lactation: bone biopsy and skeletal features in three patients. Metab Bone Dis Relat Res 5:159–165
- 224. Payne RB, Little AJ, Evans RT 1990 Albumin-adjusted calcium concentration in serum increases during normal pregnancy. Clin Chem 36:142–144
- 225. Ludwig GD 1962 Hyperparathyroidism in relation to pregnancy. N Engl J Med 267:637–642
- Shangold MM, Dor N, Welt SI, Fleischman AR, Crenshaw Jr MC 1982 Hyperparathyroidism and pregnancy: a review. Obstet Gynecol Surv 37:217–228
- 227. Wagner G, Transhol L, Melchior JC 1964 Hyperparathyroidism and pregnancy. Acta Endocrinol (Copenh) 47:549–564
- Bruce J, Strong JA 1955 Maternal hyperparathyroidism and parathyroid deficiency in the child, with account of effect of parathyroidectomy on renal function, and of attempt to transplant part of tumor. Q J Med 24:307–319
- 229. Better OS, Levi J, Grief E, Tuma S, Gellei B, Erlik D 1973 Prolonged neonatal parathyroid suppression. A sequel to asymptomatic maternal hyperparathyroidism. Arch Surg 106:722–724

- 230. Garel JM, Dumont C 1972 Distribution and inactivation of labeled parathyroid hormone in rat fetus. Horm Metab Res 4:217–221
- 231. Garel JM 1972 Distribution of labeled parathyroid hormone in rat fetus. Horm Metab Res 4:131–132
- Northrop G, Misenhimer HR, Becker FO 1977 Failure of parathyroid hormone to cross the nonhuman primate placenta. Am J Obstet Gynecol 129:449–453
- Kaplan EL, Burrington JD, Klementschitsch P, Taylor J, Deftos L 1984 Primary hyperparathyroidism, pregnancy, and neonatal hypocalcemia. Surgery 96:717–722
- Delmonico FL, Neer RM, Cosimi AB, Barnes AB, Russell PS 1976 Hyperparathyroidism during pregnancy. Am J Surg 131:328–337
 Johnstone RE, Kreindler T 1972 Hyperparathyroidism during
- Johnstone RE, Kreindler T 1972 Hyperparathyroidism during pregnancy. Obstet Gynecol 40:580–585
- 236. Wilson DT, Martin T, Christensen R, Yee AH, Reynolds C 1983 Hyperparathyroidism in pregnancy: case report and review of the literature. Can Med Assoc J 129:986–989
- 237. Rubin A, Chaykin L, Ludwig GD 1968 Maternal hyperparathyroidism and pregnancy. J Am Med Assoc 206:128–130
- 238. Higgins RV, Hisley JC 1988 Primary hyperparathyroidism in pregnancy. A report of two cases. J Reprod Med 33:726–730
- 239. Lueg MC, Dawkins WE 1983 Primary hyperparathyroidism and pregnancy. South Med J 76:1389–1392
- Lowe DK, Orwoll ES, McClung MR, Cawthon ML, Peterson CG 1983 Hyperparathyroidism and pregnancy. Am J Surg 145:611–614
- 241. Thomas BR, Bennett JD 1995 Symptomatic hypocalcemia and hypoparathyroidism in two infants of mothers with hyperparathyroidism and familial benign hypercalcemia. J Perinatol 15:23–26
- 242. Marx SJ, Attie MF, Levine MA, Spiegel AM, Downs Jr RW, Lasker RD 1981 The hypocalciuric or benign variant of familial hypercalcemia: clinical and biochemical features in fifteen kindreds. Medicine (Baltimore) 60:397–412
- 243. **Powell BR, Buist NR** 1990 Late presenting, prolonged hypocalcemia in an infant of a woman with hypocalciuric hypercalcemia. Clin Pediatr (Phila) 29:241–243
- 244. **Rude RK, Haussler MR, Singer FR** 1984 Postpartum resolution of hypocalcemia in a lactating hypoparathyroid patient. Endocrinol Jpn 31:227–233
- 245. Cundy T, Haining SA, Guilland-Cumming DF, Butler J, Kanis JA 1987 Remission of hypoparathyroidism during lactation: evidence for a physiological role for prolactin in the regulation of vitamin D metabolism. Clin Endocrinol (Oxf) 26:667–674
- Bronsky D, Kiamko RT, Moncada R, Rosenthal IM 1968 Intrauterine hyperparathyroidism secondary to maternal hypoparathyroidism. Pediatrics 42:606–613
- 247. **Grant DK** 1953 Papilloedema and fits in hypoparathyroidism. Q J Med 22:243–259
- 248. Wright AD, Joplin GF, Dixon HG 1969 Post-partum hypercalcaemia in treated hypoparathyroidism. Br Med J 1:23–25
- Blickstein I, Kessler I, Lancet M 1985 Idiopathic hypoparathyroidism with gestational diabetes. Am J Obstet Gynecol 153:649– 650
- 250. **Redell G** 1946 Parathyroprival tetany and pregnancy. Acta Obstet Gynecol Scand 26:1–10
- Blohm RW, Wurl OA, Gillespie JO, Escamilla RF 1953 Refractoriness to antitetanic therapy in a case of surgical hypoparathyroidism. J Clin Endocrinol Metab 13:519–533
- 252. Graham III WP, Gordon CS, Loken HF, Blum A, Halden A 1964 Effect of pregnancy and of the menstrual cycle on hypoparathyroidism. J Clin Endocrinol Metab 24:512–516
- 253. Markestad T, Ulstein M, Bassoe HH, Aksnes L, Aarskog D 1983 Vitamin D metabolism in normal and hypoparathyroid pregnancy and lactation. Case report. Br J Obstet Gynaecol 90:971–976
- Caplan RH, Beguin EA 1990 Hypercalcemia in a calcitriol-treated hypoparathyroid woman during lactation. Obstet Gynecol 76:485– 489
- 255. **Sadeghi-Nejad A, Wolfsdorf JI, Senior B** 1980 Hypoparathyroidism and pregnancy. Treatment with calcitriol. J Am Med Assoc 243:254–255
- 256. Salle BL, Berthezene F, Glorieux FH, Delvin EE, Berland M, David L, Varenne JP, Putet G 1981 Hypoparathyroidism during pregnancy: treatment with calcitriol. J Clin Endocrinol Metab 52: 810-813

- 257. Kurzel RB, Hagen GA 1990 Use of thiazide diuretics to reduce the hypercalciuria of hypoparathyroidism during pregnancy. Am J Perinatol 7:333–336
- Caplan RH, Wickus GG 1993 Reduced calcitriol requirements for treating hypoparathyroidism during lactation. A case report. J Reprod Med 38:914–918
- Levine MA, Spiegel AM 1995 Pseudohypoparathyroidism. In: De-Groot LJ (ed) Endocrinology. W.B. Saunders, Philadelphia, pp 1136–1150
- 260. Breslau NA, Zerwekh JE 1986 Relationship of estrogen and pregnancy to calcium homeostasis in pseudohypoparathyroidism. J Clin Endocrinol Metab 62:45–51
- 261. Zerwekh JE, Breslau NA 1986 Human placental production of 1α,25-dihydroxyvitamin D₃: biochemical characterization and production in normal subjects and patients with pseudohypoparathyroidism. J Clin Endocrinol Metab 62:192–196
- 262. Vidailhet M, Monin P, Andre M, Suty Y, Marchal C, Vert P 1980 [Neonatal hyperparathyroidism secondary to maternal hypoparathyroidism]. Arch Fr Pediatr 37:305–312
- 263. **Glass EJ, Barr DG** 1981 Transient neonatal hyperparathyroidism secondary to maternal pseudohypoparathyroidism. Arch Dis Child 56:565–568
- 264. Kirby DR, Bardbury S 1965 The hemo-chorial mouse placenta. Anat Rec 152:279–282
- 265. Enders AC 1965 A comparative study of the fine structures of the trophoblast in several hemochorial placentas. Am J Anat 116:29–68
- 266. **Ramsey EM** 1982 The Placenta: Human and Animal. Praeger, New York, p 187
- 267. Robinson NR, Atkinson DE, Jones CJ, Sibley CP 1988 Permeability of the near-term rat placenta to hydrophilic solutes. Placenta 9:361–372
- 268. David L, Anast CS 1974 Calcium metabolism in newborn infants. The interrelationship of parathyroid function and calcium, magnesium, and phosphorus metabolism in normal, sick, and hypocalcemic newborns. J Clin Invest 54:287–296
- 269. Garel JM, Barlet JP 1976 Calcium metabolism in newborn animals: the interrelationship of calcium, magnesium, and inorganic phosphorus in newborn rats, foals, lambs, and calves. Pediatr Res 10: 749–754
- Schauberger CW, Pitkin RM 1979 Maternal-perinatal calcium relationships. Obstet Gynecol 53:74–76
- 271. Fleischman AR, Lerman S, Oakes GK, Epstein MF, Chez RA, Mintz DH 1975 Perinatal primate parathyroid hormone metabolism. Biol Neonate 27:40–49
- 272. Schedewie HK, Odell WD, Fisher DA, Krutzik SR, Dodge M, Cousins L, Fiser WP 1979 Parathormone and perinatal calcium homeostasis. Pediatr Res 13:1–6
- 273. Pitkin RM, Cruikshank DP, Schauberger CW, Reynolds WA, Williams GA, Hargis GK 1980 Fetal calcitropic hormones and neonatal calcium homeostasis. Pediatrics 66:77–82
- 274. Delivoria-Papadopoulos M, Battaglia FC, Bruns PD, Meschia G 1967 Total, protein-bound, and ultrafilterable calcium in maternal and fetal plasmas. Am J Physiol 213:363–366
- 275. Wadsworth JC, Kronfeld DS, Ramberg Jr CF 1982 Parathyrin and calcium homeostasis in the fetus. Biol Neonate 41:101–109
- Math F, Davrainville JL 1979 Postnatal variations of extracellular free calcium levels in the rat. Influence of undernutrition. Experientia 35:1355–1356
- Thomas ML, Anast CS, Forte LR 1981 Regulation of calcium homeostasis in the fetal and neonatal rat. Am J Physiol 240:E367–E372
- Krukowski M, Smith JJ 1976 pH and the level of calcium in the blood of fetal and neonatal albino rats. Biol Neonate 29:148–161
- 279. Krukowski M, Smith JJ 1979 Acidosis, hypercalcemia, and hyperphosphatemia in rat fetuses near term and effects of maternal acid/base loading. Proc Soc Exp Biol Med 162:359–364
- Caple IW, Heath JA, Care AD, Heaton C, Farrugia W, Wark JD 1988 The regulation of osteoblast activity in fetal lambs by placental calcium transport and the fetal parathyroid glands. In: Jones CT (ed) Fetal and Neonatal Development. Perinatal Press, Ithaca, NY, pp 90–93
- 281. MacIsaac RJ, Heath JA, Rodda CP, Moseley JM, Care AD, Martin TJ, Caple IW 1991 Role of the fetal parathyroid glands and parathyroid hormone-related protein in the regulation of placental

transport of calcium, magnesium and inorganic phosphate. Reprod Fertil Dev 3:447–457

- 282. Moniz CF, Nicolaides KH, Tzannatos C, Rodeck CH 1986 Calcium homeostasis in second trimester fetuses. J Clin Pathol 39:838–841
- 283. Delvin EE, Glorieux FH, Salle BL, David L, Varenne JP 1982 Control of vitamin D metabolism in preterm infants: feto-maternal relationships. Arch Dis Child 57:754–757
- 284. Lima MS, Kallfelz F, Krook L, Nathanielsz PW 1993 Humeral skeletal development and plasma constituent changes in fetuses of ewes maintained on a low calcium diet from 60 days of gestation. Calcif Tissue Int 52:283–290
- 285. Miller SC, Halloran BP, DeLuca HF, Jee WS 1983 Studies on the role of vitamin D in early skeletal development, mineralization, and growth in rats. Calcif Tissue Int 35:455–460
- Chalon S, Garel JM 1985 Plasma calcium control in the rat fetus. II. Influence of fetal hormones. Biol Neonate 48:323–328
- 287. **Pic P** 1968 [Maintenance of an elevated fetal blood calcium in the absence of maternal and fetal parathyroid glands in rats]. C R Seances Soc Biol Fil 162:1043–1047
- Kovacs CS, Ho C, Seidman CE, Seidman JG, Kronenberg HM 1996 Parathyroid calcium sensing receptor regulates fetal blood calcium and fetal-maternal calcium gradient independently of the maternal calcium level. J Bone Miner Res 11[Suppl 1]:S121 (Abstract)
- Burnette JC, Simpson DM, Chandler Jr DC, Bawden JW 1968 Fetal blood calcium response to maternal parathyroid and vitamin D administration in guinea pigs. J Dent Res 47:444–446
- Krukowski M, Lehr D 1963 Parathyroid hormone and the placental barrier. Arch Int Pharmacodyn Ther 146:245–265
- 291. Garel JM, Pic P, Jost A 1971 Action de la parathormone chez de foetus de rat. Ann Endocrinol (Paris) 32:253–262
- 292. **Reynolds WA, Pitkin RM, Wezeman FH** 1975 Calcitonin effects in primate pregnancy. Am J Obstet Gynecol 122:212–218
- 293. **Stulc J, Stulcova B** 1996 Placental transfer of phosphate in anaesthetized rats. Placenta 17:487–493
- 294. Barlet JP, Davicco MJ, Rouffet J, Coxam V, Lefaivre J 1994 Short communication: parathyroid hormone-related peptide does not stimulate phosphate placental transport. Placenta 15:441–444
- 295. Moore ES, Langman CB, Favus MJ, Coe FL 1985 Role of fetal 1,25-dihydroxyvitamin D production in intrauterine phosphorus and calcium homeostasis. Pediatr Res 19:566–569
- 296. Leroyer-Alizon E, David L, Anast CS, Dubois PM 1981 Immunocytological evidence for parathyroid hormone in human fetal parathyroid glands. J Clin Endocrinol Metab 52:513–516
- 297. Weatherley ÅJ, Ross R, Pickard DW, Care AD 1983 The transfer of calcium during perfusion of the placenta and intact and thyroparathyroidectomized sheep. Placenta 4:271–277
- 298. Care AD, Caple IW, Abbas SK, Pickard DW 1986 The effect of fetal thyroparathyroidectomy on the transport of calcium across the ovine placenta to the fetus. Placenta 7:417–424
- 299. Kovacs CS, Lanske B, Hunzelman JL, Guo J, Karaplis AC, Kronenberg HM 1996 Parathyroid hormone-related peptide (PTHrP) regulates fetal-placental calcium transport through a receptor distinct from the PTH/PTHrP receptor. Proc Natl Acad Sci USA 93:15233–15238
- 300. Smith Jr FG, Alexander DP, Buckle RM, Britton HG, Nixon DA 1972 Parathyroid hormone in foetal and adult sheep: the effect of hypocalcaemia. J Endocrinol 53:339–348
- 301. Pitkin RM, Reynolds WA, Williams GA, Kawahara W, Bauman AF, Hargis GK 1980 Maternal and fetal parathyroid hormone responsiveness in pregnant primates. J Clin Endocrinol Metab 51: 1044–1047
- 302. **Dorey LG, Gell JW** 1975 Primary hyperparathyroidism during the third trimester of pregnancy. Obstet Gynecol 45:469–472
- 303. Rubin LP, Posillico JT, Anast CS, Brown EM 1991 Circulating levels of biologically active and immunoreactive intact parathyroid hormone in human newborns. Pediatr Res 29:201–207
- 304. Seki K, Furuya K, Makimura N, Mitsui C, Hirata J, Nagata I 1994 Cord blood levels of calcium-regulating hormones and osteocalcin in premature infants. J Perinat Med 22:189–194
- 305. Fairney A, Jackson D, Clayton BE 1973 Measurement of serum parathyroid hormone, with particular reference to some infants with hypocalcaemia. Arch Dis Child 48:419-424

- 306. Tsang RC, Chen IW, Friedman MA, Chen I 1973 Neonatal parathyroid function: role of gestational age and postnatal age. J Pediatr 83:728–738
- 307. Seino Y, Ishida M, Yamaoka K, Ishii T, Hiejima T, Ikehara C, Tanaka Y, Matsuda S, Shimotsuji T, Yabuuchi H, Morimoto S, Onishi T 1982 Serum calcium regulating hormones in the perinatal period. Calcif Tissue Int 34:131–135
- 308. Papantoniou NE, Papapetrou PD, Antsaklis AJ, Kontoleon PE, Mesogitis SA, Aravantinos D 1996 Circulating levels of immunoreactive parathyroid hormone-related protein and intact parathyroid hormone in human fetuses and newborns. Eur J Endocrinol 134:437–442
- Collignon H, Davicco MJ, Barlet JP 1996 Calcitonin mRNA expression and plasma calciotropic hormones in fetal lambs. Domest Anim Endocrinol 13:269–276
- Seki K, Wada S, Nagata N, Nagata I 1994 Parathyroid hormonerelated protein during pregnancy and the perinatal period. Gynecol Obstet Invest 37:83–86
- Glazier JD, Mawer EB, Sibley CP 1995 Calbindin-D_{9K} gene expression in rat chorioallantoic placenta is not regulated by 1,25dihydroxyvitamin D₃. Pediatr Res 37:720–725
- 312. Verhaeghe J, Thomasset M, Brehier A, Van Assche FA, Bouillon R 1988 1,25(OH)₂D₃ and Ca-binding protein in fetal rats: relationship to the maternal vitamin D status. Am J Physiol 254:E505–E512
- 313. Yoshizawa T, Handa Y, Uematsu Y, Sekine K, Takeda S, Yoshihara Y, Kawakami T, Sato H, Alioka K, Tanimoto K, Fukamizu A, Masushige S, Matsumoto T, Kato S 1996 Disruption of the vitamin D receptor (VDR) in the mouse [abstract]. J Bone Miner Res 11[Suppl 1]:S124
- 314. Li YC, Pirro AE, Amling M, Delling G, Baron R, Bronson R, Demay MB 1997 Targeted ablation of the vitamin D receptor: an animal model of vitamin D dependent rickets type II with alopecia. Proc Natl Acad Sci USA 94:9831–9835
- Campbell DE, Fleischman AR 1988 Rickets of prematurity: controversies in causation and prevention. Clin Perinatol 15:879–890
- Pereira GR, Zucker AH 1986 Nutritional deficiencies in the neonate. Clin Perinatol 13:175–189
- Specker BL 1994 Do North American women need supplemental vitamin D during pregnancy or lactation? Am J Clin Nutr 59[Suppl]:484S–490S
- Johnson JA, Grande JP, Roche PC, Kumar R 1996 Ontogeny of the 1,25-dihydroxyvitamin D₃ receptor in fetal rat bone. J Bone Miner Res 11:56–61
- 319. **Durand D, Barlet JP, Braithwaite GD** 1983 The influence of 1,25dihydroxycholecalciferol on the mineral content of foetal guinea pigs. Reprod Nutr Dev 23:235–244
- 320. Noff D, Edelstein S 1978 Vitamin D and its hydroxylated metabolites in the rat. Placental and lacteal transport, subsequent metabolic pathways and tissue distribution. Horm Res 9:292–300
- 321. Hollis BW, Pittard WB 1984 Evaluation of the total fetomaternal vitamin D relationships at term: evidence for racial differences. J Clin Endocrinol Metab 59:652–657
- 322. Haddad Jr JG, Boisseau V, Avioli LV 1971 Placental transfer of vitamin D_3 and 25-hydroxycholecalciferol in the rat. J Lab Clin Med 77:908–915
- 323. Leroyer-Alizon E, David L, Dubois PM 1980 Evidence for calcitonin in the thyroid gland of normal and anencephalic human fetuses: immunocytological localization, radioimmunoassay, and gel filtration of thyroid extracts. J Clin Endocrinol Metab 50:316– 321
- 324. Garel JM, Care AD, Barlet JP 1974 A radioimmunoassay for ovine calcitonin: an evaluation of calcitonin secretion during gestation, lactation and foetal life. J Endocrinol 62:497–509
- 325. Garel JM, Milhaud G, Sizonenko P 1969 [Thyrocalcitonin and the placental barrier in rats]. C R Acad Sci Hebd Seances Acad Sci D 269:1785–1787
- 326. Garel JM, Barlet JP 1978 Calcitonin in the mother, fetus and newborn. Ann Biol Anim Biochim Biophys 18:53–68
- 327. Garel JM, Milhaud G, Jost A 1968 [Hypocalcemic and hypophosphatemic action of thyrocalcitonin in fetal rats]. C R Acad Sci Hebd Seances Acad Sci D 267:344–347
- 328. Allgrove J, Manning RM, Adami S, Chayen J, O'Riordan JL 1981

Biologically active parathyroid hormone in foetal and maternal plasma. Clin Sci 60:11P (Abstract)

- 329. Abbas SK, Ratcliffe WA, Moniz C, Dixit M, Caple IW, Silver M, Fowden A, Care AD 1994 The role of parathyroid hormone-related protein in calcium homeostasis in the fetal pig. Exp Physiol 79: 527–536
- Care AD 1989 Development of endocrine pathways in the regulation of calcium homeostasis. Baillieres Clin Endocrinol Metab 3:671–688
- 331. Abbas SK, Pickard DW, Illingworth D, Storer J, Purdie DW, Moniz C, Dixit M, Caple IW, Ebeling PR, Rodda CP 1990 Measurement of parathyroid hormone-related protein in extracts of fetal parathyroid glands and placental membranes. J Endocrinol 124:319–325
- Lee K, Deeds JD, Segre GV 1995 Expression of parathyroid hormone-related peptide and its receptor messenger ribonucleic acids during fetal development of rats. Endocrinology 136:453–463
- 333. Karmali R, Schiffmann SN, Vanderwinden JM, Hendy GN, Nys-DeWolf N, Corvilain J, Bergmann P, Vanderhaeghen JJ 1992 Expression of mRNA of parathyroid hormone-related peptide in fetal bones of the rat. Cell Tissue Res 270:597–600
- 334. Bowden SJ, Emly JF, Hughes SV, Powell G, Ahmed A, Whittle MJ, Ratcliffe JG, Ratcliffe WA 1994 Parathyroid hormone-related protein in human term placenta and membranes. J Endocrinol 142:217–224
- 335. Philbrick WM, Wysolmerski JJ, Galbraith S, Holt E, Orloff JJ, Yang KH, Vasavada RC, Weir EC, Broadus AE, Stewart AF 1996 Defining the roles of parathyroid hormone-related protein in normal physiology. Physiol Rev 76:127–173
- 336. Karaplis AC, Luz A, Glowacki J, Bronson RT, Tybulewicz VL, Kronenberg HM, Mulligan RC 1994 Lethal skeletal dysplasia from targeted disruption of the parathyroid hormone-related peptide gene. Genes Dev 8:277–289
- 337. Wysolmerski JJ, Dunbar M, Karaplis AC, Broadus AE, Philbrick WM 1996 PTHrP is necessary for mammary epithelial development. J Bone Miner Res 11:S113 (Abstract)
- 338. Rodda CP, Kubota M, Heath JA, Ebeling PR, Moseley JM, Care AD, Caple IW, Martin TJ 1988 Evidence for a novel parathyroid hormone-related protein in fetal lamb parathyroid glands and sheep placenta: comparisons with a similar protein implicated in humoral hypercalcaemia of malignancy. J Endocrinol 117:261–271
- 339. Care AD 1991 The placental transfer of calcium. J Dev Physiol 15:253–257
- 340. **Garel JM** 1987 Hormonal control of calcium metabolism during the reproductive cycle in mammals. Physiol Rev 67:1–66
- Fisher GJ, Kelley LK, Smith CH 1987 ATP-dependent calcium transport across basal plasma membranes of human placental trophoblast. Am J Physiol 252:C38–C46
- 342. Borke JL, Caride A, Verma AK, Kelley LK, Smith CH, Penniston JT, Kumar R 1989 Calcium pump epitopes in placental trophoblast basal plasma membranes. Am J Physiol 257:C341–C346
- 343. **Bruns ME, Fausto A, Avioli LV** 1978 Placental calcium binding protein in rats. Apparent identity with vitamin D-dependent calcium binding protein from rat intestine. J Biol Chem 253:3186–3190
- 344. **Symonds HW**, **Sansom BF**, **Twardock AR** 1972 The measurement of the transfer of calcium and phosphorus from foetus to dam in the sheep using a whole body counter. Res Vet Sci 13:272–275
- 345. Ramberg Jr CF, Delivoria-Papadopoulos M, Crandall ED, Kronfeld DS 1973 Kinetic analysis of calcium transport across the placenta. J Appl Physiol 35:682–688
- 346. Care AD, Dutton A, Mott JC, Robinson JS, Ross R 1979 Studies of the transplacental calcium gradient in sheep [abstract]. J Physiol (Lond) 290:19P–20P
- 347. **Care AD** 1980 Calcium homeostasis in the fetus. J Dev Physiol 2:85–99
- 348. **Chalon S, Garel JM** 1985 Plasma calcium control in the rat fetus. III. Influence of alterations in maternal plasma calcium on fetal plasma calcium level. Biol Neonate 48:329–335
- 349. **Barlet JP** 1985 Calcitonin may modulate placental transfer of calcium in ewes. J Endocrinol 104:17–21
- 350. **Durand D, Barlet JP** 1981 Influence de la calcitonine maternelle sur la calcémie foetale chez la brebis. Ann Endocrinol (Paris) 42:8C

- 351. **Barlet JP** 1985 Prolactin and calcium metabolism in pregnant ewes. J Endocrinol 107:171–175
- 352. Shamley DR, Veale G, Pettifor JM, Buffenstein R 1996 Trophoblastic giant cells of the mouse placenta contain calbindin- D_{9K} but not the vitamin D receptor. J Endocrinol 150:25–32
- 353. **Ross R, Florer J, Halbert K, McIntyre L** 1989 Characterization of 1,25-dihydroxyvitamin D₃ receptors and *in vivo* targeting of [³H]-1,25(OH)₂D₃ in the sheep placenta. Placenta 10:553–567
- 354. Tanamura A, Nomura S, Kurauchi O, Furui T, Mizutani S, Tomoda Y 1995 Purification and characterization of 1,25(OH)₂D₃ receptor from human placenta. J Obstet Gynaecol 21:631–639
- 355. **Stumpf WE, Sar M, Narbaitz R, Huang S, DeLuca HF** 1983 Autoradiographic localization of 1,25-dihydroxyvitamin D₃ in rat placenta and yolk sac. Horm Res 18:215–220
- 356. **Chalon S, Garel JM** 1983 1,25-Dihydroxyvitamin D_3 injections into rat fetuses: effects on fetal plasma calcium, plasma phosphate and mineral content. Reprod Nutr Dev 23:567–573
- 357. **Durand D, Braithwaite GD, Barlet JP** 1983 The effect of 1α-hydroxycholecalciferol on the placental transfer of calcium and phosphate in sheep. Br J Nutr 49:475–480
- 358. Barlet JP, Davicco MJ, Coxam V 1992 Calcitonin modulates parathyroid hormone related peptide-stimulated calcium placental transfer. In: Cohn DV, Gennari C (eds) Calcium Regulating Hormones and Bone Metabolism. Elsevier, Amsterdam, pp 124–128
- Robinson NR, Sibley CP, Mughal MZ, Boyd RD 1989 Fetal control of calcium transport across the rat placenta. Pediatr Res 26:109–115
- 360. Shaw AJ, Mughal MZ, Maresh MJ, Sibley CP 1991 Effects of two synthetic parathyroid hormone-related protein fragments on maternofetal transfer of calcium and magnesium and release of cyclic AMP by the in-situ perfused rat placenta. J Endocrinol 129:399–404
- 361. Barri M, Abbas SK, Pickard DW, Hammonds RG, Wood WI, Caple IW, Martin TJ, Care AD 1990 Fetal magnesium homeostasis in the sheep. Exp Physiol 75:681–688
- 362. Abbas SK, Pickard DW, Rodda CP, Heath JA, Hammonds RG, Wood WI, Caple IW, Martin TJ, Care AD 1989 Stimulation of ovine placental calcium transport by purified natural and recombinant parathyroid hormone-related protein (PTHrP) preparations. Q J Exp Physiol 74:549–552
- 363. Care AD, Abbas SK, Pickard DW, Barri M, Drinkhill M, Findlay JB, White IR, Caple IW 1990 Stimulation of ovine placental transport of calcium and magnesium by mid-molecule fragments of human parathyroid hormone-related protein. Exp Physiol 75:605– 608
- 364. Wu TL, Vasavada RC, Yang K, Massfelder T, Ganz M, Abbas SK, Care AD, Stewart AF 1996 Structural and physiologic characterization of the mid-region secretory species of parathyroid hormone-related protein. J Biol Chem 271:24371–24381
- 365. Davicco MJ, Coxam V, Lefaivre J, Barlet JP 1992 Parathyroid hormone-related peptide increases urinary phosphate excretion in fetal lambs. Exp Physiol 77:377–383
- 366. MacIsaac RJ, Horne RS, Caple IW, Martin TJ, Wintour EM 1993 Effects of thyroparathyroidectomy, parathyroid hormone, and PTHrP on kidneys of ovine fetuses. Am J Physiol 264:E37–E44
- 367. Brace RA 1994 Amniotic fluid dynamics. In: Creasy RK, Resnik R (eds) Maternal-Fetal Medicine: Principles and Practice. W.B. Saunders, Philadelphia, pp 106–114
- Rebut-Bonneton C, Garel JM, Delbarre F 1983 Parathyroid hormone, calcitonin, 1,25-dihydroxycholecalciferol, and basal bone resorption in the rat fetus. Calcif Tissue Int 35:183–189
- Rebut-Bonneton C, Demignon J, Amor B, Miravet L 1983 Effect of calcitonin in pregnant rats on bone resorption in fetuses. J Endocrinol 99:347–353
- Garel JM, Geloso-Meyer A 1971 [Fetal hyperparathyroidism in rats following maternal hypoparathyroidism]. Rev Eur Etud Clin Biol 16:174–178
- 371. Aaron JE, Makins NB, Caple IW, Abbas SK, Pickard DW, Care AD 1989 The parathyroid glands in the skeletal development of the ovine foetus. Bone Miner 7:13–22
- 372. Aaron JE, Abbas SK, Colwell A, Eastell R, Oakley BA, Russell RG, Care AD 1992 Parathyroid gland hormones in the skeletal development of the ovine foetus: the effect of parathyroidectomy with calcium and phosphate infusion. Bone Miner 16:121–129
- 373. Lanske B, Karaplis AC, Lee K, Luz A, Vortkamp A, Pirro A,

Karperien M, Defize L, Ho C, Abou-Samra AB, Jüppner H, Segre GV, Kronenberg HM 1996 PTH/PTHrP receptor in early development and Indian hedgehog-regulated bone growth. Science 273: 663–666

- 374. Page LA, Haddow JE 1987 Self-limited neonatal hyperparathyroidism in familial hypocalciuric hypercalcemia. J Pediatr 111:261– 264
- 375. Stuart C, Aceto Jr T, Kuhn JP, Terplan K 1979 Intrauterine hyperparathyroidism. Postmortem findings in two cases. Am J Dis Child 133:67–70
- 376. Loughead JL, Mughal Z, Mimouni F, Tsang RC, Oestreich AE 1990 Spectrum and natural history of congenital hyperparathyroidism secondary to maternal hypocalcemia. Am J Perinatol 7: 350–355
- 377. Sann L, David L, Thomas A, Frederich A, Chapuy MC, Francois R 1976 Congenital hyperparathyroidism and vitamin D deficiency secondary to maternal hypoparathyroidism. Acta Paediatr Scand 65:381–385
- 378. Aceto Jr T, Batt RE, Bruck E, Schultz RB, Perz YR 1966 Intrauterine hyperparathyroidism: a complication of untreated maternal hypoparathyroidism. J Clin Endocrinol Metab 26:487–492
- 379. Eastell R, Edmonds CJ, de Chayal RC, McFadyen IR 1985 Prolonged hypoparathyroidism presenting eventually as second trimester abortion. Br Med J (Clin Res Ed) 291:955–956
- 380. Anderson GW, Musselman L 1942 The treatment of tetany in pregnancy. Am J Obstet Gynecol 43:547–567
- Kehrer E 1913 Die geburtschilflich-gynäkologische bedeutung der tetanie. Arch Gynaek 99:372–447
- Savani RC, Mimouni F, Tsang RC 1993 Maternal and neonatal hyperparathyroidism as a consequence of maternal renal tubular acidosis. Pediatrics 91:661–663
- 383. Levin TL, States L, Greig A, Goldman HS 1992 Maternal renal insufficiency: a cause of congenital rickets and secondary hyperparathyroidism. Pediatr Radiol 22:315–316
- 384. Hammond VE, Senior PV, Tucci J, Beck F 1996 Functional analysis of the parathyroid hormone related protein by gene targeting. J Bone Miner Res 11 [Suppl 1]:S199 (Abstract)
- 385. Brown EM, Pollak M, Seidman CE, Seidman JG, Chou YH, Riccardi D, Hebert SC 1995 Calcium-ion-sensing cell-surface receptors. N Engl J Med 333:234–240
- Chattopadhyay N, Mithal A, Brown EM 1996 The calcium-sensing receptor: a window into the physiology and pathophysiology of mineral ion metabolism. Endocr Rev 17:289–307
- 387. **Bagnoli F, Bruchi S, Garosi G, Pecciarini L, Bracci R** 1990 Relationship between mode of delivery and neonatal calcium homeostasis. Eur J Pediatr 149:800–803
- 388. Kovacs CS, Lanske B, Byrne M, Krane SM, Kronenberg HM 1997 Altered interstitial collagenase expression in the tibias of PTHrP gene-ablated and PTH/PTHrP receptor gene-ablated fetal mice. J Bone Miner Res 12[Suppl 1]:S116 (Abstract)
- 389. Macy I, Kelly H, Sloan R 1953 The Composition of Milks; Publication No. 254. National Research Council, Washington, D.C., p 63
- 390. Sowers M 1996 Pregnancy and lactation as risk factors for subsequent bone loss and osteoporosis. J Bone Miner Res 11:1052–1060
- 391. Hunscher HA 1930 Metabolism of women during the reproductive cycle. II. Calcium and phosphorus utilization in two successive lactation periods. J Biol Chem 86:37–57
- 392. Laskey MA, Prentice A, Shaw J, Zachou T, Ceesay SM, Vasquez-Velasquez L, Fraser DR 1990 Breast-milk calcium concentrations during prolonged lactation in British and rural Gambian mothers. Acta Paediatr Scand 79:507–512
- 393. Neville MC, Keller RP, Seacat J, Casey CE, Allen JC, Archer P 1984 Studies on human lactation. I. Within-feed and between-breast variation in selected components of human milk. Am J Clin Nutr 40:635–646
- 394. Vaughan LA, Weber CW, Kemberling SR 1979 Longitudinal changes in the mineral content of human milk. Am J Clin Nutr 32:2301–2306
- 395. Karra MV, Udipi SA, Kirksey A, Roepke JL 1986 Changes in specific nutrients in breast milk during extended lactation. Am J Clin Nutr 43:495–503
- 396. Donelson E, Nims B, Hunscher HA, Macy IG 1931 Metabolism of women during the reproductive cycle. IV. Calcium and phospho-

- 397. Hillman L, Sateesha S, Haussler M, Wiest W, Slatopolsky E, Haddad J 1981 Control of mineral homeostasis during lactation: interrelationships of 25-hydroxyvitamin D, 24,25-dihydroxyvitamin D, 1,25-dihydroxyvitamin D, parathyroid hormone, calcitonin, prolactin, and estradiol. Am J Obstet Gynecol 139:471–476
- 398. Specker BL, Tsang RC, Ho ML 1991 Changes in calcium homeostasis over the first year postpartum: effect of lactation and weaning. Obstet Gynecol 78:56–62
- 399. Grill V, Hillary J, Ho PM, Law FM, MacIsaac RJ, MacIsaac IA, Moseley JM, Martin TJ 1992 Parathyroid hormone-related protein: a possible endocrine function in lactation. Clin Endocrinol (Oxf) 37:405–410
- 400. Dobnig H, Kainer F, Stepan V, Winter R, Lipp R, Schaffer M, Kahr A, Nocnik S, Patterer G, Leb G 1995 Elevated parathyroid hormone-related peptide levels after human gestation: relationship to changes in bone and mineral metabolism. J Clin Endocrinol Metab 80:3699–3707
- 401. Kovacs CS, Chik CL 1995 Hyperprolactinemia caused by lactation and pituitary adenomas is associated with altered serum calcium, phosphate, parathyroid hormone (PTH), and PTH-related peptide levels. J Clin Endocrinol Metab 80:3036–3042
- 402. Greer FR, Lane J, Ho M 1984 Elevated serum parathyroid hormone, calcitonin, and 1,25-dihydroxyvitamin D in lactating women nursing twins. Am J Clin Nutr 40:562–568
- Lepre F, Grill V, Ho PW, Martin TJ 1993 Hypercalcemia in pregnancy and lactation associated with parathyroid hormone-related protein [letter]. N Engl J Med 328:666–667
- 404. Toverud SU, Harper C, Munson PL 1976 Calcium metabolism during lactation: enhanced effects of thyrocalcitonin. Endocrinology 99:371–378
- 405. Wong KM, Singer L, Ophaug RH 1980 Metabolic aspects of bone resorption in calcium-deficient lactating rats. Calcif Tissue Int 32: 213–219
- 406. Blahosova A, Neradilova M, Velicky J, Titlbach M, Marsikova L, Reisenauer R 1974 Dynamics of changes of calcium and phosphorus metabolism in relation to the morphology of parafollicular thyroid cells in rats during lactation and forced weaning. Endokrinologie 63:122–136
- 407. Lobaugh B, Boass A, Garner SC, Toverud SU 1992 Intensity of lactation modulates renal 1α-hydroxylase and serum 1,25(OH)₂D in rats. Am J Physiol 262:E840–E844
- 408. **Peng TC, Garner SC, Kusy RP, Hirsch PF** 1988 Effect of number of suckling pups and dietary calcium on bone mineral content and mechanical properties of femurs of lactating rats. Bone Miner 3: 293–304
- 409. Hodnett DW, DeLuca HF, Jorgensen NA 1992 Intestine, bone, and mammary gland contributions to maternal plasma calcium increase after abrupt weaning. Proc Soc Exp Biol Med 199:332–336
- 410. Kent GN, Price RI, Gutteridge DH, Smith M, Allen JR, Bhagat CI, Barnes MP, Hickling CJ, Retallack RW, Wilson SG 1990 Human lactation: forearm trabecular bone loss, increased bone turnover, and renal conservation of calcium and inorganic phosphate with recovery of bone mass following weaning. J Bone Miner Res 5: 361–369
- 411. Lippuner K, Zehnder HJ, Casez JP, Takkinen R, Jaeger P 1996 PTH-related protein is released into the mother's bloodstream during location: evidence for beneficial effects on maternal calciumphosphate metabolism. J Bone Miner Res 11:1394–1399
- 412. Kalkwarf HJ, Specker BL, Heubi JE, Vieira NE, Yergey AL 1996 Intestinal calcium absorption of women during lactation and after weaning. Am J Clin Nutr 63:526–531
- 413. Retallack RW, Jeffries M, Kent GN, Hitchcock NE, Gutteridge DH, Smith M 1977 Physiological hyperparathyroidism in human lactation. Calcif Tissue Res 22[Suppl]:142–146
- 414. Greer FR, Tsang RC, Searcy JE, Levin RS, Steichen JJ 1982 Mineral homeostasis during lactation—relationship to serum 1,25-dihydroxyvitamin D, 25-hydroxyvitamin D, parathyroid hormone, and calcitonin. Am J Clin Nutr 36:431–437
- 415. Affinito P, Tommaselli GA, di Carlo C, Guida F, Nappi C 1996 Changes in bone mineral density and calcium metabolism in

breastfeeding women: a one year follow-up study. J Clin Endocrinol Metab 81:2314-2318

- 416. Zinaman MJ, Hickey M, Tomai TP, Albertson BD, Simon JA 1990 Calcium metabolism in postpartum lactation: the effect of estrogen status. Fertil Steril 54:465–469
- 417. Krebs NF, Reidinger CJ, Robertson AD, Brenner M 1997 Bone mineral density changes during lactation: maternal, dietary, and biochemical correlates. Am J Clin Nutr 65:1738–1746
- Fry JM, Curnow DH, Gutteridge DH, Retallack RW 1979 Effect of chronic parathyroidectomy on calcium metabolism in the lactating rat. J Endocrinol 82:323–330
- 419. Lobaugh B, Boass A, Lester GE, Toverud SU 1990 Regulation of serum 1,25-dihydroxyvitamin D₃ in lactating rats. Am J Physiol 259:E665–E671
- 420. Stevenson JC, Hillyard CJ, MacIntyre I, Cooper H, Whitehead MI 1979 A physiological role for calcitonin: protection of the maternal skeleton. Lancet 2:769–770
- Toverud SU, Cooper CW, Munson PL 1978 Calcium metabolism during lactation: elevated blood levels of calcitonin. Endocrinology 103:472–479
- 422. Toverud SU, Boass A 1979 Hormonal control of calcium metabolism in lactation. In: Munson PL, Glover J, Diczfalusy E, Olson RE (eds) Vitamins and Hormones. Academic Press, New York, pp 303–347
- 423. Uemura H, Yasui T, Yoneda N, Irahara M, Aono T 1997 Measurement of N- and C-terminal-region fragments of parathyroid hormone-related peptide in milk from lactating women and investigation of the relationship of their concentrations to calcium in milk. J Endocrinol 153:445–451
- 424. Law FM, Moate PJ, Leaver DD, Diefenbach-Jagger H, Grill V, Ho PW, Martin TJ 1991 Parathyroid hormone-related protein in milk and its correlation with bovine milk calcium. J Endocrinol 128: 21–26
- 425. Yamamoto M, Fisher JE, Thiede MA, Caulfield MP, Rosenblatt M, Duong LT 1992 Concentrations of parathyroid hormone-related protein in rat milk change with duration of lactation and interval from previous suckling, but not with milk calcium. Endocrinology 130:741–747
- Ratcliffe WA, Thompson GE, Care AD, Peaker M 1992 Production of parathyroid hormone-related protein by the mammary gland of the goat. J Endocrinol 133:87–93
- 427. Thiede MA, Grasser WA, Petersen DN 1992 Regulated expression of parathyroid hormone-related protein in mammary blood supply supports a role in mammary blood flow. In: Cohn DV, Gennari C, Tashjian Jr AH (eds) Calcium Regulating Hormones and Bone Metabolism: Basic and Clincal Aspects. Elsevier Science Publishers, New York, pp 62–66
- Thiede MA 1994 Parathyroid hormone-related protein: a regulated calcium-mobilizing product of the mammary gland. J Dairy Sci 77:1952–1963
- 429. **Thiede MA** 1989 The mRNA encoding a parathyroid hormone-like peptide is produced in mammary tissue in response to elevations in serum prolactin. Mol Endocrinol 3:1443–1447
- 430. Thompson GE, Ratcliffe WA, Hughes S, Abbas SK, Care AD 1994 Local control of parathyroid hormone-related protein secretion by the mammary gland of the goat. Comp Biochem Physiol A 108: 485–490
- 431. **Bucht E, Carlqvist M, Hedlund B, Bremme K, Torring O** 1992 Parathyroid hormone-related peptide in human milk measured by a mid-molecule radioimmunoassay. Metabolism 41:11–16
- 432. Yamamoto M, Duong LT, Fisher JE, Thiede MA, Caulfield MP, Rosenblatt M 1991 Suckling-mediated increases in urinary phosphate and 3',5'-cyclic adenosine monophosphate excretion in lactating rats: possible systemic effects of parathyroid hormone-related protein. Endocrinology 129:2614–2622
- 433. Barlet JP, Abbas SK, Care AD, Davicco MJ, Rouffet J 1993 Parathyroid hormone-related peptide and milking-induced phosphaturia in dairy cows. Acta Endocrinol (Copenh) 129:332–336
- 434. Melton ME, D'Anza JJ, Wimbiscus SA, Grill V, Martin TJ, Kukreja SC 1990 Parathyroid hormone-related protein and calcium homeostasis in lactating mice. Am J Physiol 259:E792–E796
- 435. Sowers MF, Hollis BW, Shapiro B, Randolph J, Janney CA, Zhang D, Schork A, Crutchfield M, Stanczyk F, Russell-Aulet M 1996

Elevated parathyroid hormone-related peptide associated with lactation and bone density loss. J Am Med Assoc 276:549–554

- 436. Lippuner K, Zehnder HJ, Casez JP, Takkinen R, Jaeger P 1995 Effects of PTH-related protein (PTH-rP) on calcium-phosphate metabolism in nursing mothers. Bone 16[Suppl 1]:209S (Abstract)
- 437. Caplan RH, Wickus GG, Sloane K, Silva PD 1995 Serum parathyroid hormone-related protein levels during lactation. J Reprod Med 40:216–218
- Reid IR, Wattie DJ, Evans MC, Budayr AA 1992 Post-pregnancy osteoporosis associated with hypercalcaemia. Clin Endocrinol (Oxf) 37:298–303
- Siskind MS, Popovtzer MM 1991 Postpartum hypercalcemia in a patient with medullary sponge kidneys. Am J Kidney Dis 17:588– 590
- 440. Khosla S, van Heerden JA, Gharib H, Jackson IT, Danks J, Hayman JA, Martin TJ 1990 Parathyroid hormone-related protein and hypercalcemia secondary to massive mammary hyperplasia (letter). N Engl J Med 322:1157
- 441. Specker BL, Vieira NE, O'Brien KO, Ho ML, Heubi JE, Abrams SA, Yergey AL 1994 Calcium kinetics in lactating women with low and high calcium intakes. Am J Clin Nutr 59:593–599
 442. Prentice A, Jarjou LM, Cole TJ, Stirling DM, Dibba B,
- 442. Prentice A, Jarjou LM, Cole TJ, Stirling DM, Dibba B, Fairweather-Tait S 1995 Calcium requirements of lactating Gambian mothers: effects of a calcium supplement on breast-milk calcium concentration, maternal bone mineral content, and urinary calcium excretion. Am J Clin Nutr 62:58–67
- 443. Prentice A 1994 Maternal calcium requirements during pregnancy and lactation. Am J Clin Nutr 59[Suppl]:477S-482S
- 444. Boass A, Toverud SU, Pike JW, Haussler MR 1981 Calcium metabolism during lactation: enhanced intestinal calcium absorption in vitamin D-deprived, hypocalcemic rats. Endocrinology 109:900– 907
- 445. Anderson JJ, Garner SC, Mar MH, Boass A, Toverud SU, Parikh I 1990 The ovariectomized, lactating rat as an experimental model for osteopenia: calcium metabolism and bone changes. Bone Miner 11:43–53
- 446. **Fournier P, Susbielle H** 1952 Les échanges de calcium chez le rat au cours de la gestation, de la lactation et du sevrage. I. Influence du régime riche en calcium. J Physiol Paris 44:123–134
- 447. Komárková A, Záhor Z, Czabanová V 1967 The effect of lactation on the composition of long bones in rats. J Lab Clin Med 69:102–109
- Miller MA, Omura TH, Miller SC 1989 Increased cancellous bone remodeling during lactation in beagles. Bone 10:279–285
- 449. Sowers M, Eyre D, Hollis BW, Randolph JF, Shapiro B, Jannausch ML, Crutchfield M 1995 Biochemical markers of bone turnover in lactating and nonlactating postpartum women. J Clin Endocrinol Metab 80:2210–2216
- 450. Hirsch PF, Hagaman JR 1986 Reduced bone mass in calcitonindeficient rats whether lactating or not. J Bone Miner Res 1:199–206
- 451. **Gruber HE, Stover SJ** 1994 Maternal and weanling bone: the influence of lowered calcium intake and maternal dietary history. Bone 15:167–176
- 452. Hagaman JR, Ambrose WW, Hirsch PF 1990 A scanning electron microscopic and photon absorptiometric study of the development, prolongation, and pattern of recovery from lactation-induced osteopenia in rats. J Bone Miner Res 5:123–132
- 453. Hodnett DW, DeLuca HF, Jorgensen NA 1992 Bone mineral loss during lactation occurs in absence of parathyroid tissue. Am J Physiol 262:E230–E233
- 454. Garner SC, Boass A, Toverud SU 1990 Parathyroid hormone is not required for normal milk composition or secretion or lactationassociated bone loss in normocalcemic rats. J Bone Miner Res 5: 69–75
- 455. Brommage R, DeLuca HF 1985 Regulation of bone mineral loss during lactation. Am J Physiol 248:E182–E187
- Atkinson PJ, West RR 1970 Loss of skeletal calcium in lactating women. J Obstet Gynaecol Br Commonw 77:555–560
- 457. Sorenson JA, Cameron JR 1967 A reliable *in vivo* measurement of bone mineral content. J Bone Joint Surg [Am] 49:481–497
- 458. Hayslip CC, Klein TA, Wray HL, Duncan WE 1989 The effects of lactation on bone mineral content in healthy postpartum women. Obstet Gynecol 73:588–592
- 459. Chan GM, Slater P, Ronald N, Roberts CC, Thomas MR, Folland

D, Jackson R 1982 Bone mineral status of lactating mothers of different ages. Am J Obstet Gynecol 144:438-441

- 460. Chan GM, Ronald N, Slater P, Hollis J, Thomas MR 1982 Decreased bone mineral status in lactating adolescent mothers. J Pediatr 101:767–770
- 461. Chan GM, Roberts CC, Folland D, Jackson R 1982 Growth and bone mineralization of normal breast-fed infants and the effects of lactation on maternal bone mineral status. Am J Clin Nutr 36:438– 443
- 462. Caird LE, Reid-Thomas V, Hannan WJ, Gow S, Glasier AF 1994 Oral progestogen-only contraception may protect against loss of bone mass in breast-feeding women. Clin Endocrinol (Oxf) 41:739– 745
- 463. Laskey MA, Prentice A 1997 Effect of pregnancy on recovery of lactational bone loss (letter). Lancet 349:1518–1519
- 464. Kalkwarf HJ, Specker BL 1995 Bone mineral loss during lactation and recovery after weaning. Obstet Gynecol 86:26–32
 465. Chan GM, McMurry M, Westover K, Engelbert-Fenton K,
- 465. Chan GM, McMurry M, Westover K, Engelbert-Fenton K, Thomas MR 1987 Effects of increased dietary calcium intake upon the calcium and bone mineral status of lactating adolescent and adult women. Am J Clin Nutr 46:319–323
- 466. Laskey MA, Prentice A, Jarjou LM, Beavan S 1996 Lactational changes in bone mineral of the lumbar spine are influenced by breast-milk output but not calcium intake, breast-milk calcium concentration, or vitamin-D receptor genotype. J Bone Miner Res 11:1815 (Abstract)
- 467. Sowers M, Corton G, Shapiro B, Jannausch ML, Crutchfield M, Smith ML, Randolph JF, Hollis B 1993 Changes in bone density with lactation. J Am Med Assoc 269:3130–3135
- 468. Revilla R, Revilla M, Hernandez ER, Villa LF, Varela L, Rico H 1995 Evidence that the loss of bone mass induced by GnRH agonists is not totally recovered. Maturitas 22:145–150
- 469. Roux C, Pelissier C, Listrat V, Kolta S, Simonetta C, Guignard M, Dougados M, Amor B 1995 Bone loss during gonadotropin releasing hormone agonist treatment and use of nasal calcitonin. Osteoporos Int 5:185–190
- 470. Fogelman I, Fentiman I, Hamed H, Studd JW, Leather AT 1994 Goserelin (Zoladex) and the skeleton. Br J Obstet Gynaecol 101[Suppl 10]:19–23
- 471. **Mukherjee T, Barad D, Turk R, Freeman R** 1996 A randomized, placebo-controlled study on the effect of cyclic intermittent etidronate therapy on the bone mineral density changes associated with six months of gonadotropin-releasing hormone agonist treatment. Am J Obstet Gynecol 175:105–109
- 472. **Taga M, Minaguchi H** 1996 Reduction of bone mineral density by gonadotropin-releasing hormone agonist, nafarelin, is not completely reversible at 6 months after the cessation of administration. Acta Obstet Gynecol Scand 75:162–165
- 473. Newhall-Perry K, Holloway L, Osburn L, Monroe SE, Heinrichs L, Henzl M, Marcus R 1995 Effects of a gonadotropin-releasing hormone agonist on the calcium-parathyroid axis and bone turn-over in women with endometriosis. Am J Obstet Gynecol 173:824–829
- 474. Orwoll ES, Yuzpe AA, Burry KA, Heinrichs L, Buttram Jr VC, Hornstein MD 1994 Nafarelin therapy in endometriosis: long-term effects on bone mineral density. Am J Obstet Gynecol 171:1221– 1225
- 475. **Rico H, Arnanz F, Revilla M, Perera S, Iritia M, Villa LF, Arribas** I 1993 Total and regional bone mineral content in women treated with GnRH agonists. Calcif Tissue Int 52:354–357
- 476. Paoletti AM, Serra GG, Cagnacci A, Vacca AM, Guerriero S, Solla E, Melis GB 1996 Spontaneous reversibility of bone loss induced by gonadotropin-releasing hormone analog treatment. Fertil Steril 65:707–710
- 477. Howell R, Edmonds DK, Dowsett M, Crook D, Lees B, Stevenson JC 1995 Gonadotropin-releasing hormone analogue (goserelin) plus hormone replacement therapy for the treatment of endometriosis: a randomized controlled trial. Fertil Steril 64:474–481
- 478. Uemura T, Mohri J, Osada H, Suzuki N, Katagiri N, Minaguchi H 1994 Effect of gonadotropin-releasing hormone agonist on the bone mineral density of patients with endometriosis. Fertil Steril 62:246–250
- 479. Eckstein N, Foldes J, Feinstein Y, Vagman I, Eshel A, Steinberg

R, **Statter M**, **Limor R**, **Ayalon D** 1992 Calcium homeostasis, bone metabolism and safety aspects during long-term treatment with a GnRH agonist. Maturitas 15:25–32

- 480. Stiegler C, Leb G, Kleinert R, Warnkross H, Ramschak-Schwarzer S, Lipp R, Clarici G, Krejs GJ, Dobnig H 1995 Plasma levels of parathyroid hormone-related peptide are elevated in hyperprolactinemia and correlated to bone density status. J Bone Miner Res 10:751–759
- 481. Sowers M, Randolph J, Shapiro B, Jannausch M 1995 A prospective study of bone density and pregnancy after an extended period of lactation with bone loss. Obstet Gynecol 85:285–289
- 482. Koetting CA, Wardlaw GM 1988 Wrist, spine, and hip bone density in women with variable histories of lactation. Am J Clin Nutr 48:1479–1481
- 483. Feldblum PJ, Zhang J, Rich LE, Fortney JA, Talmage RV 1992 Lactation history and bone mineral density among perimenopausal women. Epidemiology 3:527–531
- 484. Dequeker J, Tobing L, Rutten V, Geusens P 1991 Relative risk factors for osteoporotic fracture: a pilot study of the MEDOS questionnaire. Clin Rheumatol 10:49–53
- 485. Aloia JF, Cohn SH, Vaswani A, Yeh JK, Yuen K, Ellis K 1985 Risk factors for postmenopausal osteoporosis. Am J Med 78:95–100
- 486. Aloia JF, Vaswani AN, Yeh JK, Ross P, Ellis K, Cohn SH 1983 Determinants of bone mass in postmenopausal women. Arch Intern Med 143:1700–1704
- 487. Berning B, van Kuijk C, Schutte HE, Kuiper JW, Drogendijk AC, Fauser BC 1993 Determinants of lumbar bone mineral density in normal weight, non-smoking women soon after menopause. A study using clinical data and quantitative computed tomography. Bone Miner 21:129–139
- 488. Wardlaw GM, Pike AM 1986 The effect of lactation on peak adult shaft and ultra-distal forearm bone mass in women. Am J Clin Nutr 44:283–286
- Glasier A, McNeilly AS, Howie PW 1984 The prolactin response to suckling. Clin Endocrinol (Oxf) 21:109–116
- 490. Wasnich Ř, Yano K, Vogel J 1983 Postmenopausal bone loss at multiple skeletal sites: relationship to estrogen use. J Chronic Dis 36:781–790
- 491. Hiyaoka A, Yoshida T, Cho F, Yoshikawa Y 1996 Changes in bone mineral density of lumbar vertebrae after parturition in African green monkeys (*Cercopithecus aethiops*). Exp Anim 45:257–259
- 492. Yamamoto N, Takahashi HE, Tanizawa T, Kawashima T, Endo N 1994 Bone mineral density and bone histomorphometric assessments of postpregnancy osteoporosis: a report of five patients. Calcif Tissue Int 54:20–25
- 493. Ratcliffe WA 1992 Role of parathyroid hormone-related protein in lactation. Clin Endocrinol (Oxf) 37:402–404
- 494. Cathebras P, Cartry O, Sassolas G, Rousset H 1996 [Hypercalcemia induced by lactation in 2 patients with treated hypoparathyroidism]. Rev Med Interne 17:675–676
- 495. Nishioka T, Yasuda T, Niimi H, Nakajima H 1988 Evidence that calcitonin plays a role in the postnatal increase of serum 1α,25dihydroxyvitamin D. Eur J Pediatr 147:148–152
- Loughead JL, Mimouni F, Tsang RC 1988 Serum ionized calcium concentrations in normal neonates. Am J Dis Child 142:516–518
- 497. Martinez ME, Catalan P, Lisbona A, Sanchez-Cabezudo MJ, Pallardo F, Jans I, Bouillon R 1994 Serum osteocalcin concentrations in diabetic pregnant women and their newborns. Horm Metab Res 26:338–342
- 498. Nishioka T, Yasuda T, Niimi H 1991 A discordant movement in urine calcium excretion in relation to serum calcium and parathyroid function occurring immediately after birth. Acta Paediatr Scand 80:590–595
- 499. Loughead JL, Mimouni F, Tsang RC, Khoury JC 1991 A role for magnesium in neonatal parathyroid gland function? J Am Coll Nutr 10:123–126
- 500. **Mimouni F, Loughead JL, Tsang RC, Khoury J** 1990 Postnatal surge in serum calcitonin concentrations: no contribution to neonatal hypocalcemia in infants of diabetic mothers. Pediatr Res 28: 493–495
- 501. Loughead JL, Mimouni F, Ross R, Tsang RC 1990 Postnatal changes in serum osteocalcin and parathyroid hormone concentrations. J Am Coll Nutr 9:358–362

- 502. Tsang RC, Light IJ, Sutherland JM, Kleinman LI 1973 Possible pathogenetic factors in neonatal hypocalcemia of prematurity. The role of gestation, hyperphosphatemia, hypomagnesemia, urinary calcium loss, and parathormone responsiveness. J Pediatr 82: 423–429
- 503. Romagnoli C, Zecca E, Tortorolo G, Diodato A, Fazzini G, Sorcini-Carta M 1987 Plasma thyrocalcitonin and parathyroid hormone concentrations in early neonatal hypocalcaemia. Arch Dis Child 62:580–584
- 504. Venkataraman PS, Blick KE, Fry HD, Rao RK 1985 Postnatal changes in calcium-regulating hormones in very-low-birth-weight infants. Effect of early neonatal hypocalcemia and intravenous calcium infusion on serum parathyroid hormone and calcitonin homeostasis. Am J Dis Child 139:913–916
- 505. **Strewler GJ, Nissenson RA** 1993 Parathyroid hormone-related protein. In: Favus MJ (ed) Primer on the Metabolic Bone Diseases and Disorders of Mineral Metabolism. Raven Press, New York, pp 61–63
- 506. **Major BJ, Ho PM, Moseley JM, Martin TJ, Leaver DD** 1995 Circulating levels of parathyroid hormone related protein in neonates. Bone 16[Suppl]:204S (Abstract)
- 507. Rong H, Hydbring E, Olsson K, Burtis WJ, Rankin W, Grill V, Bucht E 1997 Parathyroid hormone-related protein in neonatal and reproductive goats determined by a sensitive time-resolved immunofluorometric assay. Eur J Endocrinol 136:546–551
- 508. Kukreja SC, D'Anza JJ, Melton ME, Wimbiscus SA, Grill V, Martin TJ 1991 Lack of effects of neutralization of parathyroid hormone-related protein on calcium homeostasis in neonatal mice. J Bone Miner Res 6:1197–1201
- 509. Halbert KE, Tsang RC 1992 Neonatal calcium, phosphorus, and magnesium homeostasis. In: Polin RA, Fox WW (eds) Fetal and Neonatal Physiology. W.B. Saunders, Philadelphia, pp 1745–1761
- 510. Ghishan FK, Parker P, Nichols S, Hoyumpa A 1984 Kinetics of intestinal calcium transport during maturation in rats. Pediatr Res 18:235–239
- 511. Ghishan FK, Jenkins JT, Younoszai MK 1980 Maturation of calcium transport in the rat small and large intestine. J Nutr 110: 1622–1628
- 512. Halloran BP, DeLuca HF 1980 Calcium transport in small intestine during early development: role of vitamin D. Am J Physiol 239: G473–G479
- 513. Buchowski MS, Miller DD 1991 Lactose, calcium source and age affect calcium bioavailability in rats. J Nutr 121:1746–1754
- 514. **Pansu D, Chapuy MC, Milani M, Bellaton C** 1976 Transepithelial calcium transport enhanced by xylose and glucose in the rat jejunal ligated loop. Calcif Tissue Res 21[Suppl]:45–52
- 515. Leichter J, Tolensky AF 1975 Effect of dietary lactose on the absorption of protein, fat and calcium in the postweaning rat. Am J Clin Nutr 28:238–241
- 516. Halloran BP, DeLuca HF 1981 Appearance of the intestinal cytosolic receptor for 1,25-dihydroxyvitamin D₃ during neonatal development in the rat. J Biol Chem 256:7338–7342
- 517. Bruns ME, Bruns DE, Avioli L 1979 Vitamin D-dependent calciumbinding protein of rat intestine: changes during postnatal development and sensitivity to 1,25-dihydroxycholecalciferol. Endocrinology 105:934–938
- 518. Giles MM, Fenton MH, Shaw B, Elton RA, Clarke M, Lang M, Hume R 1987 Sequential calcium and phosphorus balance studies in preterm infants. J Pediatr 110:591–598
- 519. Shaw JC 1976 Evidence for defective skeletal mineralization in low-birthweight infants: the absorption of calcium and fat. Pediatrics 57:16–25
- 520. **Senterre J, Salle B** 1982 Calcium and phosphorus economy of the preterm infant and its interaction with vitamin D and its metabolites. Acta Paediatr Scand Suppl 296:85–92
- 521. Barltrop D, Mole RH, Sutton A 1977 Absorption and endogenous faecal excretion of calcium by low birthweight infants on feeds with varying contents of calcium and phosphate. Arch Dis Child 52: 41–49
- 522. Kobayashi A, Kawai S, Obe Y, Nagashima Y 1975 Effects of dietary lactose and lactase preparation on the intestinal absorption of calcium and magnesium in normal infants. Am J Clin Nutr 28:681–683
- 523. Kocian J, Skala I, Bakos K 1973 Calcium absorption from milk and

lactose-free milk in healthy subjects and patients with lactose intolerance. Digestion 9:317–324

- 524. Hollis BW, Lowery JW, Pittard WB, Guy DG, Hansen JW 1996 Effect of age on the intestinal absorption of vitamin D₃-palmitate and nonesterified vitamin D₂ in the term human infant. J Clin Endocrinol Metab 81:1385–1388
- 525. Karlén J, Aperia A, Zetterstrüm R 1985 Renal excretion of calcium and phosphate in preterm and term infants. J Pediatr 106:814–819
- 526. Ghazali S, Barratt TM 1974 Urinary excretion of calcium and magnesium in children. Arch Dis Child 49:97–101
- 527. **Guignard JP, Torrado A, Da Cunha O, Gautier E** 1975 Glomerular filtration rate in the first three weeks of life. J Pediatr 87:268–272
- 528. Linarelli LG 1972 Newborn urinary cyclic AMP and developmental renal responsiveness to parathyroid hormone. Pediatrics 50:14–23
- 529. Mallet É, Basuyau JP, Brunelle P, Devaux AM, Fessard C 1978 Neonatal parathyroid secretion and renal receptor maturation in premature infants. Biol Neonate 33:304–308
- 530. Widdowson EM, McCance RA 1965 The metabolism of phosphorus, calcium, magnesium and strontium. Pediatr Clin North Am 12:595–614
- 531. Ziegler EE, O'Donnell AM, Nelson SE, Fomon SJ 1976 Body composition of the reference fetus. Growth 40:329–341
- 532. Sparks JW 1984 Human intrauterine growth and nutrient accretion. Semin Perinatol 8:74–93
- 533. Cockburn F, Belton NR, Purvis RJ, Giles MM, Brown JK, Turner TL, Wilkinson EM, Forfar JO, Barrie WJ, McKay GS, Pocock SJ 1980 Maternal vitamin D intake and mineral metabolism in mothers and their newborn infants. Br Med J 281:11–14
- 534. Brooke OG, Brown IR, Bone CD, Carter ND, Cleeve HJ, Maxwell JD, Robinson VP, Winder SM 1980 Vitamin D supplements in pregnant Asian women: effects on calcium status and fetal growth. Br Med J 280:751–754
- 535. **Reif S, Katzir Y, Eisenberg Z, Weisman Y** 1988 Serum 25-hydroxyvitamin D levels in congenital craniotabes. Acta Paediatr Scand 77:167–168
- 536. Specker BL, Ho ML, Oestreich A, Yin TA, Shui QM, Chen XC, Tsang RC 1992 Prospective study of vitamin D supplementation and rickets in China. J Pediatr 120:733–739
- 537. Brunvand L, Quigstad E, Urdal P, Haug E 1996 Vitamin D deficiency and fetal growth. Early Hum Dev 45:27–33
- 538. **Garel JM** 1983 Parathyroid hormone, calcitonin and mineral metabolism in the mammalian fetus and neonate. In: Hollick MF, Anast CS, Gray TK (eds) Perinatal Calcium and Phosphorus Metabolism. Excerpta Medica, Amsterdam, pp 71–104
- 539. **Krukowski M, Kahn AJ** 1980 The role of parathyroid hormone in mineral homeostasis and bone modeling in suckling rat pups. Metab Bone Dis Relat Res 2:257–260
- 540. Minton SD, Steichen JJ, Tsang RC 1979 Bone mineral content in term and preterm appropriate-for-gestational-age infants. J Pediatr 95:1037–1042
- 541. Steichen JJ, Asch PA, Tsang RC 1988 Bone mineral content measurement in small infants by single-photon absorptiometry: current methodologic issues. J Pediatr 113:181–187
- 542. Greer FR, McCormick A 1986 Bone growth with low bone mineral content in very low birth weight premature infants. Pediatr Res 20:925–928
- 543. **Greer FR** 1988 Determination of radial bone mineral content in low birth weight infants by photon absorptiometry. J Pediatr 113:213– 219
- 544. Steichen JJ, Gratton TL, Tsang RC 1980 Osteopenia of prematurity: the cause and possible treatment. J Pediatr 96:528–534
- 545. Koo WW, Sherman R, Succop P, Ho M, Buckley D, Tsang RC 1989 Serum vitamin D metabolites in very low birth weight infants with and without rickets and fractures. J Pediatr 114:1017–1022
- 546. McIntosh N, De Curtis M, Williams J 1986 Failure of mineral supplementation to reduce incidence of rickets in very-low-birth-weight infants (letter). Lancet 2:981–982
- 547. Steichen JJ, Tsang RC, Greer FR, Ho M, Hug G 1981 Elevated serum 1,25 dihydroxyvitamin D concentrations in rickets of very low-birth-weight infants. J Pediatr 99:293–298

- 548. **Gradus D, Le Roith D, Karplus M, Zmora E, Grief M, Bar-Ziv J** 1981 Congenital hyperparathyroidism and rickets: secondary to maternal hypoparathyroidism and vitamin D deficiency. Isr J Med Sci 17:705–708
- 549. Key Jr LL, Thorne M, Pitzer B, Volberg F, Turner C 1990 Management of neonatal hyperparathyroidism with parathyroidectomy and autotransplantation. J Pediatr 116:923–926
- 550. Tsang RC, Venkataraman P, Ho M, Steichen JJ, Whitsett J, Greer F 1984 The development of pseudohypoparathyroidism. Involvement of progressively increasing serum parathyroid hormone concentrations, increased 1,25-dihydroxyvitamin D concentrations, and 'migratory' subcutaneous calcifications. Am J Dis Child 138: 654–658
- 551. Minagawa M, Yasuda T, Kobayashi Y, Niimi H 1995 Transient pseudohypoparathyroidism of the neonate. Eur J Endocrinol 133: 151–155
- 552. **Mimouni F, Tsang RC** 1992 Pathophysiology of neonatal hypocalcemia. In: Polin RA, Fox WW (eds) Fetal and Neonatal Physiology. W.B. Saunders, Philadelphia, pp 1761–1767
- 553. Specker BL, Tsang RC, Ho ML, Landi TM, Gratton TL 1991 Low serum calcium and high parathyroid hormone levels in neonates fed 'humanized' cow's milk-based formula. Am J Dis Child 145: 941–945
- 554. Noguchi A, Eren M, Tsang RC 1980 Parathyroid hormone in hypocalcemic and normocalcemic infants of diabetic mothers. J Pediatr 97:112–114
- 555. **Strand CE, Ehrenkranz RA** 1994 Infants of diabetic mothers. In: Lebovitz HE (ed) Therapy for Diabetes Mellitus and Related Disorders. American Diabetes Association, Alexandria, VA, pp 32–37
- 556. **Freinkel N, Ogata E, Metzger BE** 1990 The offspring of the mother with diabetes. In: Rifkin H, Porte Jr D (eds) Diabetes Mellitus: Theory and Practice. Elsevier, New York, pp 651–659
- 557. Demarini S, Mimouni F, Tsang RC, Khoury J, Hertzberg V 1994 Impact of metabolic control of diabetes during pregnancy on neonatal hypocalcemia: a randomized study. Obstet Gynecol 83:918– 922
- 558. Tsang RC, Chen I, Friedman MA, Gigger M, Steichen J, Koffler H, Fenton L, Brown D, Pramanik A, Keenan W, Strub R, Joyce T 1975 Parathyroid function in infants of diabetic mothers. J Pediatr 86:399-404
- 559. Tsang RC, Strub R, Brown DR, Steichen J, Hartman C, Chen IW 1976 Hypomagnesemia in infants of diabetic mothers: perinatal studies. J Pediatr 89:115–119
- 560. Mimouni F, Tsang RC, Hertzberg VS, Miodovnik M 1986 Polycythemia, hypomagnesemia, and hypocalcemia in infants of diabetic mothers. Am J Dis Child 140:798–800
- 561. Martinez ME, Catalan P, Balaguer G, Lisbona A, Quero J, Reque A, Pallardo LF 1991 25(OH)D levels in diabetic pregnancies relation with neonatal hypocalcemia. Horm Metab Res 23:38–41
- 562. Salle B, David L, Glorieux F, Delvin EE, Louis JJ, Troncy G 1982 Hypocalcemia in infants of diabetic mothers. Studies in circulating calciotropic hormone concentrations. Acta Paediatr Scand 71:573– 577
- 563. Donatelli M, Bucalo ML, Russo V, Cerasola GA 1984 Calcium hormones in diabetic pregnancy. Boll Soc Ital Biol Sper 60:1503–1508
- 564. Sgambato S, Passariello N, Buoninconti R, Caserta R, Paolisso G 1986 Plasma calcitonin variations in normal women and in women with family history of diabetes during pregnancy. Diabete Metab 12:297–301
- 565. Mimouni F, Tsang RC, Hertzberg VS, Neumann V, Ellis K 1989 Parathyroid hormone and calcitriol changes in normal and insulindependent diabetic pregnancies. Obstet Gynecol 74:49–54
- 566. Birdsey TJ, Husain SM, Garland HO, Sibley CP 1995 The effect of diabetes mellitus on urinary calcium excretion in pregnant rats and their offspring. J Endocrinol 145:11–18
- 567. Husain SM, Birdsey TJ, Glazier JD, Mughal MZ, Garland HO, Sibley CP 1994 Effect of diabetes mellitus on maternofetal flux of calcium and magnesium and calbindin_{9K} mRNA expression in rat placenta. Pediatr Res 35:376–381